

VOLUME

6

Regis Technologies

**Chiral
Application
Guide**



DURABLE, HIGH EFFICIENCY CHIRAL COLUMNS

Since 1980, Regis Technologies, Inc. has proven to be a leader in chiral separations and services. Our support system to the analytical and preparative chromatographer is a model other manufacturers of chiral products try to emulate. Our technical expertise, professional staff, and worldwide distribution network is highly respected and praised in the chiral community.

Regis offers four different types of Chiral Stationary Phases (CSP's):

- Covalently bonded Pirkle-Type Concept
- Protein-based
- Covalently bonded 18 Crown-ether

The complete line of Pirkle-Type CSP's are manufactured on-site at our cGMP facility in Morton Grove, Illinois. Columns range from analytical to preparative in size. Since Regis packs their own columns, custom sized columns are easily attainable.

Regis maintains an extended inventory of Protein-based columns manufactured by **ChromTech** in the United Kingdom and the covalently bonded 18 Crown-Ether columns manufactured by **RStech** in South Korea. Information on these product lines is readily available on our website at www.registech.com.

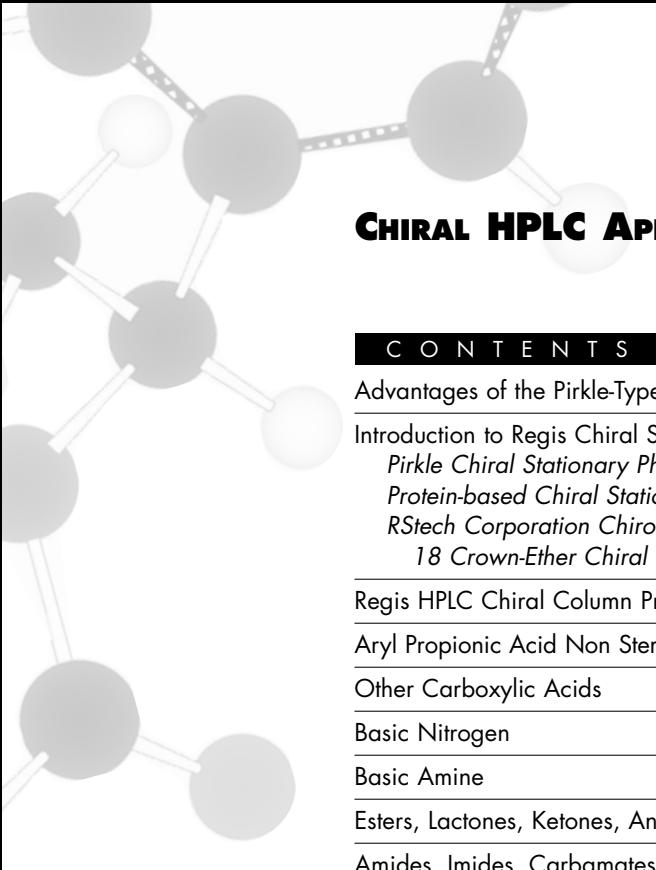
As evidence of our commitment to the scientist in the lab, Regis is pleased to present its **Chiral Application Guide VI**. This new guide contains over 500 specific chiral applications using a variety of chiral column types. We have also included a few new sections in this guide. We added a **Frequently Asked Question** section and a **Quick Scheme Method Development** section.

For applications not listed in this guide, Regis maintains a dedicated chiral separations lab and chiral separations scientist. This enables Regis to offer a **Free Chiral Screening Service** to the scientific community. For specific column types, services or applications not listed in this guide, please contact Regis directly or visit our website for continuously updated information.



All Regis Chiral Separations products must meet rigorous manufacturing and quality control specifications before release.

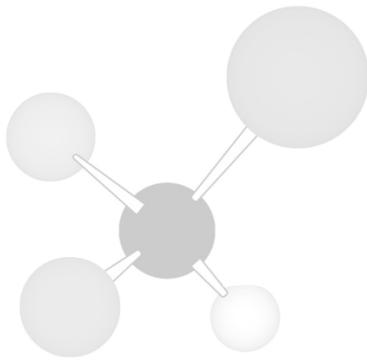
Regis Technologies, Inc.



CHIRAL HPLC APPLICATION GUIDE VI

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Advantages of the Pirkle-Type Chiral Stationary Phases

Universal Solvent Capability

The entire family of Regis' Pirkle-Type Chiral Stationary Phases (CSP's) can be used in **BOTH** normal and reversed-phase solvents. Due to the fact that all of the Pirkle-Type CSP's are covalently bonded, the columns can tolerate all commonly used mobile phase combinations.

Column Durability

Another advantage of covalent bonding is column durability. Listed below are a few distinct benefits associated with the Pirkle-Type CSP's:

- Long lasting columns
- Bonded selector will not leach off the silica gel
- Can tolerate sample overload
- Can utilize strong solvents for cleaning
- Columns are fully reversible
- Compatible in SFC and SMB applications

Ability to Invert Elution Order

All of the Pirkle-Type CSP's are available in both enantiomeric forms. This allows the Chromatographer to invert the elution order of the enantiomers by simply switching columns. This advantage is essential when determining enantiomeric purity when the trace enantiomer should elute before the major. Elution order is also important in preparative chromatography because when the desired enantiomer elutes first, purity and production efficiency increases.

Chromatographic Efficiency

Unlike most Chiral columns on the market, Pirkle-Type Chiral HPLC columns show excellent chromatographic efficiency. The high density of binding sites allows larger amounts of sample to be injected without major changes in column performance.

Ease of Scale-up

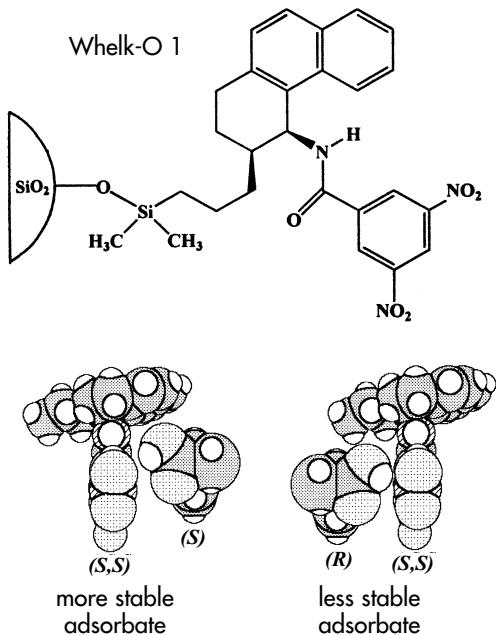
Pirkle-Type CSP's were designed to allow the Chromatographer to scale-up their separation from analytical to preparative in a linear fashion. Regis uses the highest grade silica gels available on the market today. The 5-micron CSP's are bonded on Exsil®. Our 10-micron and 16-micron CSP's are bonded on Kromasil®. Synthesis of the chiral selectors, bonding of the different CSP's, and column production is all performed by Regis in one facility. This allows Regis total control over the product line. This also allows Regis to perform special requests for the customer, including custom bonding and custom column packing.

Pirkle Chiral Stationary Phases

The Pirkle-Concept Chiral Stationary Phases generally fall into three classes: π -electron acceptor/ π -electron donors, the π -electron acceptors and the π -electron donors. With Pirkle Phases, chiral recognition occurs at binding sites. Major binding sites are classified as π -basic or π -acidic aromatic rings, acidic sites, basic sites, and steric interaction sites. Aromatic rings are potential sites for π - π interactions. Acidic sites supply hydrogens for potential intermolecular hydrogen bonds—the hydrogen is often an amido proton (N-H) from an amide, carbamate, urea, or amine. Basic sites, such as π -electrons, sulfinyl or phosphinyl oxygens, and hydroxy or ether oxygens, may also be involved in hydrogen bond formation. Steric interactions may also occur between large groups.

π -Electron Acceptor/ π -Electron Donor Phases

- WHELK-O 1
- WHELK-O 2
- ULMO

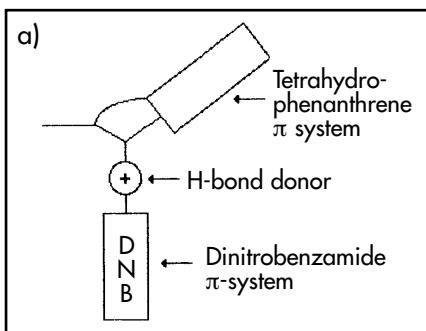


The latest and most revolutionary addition to the Pirkle-Concept series is the π -electron acceptor/ π -electron donor phase. This concept is an innovative incorporation of both π -acceptor and π -donor characteristics, resulting in a phase that can be used for a wide variety of compound groups.

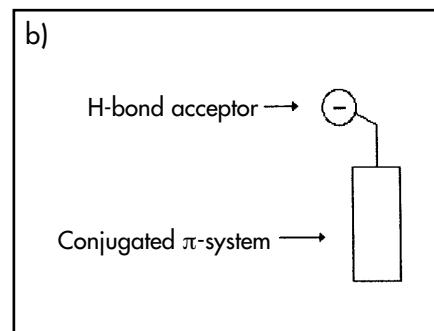
WHELK-O 1

The Whelk-O 1 Chiral Stationary Phase is based on 1-(3,5-Dinitrobenzamido)-1,2,3,4-tetrahydrophenanthrene. This phase allows separation of underderivatized racemates from a number of families including amides, epoxides, esters, ureas, carbamates, ethers, aziridines, phosphonates, aldehydes, ketones, carboxylic acids, and alcohols.

The Whelk-O 1 was originally designed for the separation of underderivatized non-steroidal anti-inflammatory drugs (NSAIDs). This π -electron acceptor/ π -electron donor phase exhibits an extraordinary degree of generality, allowing resolution of a wide variety of underderivatized racemates. This broad versatility observed on the Whelk-O 1 column, compares favorably with polysaccharide-derived chiral stationary phases. In addition, because of the Whelk-O 1's covalent nature, this chiral phase is compatible with all commonly used mobile phases, including aqueous systems—a distinct advantage over polysaccharide-derived chiral stationary phases. Other advantages include column durability, excellent efficiency, elution order inversion allowing availability of both enantiomeric forms, and excellent preparative capacity.



a) Schematic diagram showing key functional groups of the Whelk-O 1 involved in chiral recognition.



b) Schematic diagram showing generalized structure of analytes which are resolved on the Whelk-O 1.

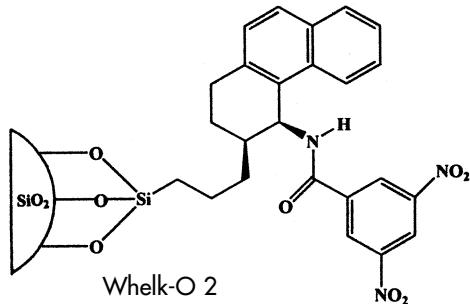
Introduction to Regis Chiral Stationary Phases

REGIS

WHELK-O 2

The Whelk-O 2 is the covalent trifunctional version of the Whelk-O 1. The Whelk-O 2 retains the same chiral selector but modifies the support to silica from a monofunctional linkage to a trifunctional. In most cases, the enantioselectivity remains the same allowing the separation of the analogous family of racemates as does the Whelk-O 1.

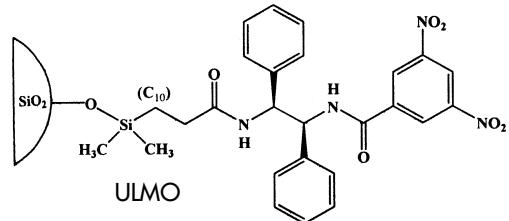
Whelk-O 2 was designed to enhance the stability of the stationary phase due to hydrolysis while using strong organic modifiers such as trifluoroacetic acid. The Whelk-O 2 is ideal for preparative separations since the material is bonded on 10 µm 100 Å spherical Kromasil silica. This allows the preparative chromatographer to perform method development on their analytical column and immediately scale-up to larger diameter columns.



ULMO

The ULMO chiral stationary phase was developed by Austrian Researchers, Uray, Lindner, and Maier. This CSP has a general ability to separate the enantiomers of many racemate classes, and is particularly good at separating the enantiomers of aryl carbinols.

The ULMO CSP is based on a 3,5-Dinitrobenzoyl derivative of diphenylethylenediamine.



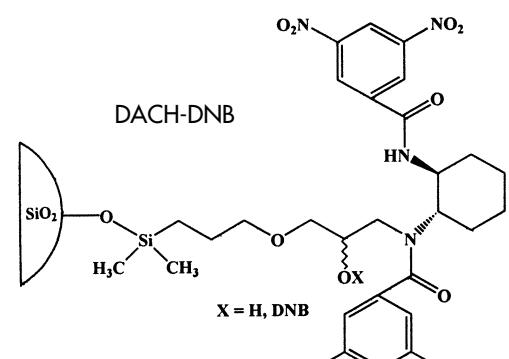
π-Electron Acceptor Phases

- DACH-DNB
- Pirkle 1-J
- α-Burke 2
- β-Gem 1
- Leucine
- Phenylglycine

The π-electron acceptor Pirkle Chiral Stationary Phases can be used to separate a wide range of enantiomers without derivatization, as demonstrated for the following classes of solutes: secondary benzyl alcohols, mandelic acid analogs, α-hydroxy-α-aryl phosphates, α-tetralol analogs, propranolol analogs, β-hydroxy-aryl sulfoxides, alkyl-aryl sulfoxides, diaryl sulfoxides, aryl-substituted cyclic phthalides, aryl-substituted lactams, aryl-substituted succinimides, aryl-substituted hydantoins, bi-β-naphthol and its analogs, and α-aryl acetamides.

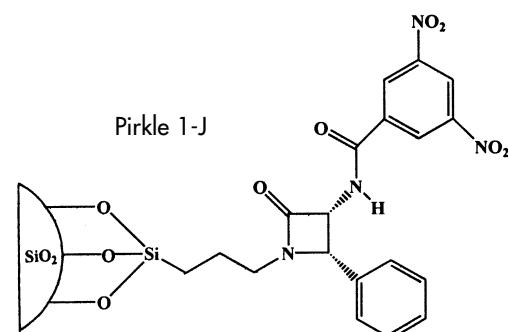
DACH-DNB

The innovative DACH-DNB CSP was designed by Italian chemists Dr. Francesco Gasparrini, Misiti and Villani at Rome University "La Sapienza." The DACH-DNB CSP; which contains the 3,5-dinitrobenzoyl derivative of 1,2-diaminocyclohexane, has been found to resolve a broad range of racemate classes including amides, alcohols, esters, ketones, acids, sulfoxides, phosphine oxides, selenoxides, phosphonates, thiophosphineoxides, phosphineselenides, phosphine-boranes, β-lactams, organometallics, atropisomers and heterocycles.

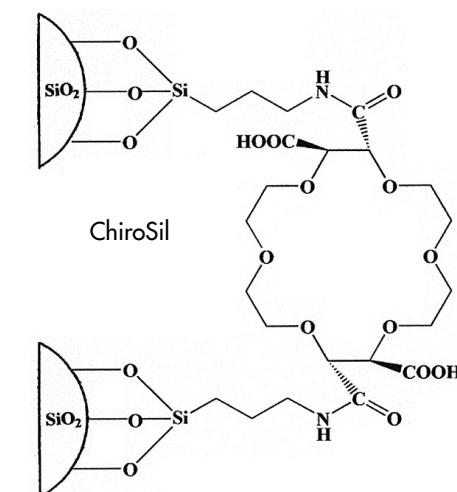
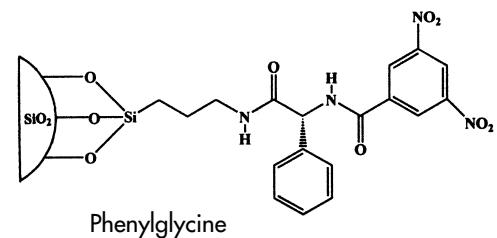
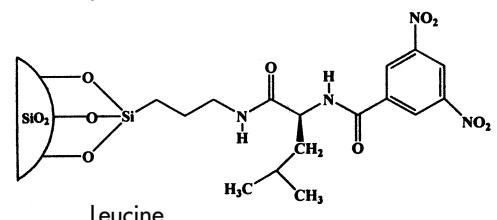
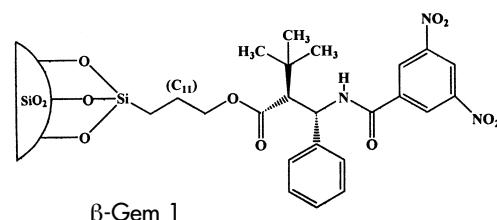
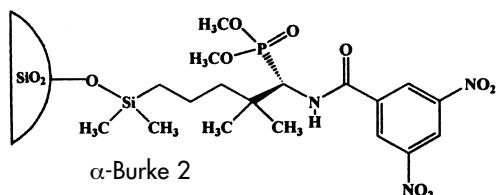


PIRKLE 1-J

The Pirkle 1-J CSP is based on 3-(3,5-Dinitrobenzamido)-4-phenyl-β-lactam. This unusual β-lactam structure significantly alters its molecular recognition properties. The Pirkle 1J is useful for the direct separation of underderivatized β-blocker enantiomers. It can also be used for the separation of the enantiomers of arylpropionic acid NSAID's as well as other drugs.



REGIS Introduction to Regis Chiral Stationary Phases



α -BURKE 2

The α -Burke 2 phase, first prepared by J. A. Burke III, a graduate student of Dr. Pirkle, is derived from dimethyl N-3,5-dinitro-benzoyl- α -amino-2,2-dimethyl-4-pentenyl phosphonate. The α -Burke 2 has been specifically designed to directly separate the enantiomers of β -blockers without chemical derivatization, but this chiral phase is not limited solely to the separation of β -blocker enantiomers. It also resolves the enantiomers of many compounds separated on π -acceptor Pirkle type chiral stationary phases.

β -GEM 1

β -Gem 1 is a π -acceptor chiral stationary phase and is derived from N-3,5-dinitrobenzoyl-3-amino-3-phenyl-2-(1,1-dimethylethyl)-propanoate.

For a great many analytes, this chiral phase considerably outperforms its widely used analog, phenylglycine. It can separate anilide derivatives of a wide variety of chiral carboxylic acids, including nonsteroidal anti-inflammatory agents.

LEUCINE

The leucine CSP is based on the 3,5-dinitrobenzoyl derivative of leucine. This π -acceptor phase demonstrates enhanced enantioselectivities for several classes of compounds, including benzodiazepines.

PHENYLGLYCINE

Phenylglycine is based on the 3,5-dinitrobenzoyl derivative of phenylglycine.

This CSP resolves a wide variety of compounds which contain π -basic groups. These include: aryl-substituted cyclic sulfoxides, bi- β -naphthol and its analogs, α -indanol and α -tetralol analogs, and aryl-substituted hydantoins.

Protein-Based Chiral Stationary Phases

Regis Technologies carries a line of protein-based chiral columns manufactured by ChromTech AB, United Kingdom. For additional product information and a Protein-Based Chiral Stationary Phase application guide, please contact Regis directly.

- Chiral AGP (α -glycoprotein)
- Chiral CBH (cellobiohydrolase)
- Chiral HSA (human serum albumin)

RStech Corporation ChiroSil® RCA(+) and SCA(-) 18Crown-Ether Chiral Stationary Phases

- ChiroSil RCA(+)
- ChiroSil SCA (-)

Developed by RStech Corporation in Daejeon, Korea, the ChiroSil phase is the newest addition to our chiral line of columns. This phase is prepared by a covalent trifunctional bonding of (+) or (-)-(18-Crown-6)-tetracarboxylic acid as the chiral selector.

This phase which is available in analytical as well as preparative columns, is an excellent choice for the separation of amino acids and compounds containing primary amines.

Like our other line of columns, this phase is highly durable, has universal solvent compatibility, and has the ability to invert elution order.

As described above, Regis' Chiral columns can be used to separate a wide variety of enantiomers in numerous compound groups. Please refer to the Product List on page 7 for particular column types, sizes, configurations and product numbers. Consult the application separation data section that begins on page 8 for information regarding specific chiral separations on a wide variety of compounds. See Application Indexes on page 87.

Regis Chiral Column Product List

REGIS

PRODUCT	PARTICLE SIZE	COLUMN DIMENSIONS	PRODUCT#	PRODUCT	PARTICLE SIZE	COLUMN DIMENSIONS	PRODUCT#
(R,R)-Whelk-O 1	5 µm, 100 Å	25 cm x 4.6 mm	786201	(R,R)-DACH-DNB	5 µm, 100 Å	25 cm x 4.6 mm	788101
(R,R)-Whelk-O 1	5 µm, 100 Å	25 cm x 10.0 mm	786202	(R,R)-DACH-DNB	5 µm, 100 Å	25 cm x 10.0 mm	788102
(R,R)-Whelk-O 1	5 µm, 100 Å	25 cm x 30.0 mm	786205	(R,R)-DACH-DNB	5 µm, 100 Å	25 cm x 30.0 mm	788104
(S,S)-Whelk-O 1	5 µm, 100 Å	25 cm x 4.6 mm	786101	(S,S)-DACH-DNB	5 µm, 100 Å	25 cm x 4.6 mm	788201
(S,S)-Whelk-O 1	5 µm, 100 Å	25 cm x 10.0 mm	786102	(S,S)-DACH-DNB	5 µm, 100 Å	25 cm x 10.0 mm	788202
(S,S)-Whelk-O 1	5 µm, 100 Å	25 cm x 30.0 mm	786105	(S,S)-DACH-DNB	5 µm, 100 Å	25 cm x 30.0 mm	788204
(R,R)-Whelk-O 1	10 µm, 100 Å	25 cm x 4.6 mm	786515	(R,R)-DACH-DNB	10 µm, 100 Å	25 cm x 21.1 mm	788103
(R,R)-Whelk-O 1	10 µm, 100 Å	25 cm x 10.0 mm	786525	(R,R)-DACH-DNB	10 µm, 100 Å	25 cm x 30.0 mm	788707
(R,R)-Whelk-O 1	10 µm, 100 Å	25 cm x 21.1 mm	786535	(R,R)-DACH-DNB	10 µm, 100 Å	25 cm x 50.0 mm	788708
(R,R)-Whelk-O 1	10 µm, 100 Å	50 cm x 21.1 mm	786545	(R,R)-DACH-DNB	10 µm, 100 Å	50 cm x 30.0 mm	788712
(R,R)-Whelk-O 1	10 µm, 100 Å	25 cm x 30.0 mm	786708	(R,R)-DACH-DNB	10 µm, 100 Å	50 cm x 50.0 mm	788709
(R,R)-Whelk-O 1	10 µm, 100 Å	25 cm x 50.0 mm	786709	(S,S)-DACH-DNB	10 µm, 100 Å	25 cm x 21.1 mm	788203
(R,R)-Whelk-O 1	10 µm, 100 Å	50 cm x 30.0 mm	786713	(S,S)-DACH-DNB	10 µm, 100 Å	25 cm x 30.0 mm	788701
(R,R)-Whelk-O 1	10 µm, 100 Å	50 cm x 50.0 mm	786710	(S,S)-DACH-DNB	10 µm, 100 Å	25 cm x 50.0 mm	788702
(S,S)-Whelk-O 1	10 µm, 100 Å	25 cm x 4.6 mm	786615	(S,S)-DACH-DNB	10 µm, 100 Å	50 cm x 30.0 mm	788715
(S,S)-Whelk-O 1	10 µm, 100 Å	25 cm x 10.0 mm	786625	(S,S)-DACH-DNB	10 µm, 100 Å	50 cm x 50.0 mm	788705
(S,S)-Whelk-O 1	10 µm, 100 Å	25 cm x 21.1 mm	786635				
(S,S)-Whelk-O 1	10 µm, 100 Å	50 cm x 21.1 mm	786645	(3R,4S)-Pirkle 1-J	5 µm, 100 Å	25 cm x 4.6 mm	731044
(S,S)-Whelk-O 1	10 µm, 100 Å	25 cm x 30.0 mm	786702	(3R,4S)-Pirkle 1-J	5 µm, 100 Å	25 cm x 10.0 mm	731244
(S,S)-Whelk-O 1	10 µm, 100 Å	25 cm x 50.0 mm	786703	(3S,4R)-Pirkle 1-J	5 µm, 100 Å	25 cm x 4.6 mm	731045
(S,S)-Whelk-O 1	10 µm, 100 Å	50 cm x 30.0 mm	786716	(3S,4R)-Pirkle 1-J	5 µm, 100 Å	25 cm x 10.0 mm	731245
(S,S)-Whelk-O 1	10 µm, 100 Å	50 cm x 50.0 mm	786704				
(R,R)-Whelk-O 2	10 µm, 100 Å	25 cm x 4.6 mm	786315	(R,R)- α -Burke 2	5 µm, 100 Å	25 cm x 4.6 mm	735035
(R,R)-Whelk-O 2	10 µm, 100 Å	25 cm x 10.0 mm	786325	(R,R)- α -Burke 2	5 µm, 100 Å	25 cm x 10.0 mm	735235
(R,R)-Whelk-O 2	10 µm, 100 Å	25 cm x 21.1 mm	786335	(S,S)- α -Burke 2	5 µm, 100 Å	25 cm x 4.6 mm	735037
(R,R)-Whelk-O 2	10 µm, 100 Å	50 cm x 21.1 mm	786345	(S,S)- α -Burke 2	5 µm, 100 Å	25 cm x 10.0 mm	735237
(R,R)-Whelk-O 2	10 µm, 100 Å	25 cm x 30.0 mm	786727				
(R,R)-Whelk-O 2	10 µm, 100 Å	25 cm x 50.0 mm	786728	(R,R)- β -Gem 1	5 µm, 100 Å	25 cm x 4.6 mm	731043
(R,R)-Whelk-O 2	10 µm, 100 Å	50 cm x 30.0 mm	786732	(R,R)- β -Gem 1	5 µm, 100 Å	25 cm x 10.0 mm	731243
(R,R)-Whelk-O 2	10 µm, 100 Å	50 cm x 50.0 mm	786729	(S,S)- β -Gem 1	5 µm, 100 Å	25 cm x 4.6 mm	731029
(S,S)-Whelk-O 2	10 µm, 100 Å	25 cm x 4.6 mm	786415	(S,S)- β -Gem 1	5 µm, 100 Å	25 cm x 10.0 mm	731229
(S,S)-Whelk-O 2	10 µm, 100 Å	25 cm x 10.0 mm	786425				
(S,S)-Whelk-O 2	10 µm, 100 Å	25 cm x 21.1 mm	786435	D-Leucine	5 µm, 100 Å	25 cm x 4.6 mm	731054
(S,S)-Whelk-O 2	10 µm, 100 Å	50 cm x 21.1 mm	786445	D-Leucine	5 µm, 100 Å	25 cm x 10.0 mm	731254
(S,S)-Whelk-O 2	10 µm, 100 Å	25 cm x 30.0 mm	786721	L-Leucine	5 µm, 100 Å	25 cm x 4.6 mm	731041
(S,S)-Whelk-O 2	10 µm, 100 Å	25 cm x 50.0 mm	786722	L-Leucine	5 µm, 100 Å	25 cm x 10.0 mm	731241
(S,S)-Whelk-O 2	10 µm, 100 Å	50 cm x 30.0 mm	786736				
(S,S)-Whelk-O 2	10 µm, 100 Å	50 cm x 50.0 mm	786723	D-Phenylglycine	5 µm, 100 Å	25 cm x 4.6 mm	731021
(R,R)-ULMO	5 µm, 100 Å	25 cm x 4.6 mm	787200	D-Phenylglycine	5 µm, 100 Å	25 cm x 10.0 mm	731221
(R,R)-ULMO	5 µm, 100 Å	25 cm x 10.0 mm	787201	L-Phenylglycine	5 µm, 100 Å	25 cm x 4.6 mm	731024
(R,R)-ULMO	5 µm, 100 Å	25 cm x 30.0 mm	787203	L-Phenylglycine	5 µm, 100 Å	25 cm x 10.0 mm	731224
(S,S)-ULMO	5 µm, 100 Å	25 cm x 4.6 mm	787100				
(S,S)-ULMO	5 µm, 100 Å	25 cm x 10.0 mm	787101	Chiral AGP	5 µm, 300 Å	10 cm x 4.0 mm	732200
(S,S)-ULMO	5 µm, 100 Å	25 cm x 30.0 mm	787103	Chiral AGP	5 µm, 300 Å	15 cm x 4.0 mm	732199
(R,R)-ULMO	10 µm, 100 Å	25 cm x 21.1 mm	787202	Chiral CBH	5 µm, 300 Å	10 cm x 4.0 mm	732350
(R,R)-ULMO	10 µm, 100 Å	25 cm x 30.0 mm	787707	Chiral CBH	5 µm, 300 Å	15 cm x 4.0 mm	732351
(R,R)-ULMO	10 µm, 100 Å	25 cm x 50.0 mm	787708	Chiral HSA	5 µm, 300 Å	10 cm x 4.0 mm	732240
(R,R)-ULMO	10 µm, 100 Å	50 cm x 30.0 mm	787712	Chiral HSA	5 µm, 300 Å	15 cm x 4.0 mm	732239
(R,R)-ULMO	10 µm, 100 Å	50 cm x 50.0 mm	787709				
(S,S)-ULMO	10 µm, 100 Å	25 cm x 21.1 mm	787102	ChiroSil® RCA(+)	5 µm, 100 Å	15 cm x 4.6 mm	799001
(S,S)-ULMO	10 µm, 100 Å	25 cm x 30.0 mm	787701	ChiroSil® RCA(+)	5 µm, 100 Å	25 cm x 4.6 mm	799002
(S,S)-ULMO	10 µm, 100 Å	25 cm x 50.0 mm	787702	ChiroSil® SCA(-)	5 µm, 100 Å	15 cm x 4.6 mm	799101
(S,S)-ULMO	10 µm, 100 Å	50 cm x 30.0 mm	787715	ChiroSil® SCA(-)	5 µm, 100 Å	25 cm x 4.6 mm	799102
(S,S)-ULMO	10 µm, 100 Å	50 cm x 50.0 mm	787703				

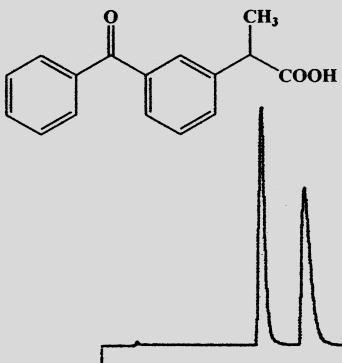
Bulk material is available for all Chiral Stationary Phases

For column dimensions not listed, please contact Regis

NOTE: All columns (except the protein-based columns) listed contain chiral stationary phases that are covalently bound on 5 µm or 10 µm 100 Å spherical silica. A large variety of column dimensions and/or particle sizes are available upon request.

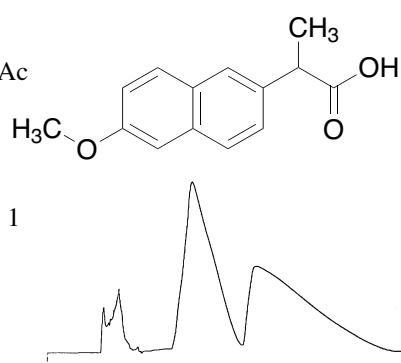
Ketoprofen

Ketoprofen
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (47/47/6)
 $\text{CH}_2\text{Cl}_2/\text{Hexane/Ethanol} + 0.01 \text{ M Ammonium Acetate}$
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 11.0 min
 $k'_1 = 3.63$
 $\alpha = 1.35$
 reference 46

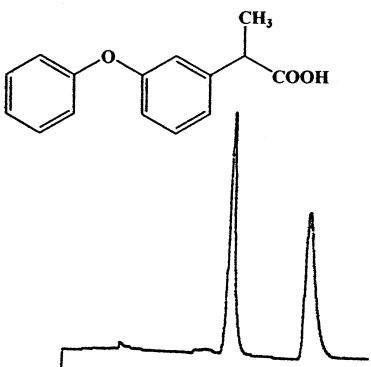
**Naproxen (semi prep)**

Naproxen (semi prep on analytical column)
 80:20:0.5 hexane/IPA/HOAc
 1 ml/min; 300 nm
 Run Time = 18 min
 inject 400 μl @

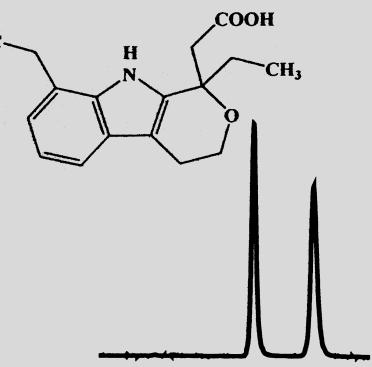
31.5 mg/ml = 12.6 mg
 4.6 mm x 25 cm Whelk-O 1
 reference 6

**Fenoprofen**

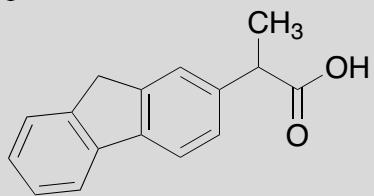
Fenoprofen
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 $\text{Hexane/IPA} + 0.1\% \text{ Acetic Acid}$
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 2.62$
 $\alpha = 1.66$
 reference 46

**Etodolac**

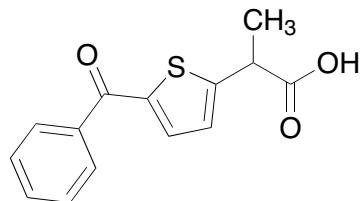
Etodolac
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 $\text{Hexane/IPA} + 0.1\% \text{ TFA}$
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 2.43$
 $\alpha = 1.50$
 reference 48

**Cicloprofen**

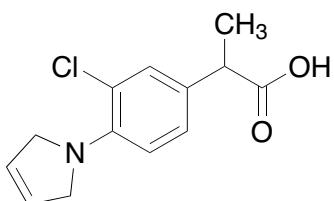
Cicloprofen
 20% IPA/hex, 1g/L NH_4OAc
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.16$
 $\alpha = 2.15$
 reference 4

**Tiaprofenic Acid**

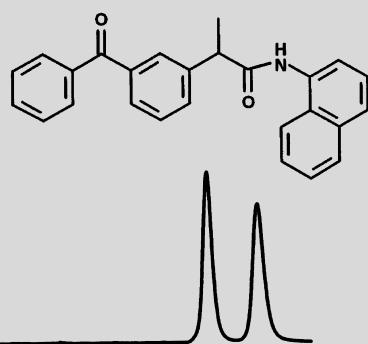
Tiaprofenic Acid
 20% IPA/hex, 1g/L NH_4OAc
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.02$
 $\alpha = 1.09$
 reference 4

**Pirprofen**

Pirprofen
 20% IPA/hex, 1g/L NH_4OAc
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.85$
 $\alpha = 1.81$
 reference 4

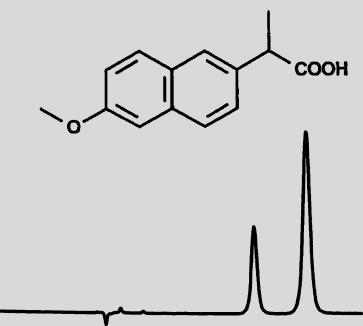
**Ketoprofen as 1-naphthylamide**

Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (70/30)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 230 nm
 Run Time = 13 min
 $k'_1 = 1.51$
 $\alpha = 1.25$
 reference 48

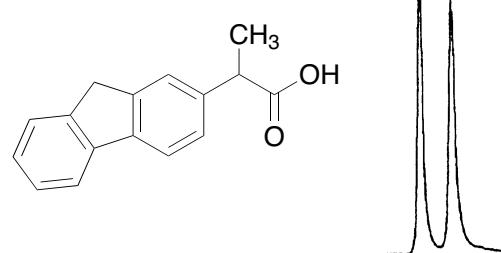


Naproxen (R:S=30:70)

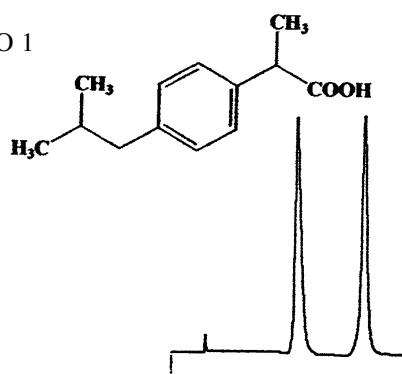
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase (90/10)
Heptane/IPA + 0.1% TFA
Flow Rate = 1.0 mL/min
Detection = UV 230 nm
Run Time = 8.5 min
 $k'_1 = 1.54$
 $\alpha = 1.34$
reference 48

**Cicloprofen**

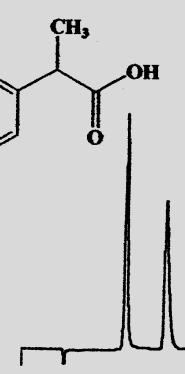
Cicloprofen
70:30:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.48$
 $\alpha = 1.35$
reference 26

**Ibuprofen**

Ibuprofen
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA +
0.01 M Ammonium
Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 11.8 min
 $k'_1 = 3.21$
 $\alpha = 1.72$
reference 46

**Naproxen (Reversed Phase)**

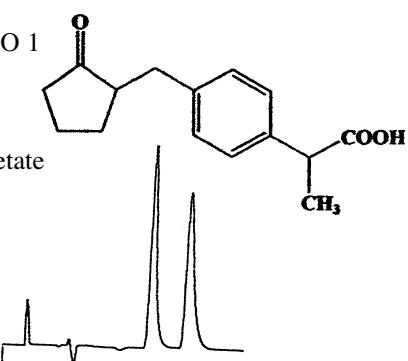
Naproxen (Reversed Phase)
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (80/20)
CH₃OH/H₂O +
0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 10.0 min
 $k'_1 = 1.63$
 $\alpha = 1.64$
reference 46

**Naproxen (Normal Phase)**

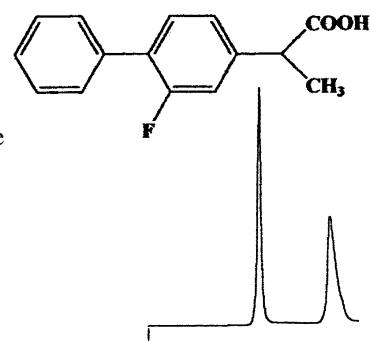
Naproxen (Normal Phase)
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (60/40)
Hexane/IPA +
0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 10.5 min
 $k'_1 = 1.40$
 $\alpha = 2.03$
reference 46

Loxoprofen

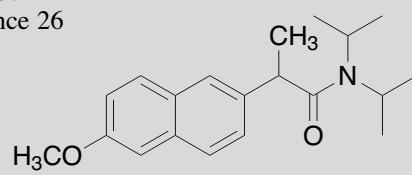
Loxoprofen
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (85/15)
Hexane/Ethanol +
0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 5.41$
 $\alpha = 1.30$
reference 46

**Flurbiprofen**

Flurbiprofen
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA +
0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 20.5 min
 $k'_1 = 5.90$
 $\alpha = 1.76$
reference 46

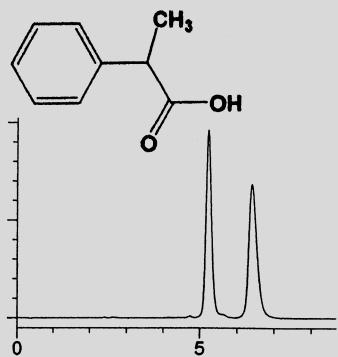
**Naproxen Diisopropyl Amide**

Naproxen Diisopropyl Amide
10%EtOH/hex
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.23$
 $\alpha = 1.53$
reference 26

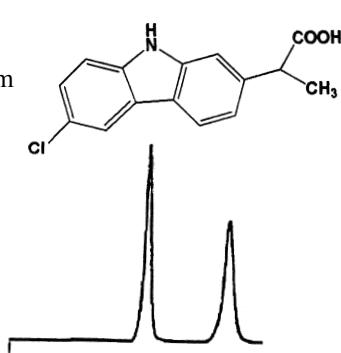


Hydratropic Acid

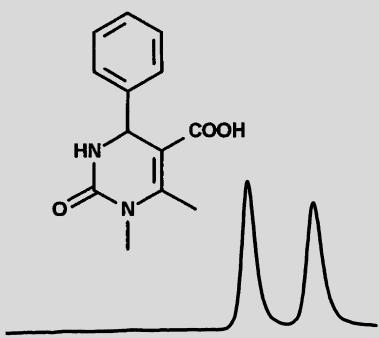
Hydratropic Acid
 Column = (R,R)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase: (98/2)
 Hexane/IPA
 + 0.1% Acetic Acid
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 1.89$
 $\alpha = 1.34$
 reference 46

**Carprofen**

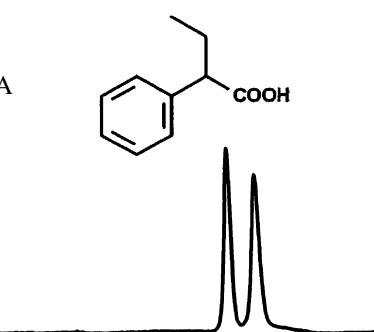
Carprofen
 Column = (R,R)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase: (85/15)
 Hexane/IPA
 + 0.1% Acetic Acid
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 4.70$
 $\alpha = 1.73$
 reference 46

**Tetrahydropyrimidine Carboxylic Acid**

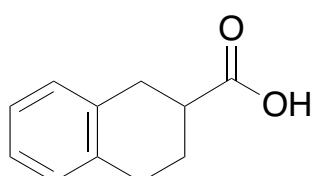
Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 Heptane/IPA + 0.1% TFA
 Flow Rate: 1.0 mL/min
 Detection: UV 215 nm
 Run Time: 14 min
 $k'_1 = 3.38$
 $\alpha = 1.21$
 reference 48

**Other Carboxylic Acids****Phenylbutyric acid**

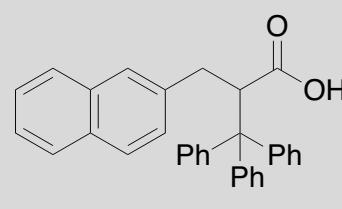
Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Heptane/IPA + 0.1% TFA
 Flow Rate = 2.0 mL/min
 Detection = UV 215 nm
 Run Time = 6.5 min
 $k'_1 = 3.19$
 $\alpha = 1.16$
 reference 48



99:1:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 17 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.06$
 $\alpha = 1.28$
 reference 18



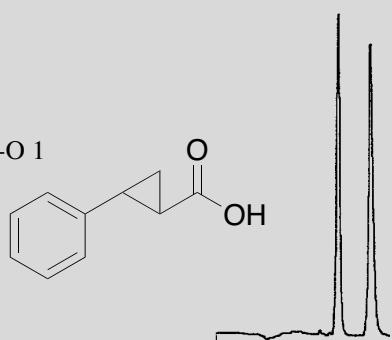
99:1:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 16 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.45$
 $\alpha = 1.38$
 reference 18



REGIS Other Carboxylic Acids

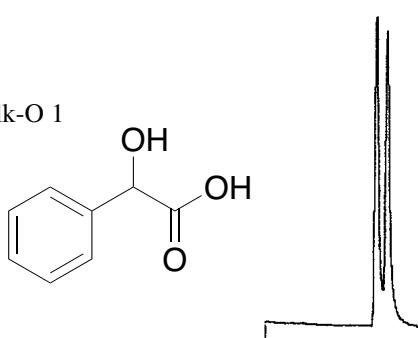
2-Phenylcyclopropane Carboxylate

2-Phenylcyclopropane
Carboxylate
99:1 hexane/IPA
1 ml/min; 220 nm
Run Time = 18 min
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.19$
 $\alpha = 1.34$
reference 18



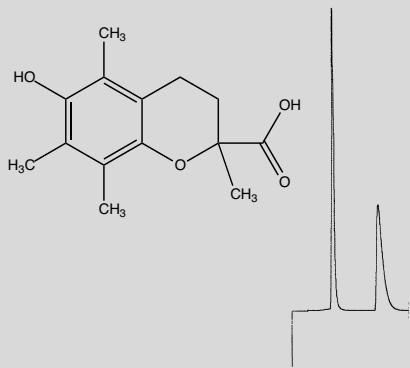
Mandelic Acid

Mandelic Acid
0.1% HOAc in water
1 ml/min; 254 nm
Run Time = 13 min
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.08$
 $\alpha = 1.13$
reference 18

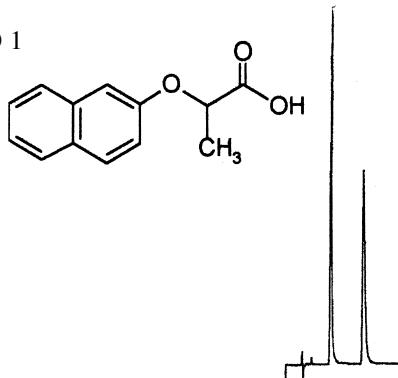


Trolox

Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA +
0.1% Acetic acid
Flow Rate = 1.5 mL/min
Detection = UV 280 nm
Run Time = 12.5 min
 $k'_1 = 2.18$
 $\alpha = 2.68$
reference 46

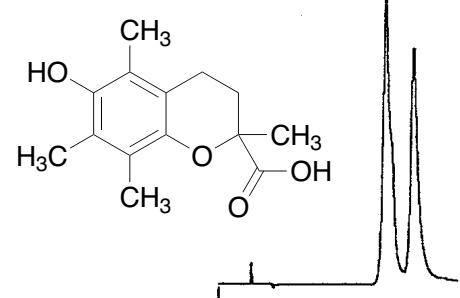


Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 8.5 min
 $k'_1 = 2.03$
 $\alpha = 2.10$
reference 49



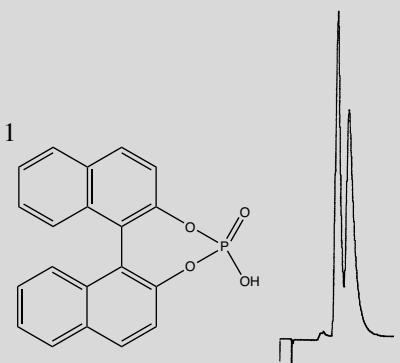
Trolox

Trolox
95:5:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 19 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 5.09$
 $\alpha = 1.21$
reference 18



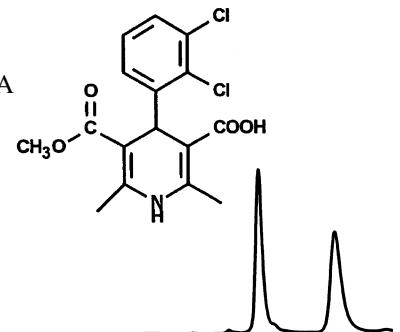
1,1'-binaphthyl-2,2'-diylhydrogen phosphate

56:44 H₂O/MeOH, 0.1%
HOAc
1 ml/min; 254 nm
Run Time = 18 min
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.46$
 $\alpha = 1.27$

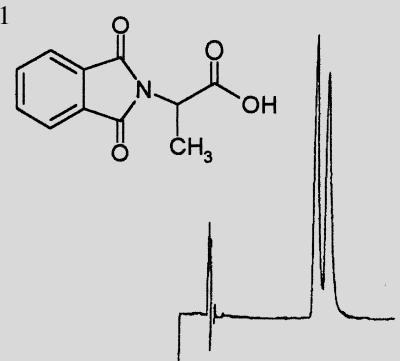


Calcium Channel Blocker

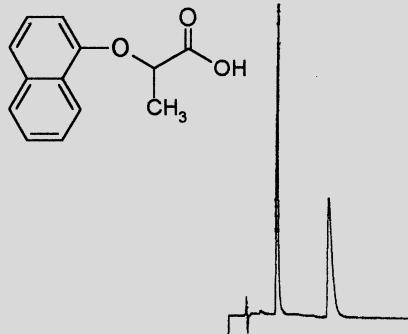
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA + 0.1% TFA
Flow Rate = 1.0 mL/min
Detection = UV 230 nm
Run Time = 6 min
 $k'_1 = 0.55$
 $\alpha = 2.06$
reference 48



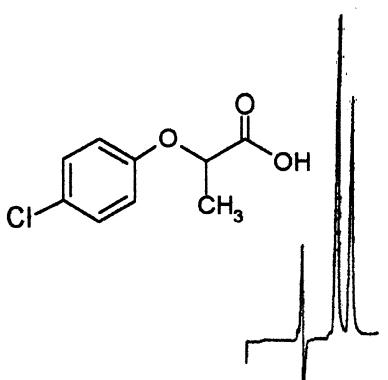
Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 8.5 min
 $k'_1 = 4.20$
 $\alpha = 1.11$
reference 50



Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA + 0.1%
 Trifluoroacetic Acid
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.0 min
 $k'_1 = 2.07$
 $\alpha = 2.62$
 reference 49

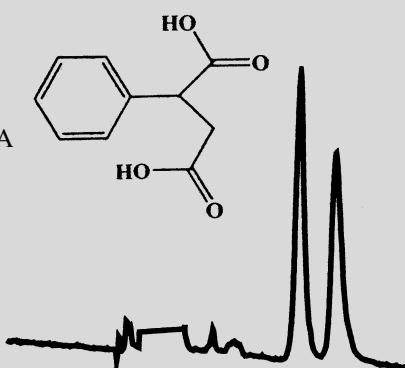


Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase: (95/5)
 Hexane/IPA + 0.1%
 Trifluoroacetic Acid
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 3.5 min
 $k'_1 = 0.84$
 $\alpha = 1.36$
 reference 49



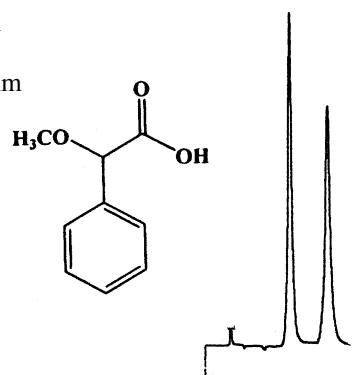
Phenylsuccinic Acid

Phenylsuccinic Acid
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA + 0.1% TFA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 8.5 min
 $k'_1 = 1.71$
 $\alpha = 1.22$
 reference 48

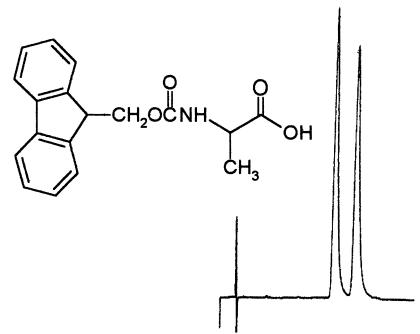


α -Methoxyphenyl Acetic Acid

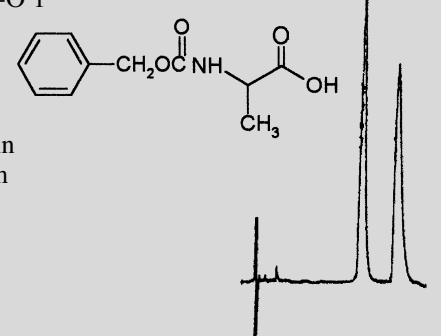
α -Methoxyphenyl Acetic Acid
 Column = (S,S)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (90/10)
 Hexane/Ethanol +
 0.01 M Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 220 nm
 Run Time = 10.0 min
 $k'_1 = 2.96$
 $\alpha = 1.61$
 reference 46



Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA + 0.1%
 Trifluoroacetic Acid
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 7.24$
 $\alpha = 1.22$
 reference 50

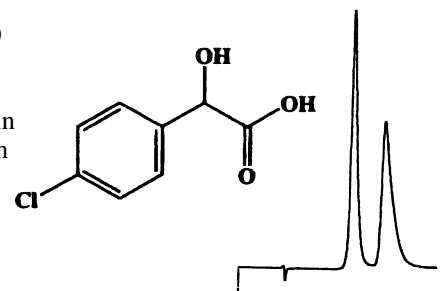


Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA + 0.1%
 Trifluoroacetic Acid
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 11.5 min
 $k'_1 = 5.44$
 $\alpha = 1.34$
 reference 50



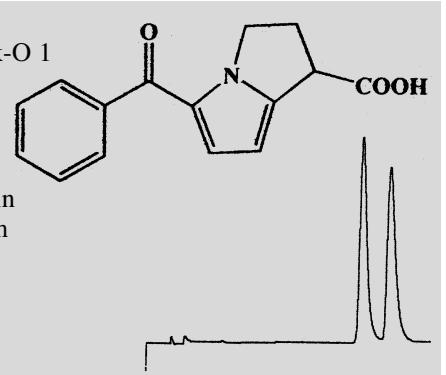
4-Chloromandelic Acid

4-Chloromandelic Acid
 Column = (R,R)-Whelk-O 2
 25 cm x 4.6 mm
 Mobile Phase = (70/30)
 H₂O/CH₃OH
 + 0.1% Acetic Acid
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.0 min
 $k'_1 = 1.95$
 $\alpha = 1.43$
 reference 46



Ketorolac

Ketorolac
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 Hexane/IPA +
 0.1% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 20.0 min
 $k'_1 = 8.87$
 $\alpha = 1.15$
 reference 46

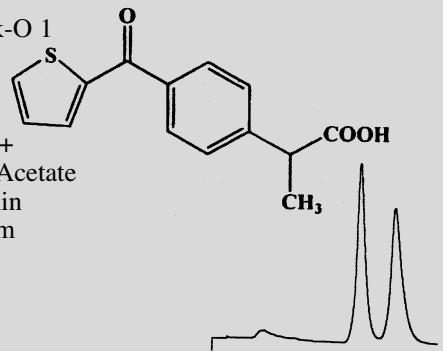


REGIS Other Carboxylic Acids

Suprofen

Suprofen

Column = (S,S)-Whelk-O 1
10/100 (FEC)
25 cm x 4.6 mm
Mobile Phase =
(80/20) Hexane/IPA +
0.01 M Ammonium Acetate
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 18.0 min
 $k'_1 = 9.76$
 $\alpha = 1.27$
reference 46

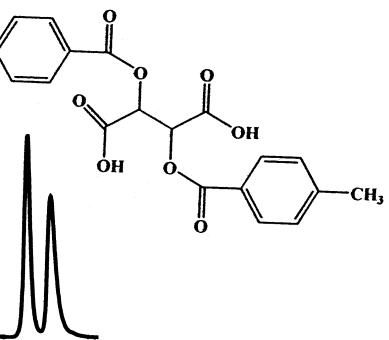


Ditoluoyltartaric Acid

Ditoluoyltartaric Acid

Column: (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA + 0.1% TFA

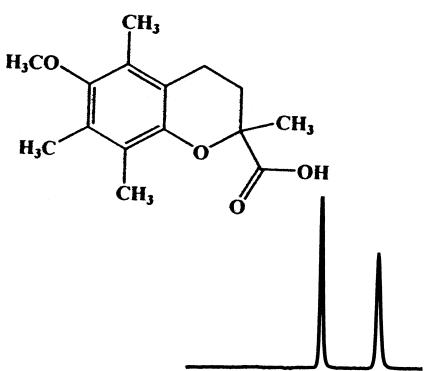
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 12.0 min
 $k'_1 = 2.47$
 $\alpha = 1.19$
reference 48



Trolox-methylether

Trolox-methylether

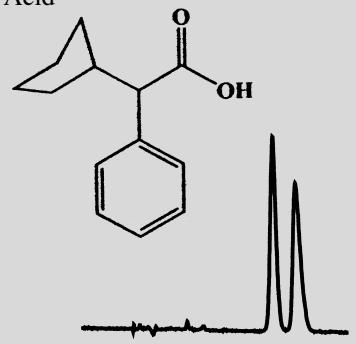
Column: (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA
+ 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 6.0 min
 $k'_1 = 0.32$
 $\alpha = 2.50$
reference 48



1-Cyclopentyl-1-phenylacetic Acid

1-Cyclopentyl-1-phenylacetic Acid

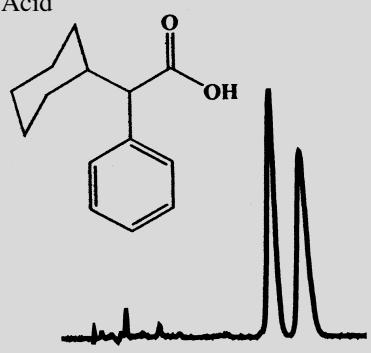
Column: (S,S)-ULMO 25 cm
x 4.6 mm
Mobile Phase: (99/1)
Hexane/IPA + 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 12.0 min
 $k'_1 = 2.46$
 $\alpha = 1.19$
reference 48



1-Cyclohexyl-1-phenylacetic Acid

1-Cyclohexyl-1-phenylacetic Acid

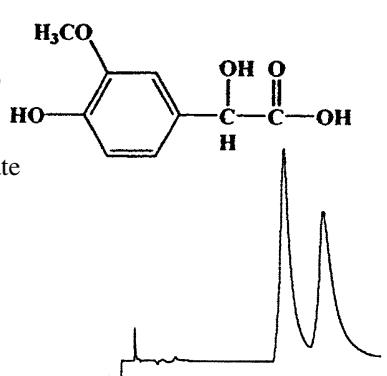
Column: (S,S)-ULMO 25 cm
x 4.6 mm
Mobile Phase: (99/1)
Hexane/IPA + 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 13.0 min
 $k'_1 = 2.53$
 $\alpha = 1.18$
reference 48



Vanilmandelic Acid

Vanilmandelic Acid

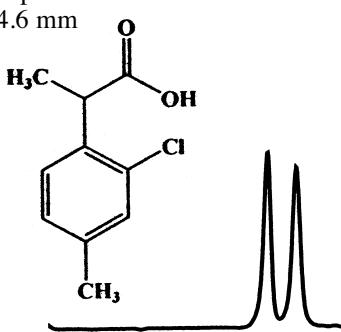
Column: (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6
Mobile Phase: (85/15)
Hexane/Ethanol +
0.01 M Ammonium Acetate
Flow Rate: 2.0 mL/min
Detection: UV 254 nm
Run Time: 22.0 min
 $k'_1 = 12.34$
 $\alpha = 1.27$
reference 46



2-(2-Chloro-4-methylphenoxy)propionic Acid

2-(2-Chloro-4-methylphenoxy)propionic Acid

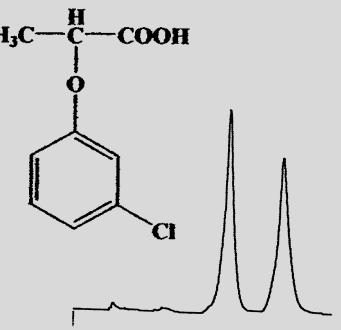
Column: (S,S)-ULMO 25 cm x 4.6 mm
Mobile Phase: (99/1)
Hexane/IPA + 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 11.0 min
 $k'_1 = 2.22$
 $\alpha = 1.11$
reference 48



2-(3-Chlorophenoxy) Propionic Acid

2-(3-Chlorophenoxy) Propionic Acid

Column: (R,R)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6
Mobile Phase: (99/1)
Hexane/IPA
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 17.0 min
 $k'_1 = 6.09$
 $\alpha = 1.42$
reference 46



4-(Trifluoromethyl)mandelic Acid

4-(Trifluoromethyl)mandelic Acid

Column: (S,S)-Whelk-O 1

25 cm x 4.6

Mobile Phase: (92/8)

Hexane/Ethanol +

0.01 M Ammonium

Acetate

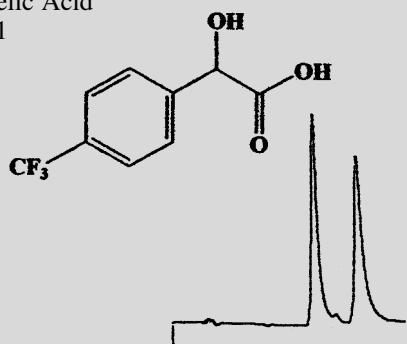
Flow Rate: 1.5 mL/min

Detection: UV 254 nm

Run Time: 11.0 min

 $k'_1 = 3.59$ $\alpha = 1.40$

reference 46

**2,3-Dibenzoyl-Tartaric Acid**

2,3-Dibenzoyl-Tartaric Acid

Column: (R,R)-ULMO

10/100 25 cm x 4.6 mm

Mobile Phase: (90/10)

Hexane/Ethanol +

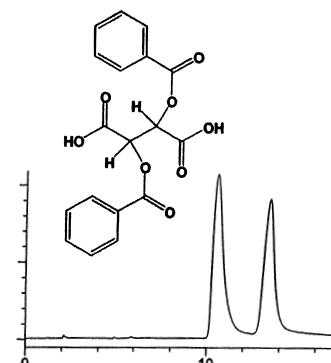
10 mM Ammonium Acetate

Flow Rate: 1.5 mL/min

Detection: UV 254 nm

 $k'_1 = 4.87$ $\alpha = 1.33$

reference 46

**Basic Nitrogen REGIS****Troger's Base**

Column: (R,R)-Whelk-O 1

(10/100) (FEC) 25 cm x 4.6 mm

Mobile Phase: (96/4) Hexane/Ethanol

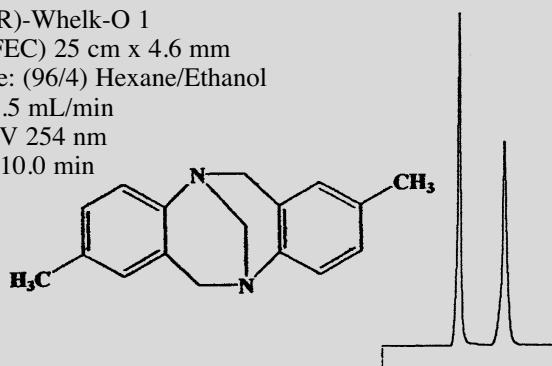
Flow Rate: 1.5 mL/min

Detection: UV 254 nm

Run Time = 10.0 min

 $k'_1 = 2.52$ $\alpha = 1.80$

reference 46

**Basic Amine REGIS****trans-11,12-Diamino-9,10-dihydro-9,10-ethanoanthracene**

trans-11,12-Diamino-9,10-

dihydro-9,10-ethanoanthracene

Column = ChiroSil® RCA(+)

15 cm x 4.6 mm

Mobile Phase = (80/20)

CH₂OH/H₂O

+ 0.1% Phosphoric acid

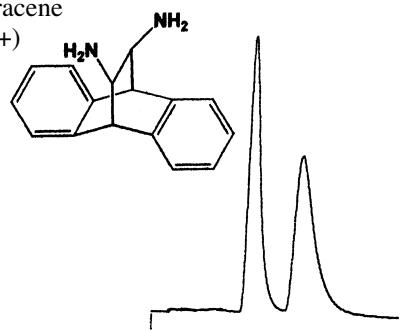
Flow Rate = 1.0 mL/min

Detection = UV 220 nm

Run Time = 10.7 min

 $k'_1 = 3.22$ $\alpha = 1.65$

reference 46



30% EtOH/hexane

1 ml/min; 254 nm

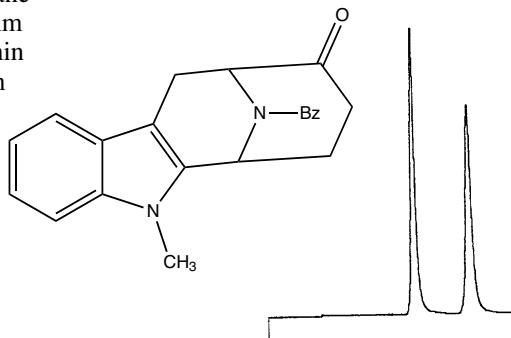
run time = 18 min

4.6 mm x 25 cm

Whelk-O 1

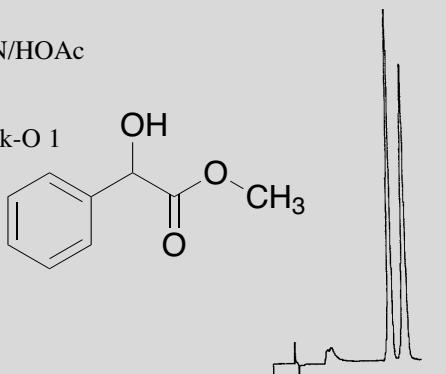
 $k'_1 = 2.46$ $\alpha = 2.09$

reference 18



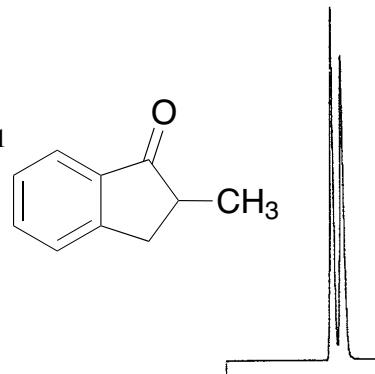
Methyl Mandelate

Methyl Mandelate
 73:27:0.1 H₂O/CH₃CN/HOAc
 1 ml/min; 254 nm
 Run Time = 20 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 5.27$
 $\alpha = 1.15$
 reference 18

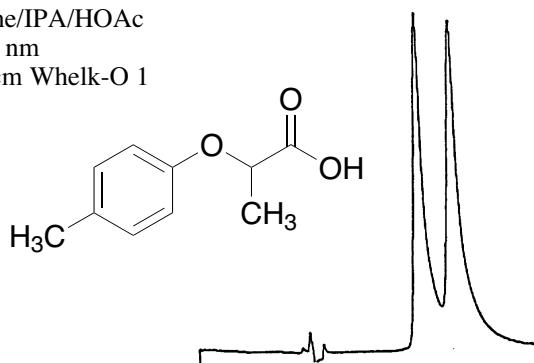


2-Methyl-1-Indanone

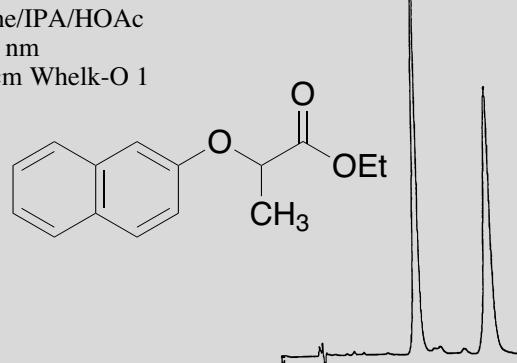
2-Methyl-1-Indanone
 99:1 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 15 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.00$
 $\alpha = 1.12$
 reference 18



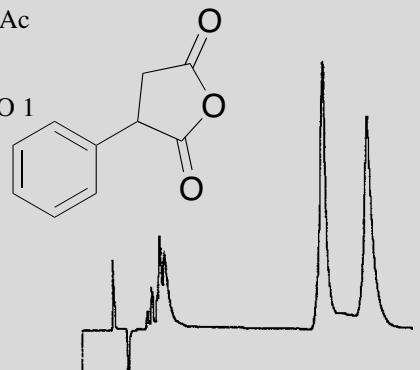
95:5:0.5 hexane/IPA/HOAc
 1 ml/min; 280 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.27$
 $\alpha = 1.28$
 reference 26



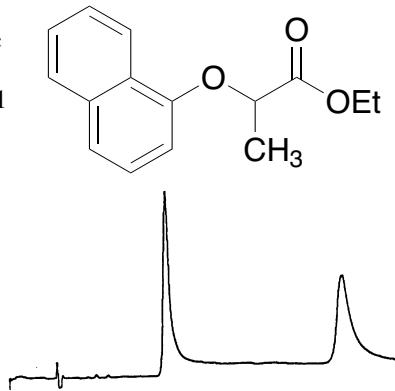
95:5:0.5 hexane/IPA/HOAc
 1 ml/min; 280 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.85$
 $\alpha = 1.70$
 reference 26



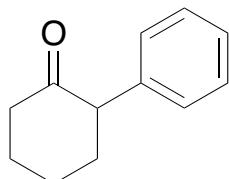
93:7:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 run time = 30 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 8.10$
 $\alpha = 1.21$
 reference 18



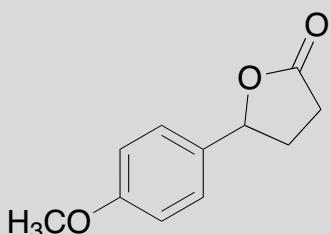
95:5:0.5 hexane/IPA/HOAc
 1 ml/min; 280 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.75$
 $\alpha = 2.53$
 reference 26



90:10 hexane/IPA
 1 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.41$
 $\alpha = 1.81$
 reference 7



20% IPA/hex
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 5.66$
 $\alpha = 1.29$
 reference 7



3-Methyl-1-Indanone

3-Methyl-1-Indanone

 $k'_1 = 6.11$ $\alpha = 1.18$

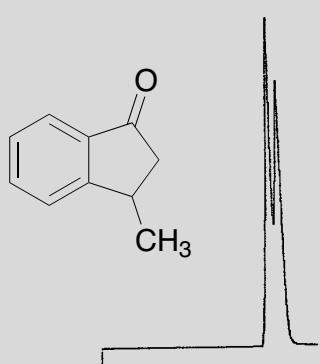
99:1 hexane/IPA

1 ml/min; 254 nm

Run Time = 20 min

4.6 mm x 25 cm Whelk-O 1

reference 18

**DPHB**

DPHB

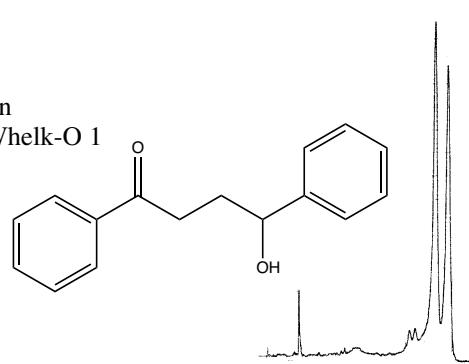
6% EtOH/hexane

1 ml/min; 254 nm

Run Time = 41 min

4.6 mm x 25 cm Whelk-O 1

reference 29



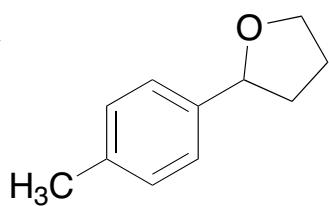
20% IPA/hex

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 1.17$ $\alpha = 1.66$

reference 7



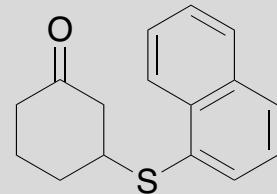
98:2 hexane/IPA

1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 7.82$ $\alpha = 1.12$

reference 7



MeOH/IPA/hexane

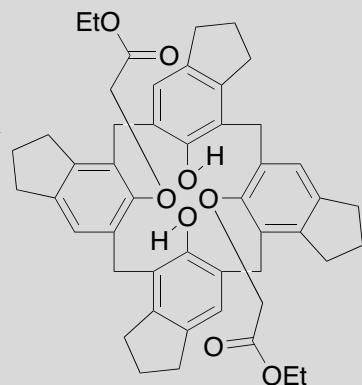
1 ml/min; 254 nm

Run Time = 17 min

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 12.73$ $\alpha = 1.16$

reference 19



Column = L-Leucine

25 cm x 4.6 mm

Mobile Phase = (99.5/0.5)

Hexane/IPA

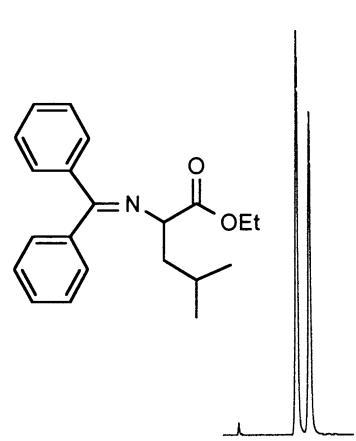
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 11.5 min

 $k'_1 = 2.42$ $\alpha = 1.21$

reference 57



97:3 hexane/IPA

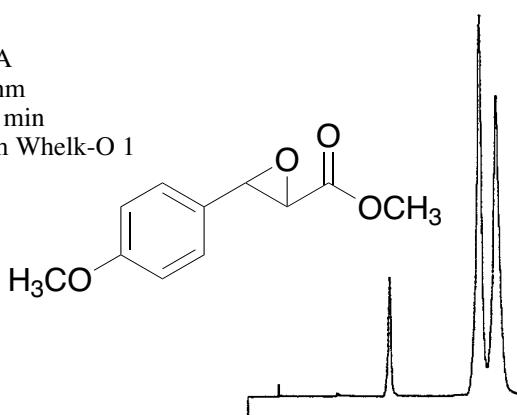
1 ml/min; 254 nm

Run Time = 27 min

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 8.46$ $\alpha = 1.08$

reference 18



10% IPA/hexane

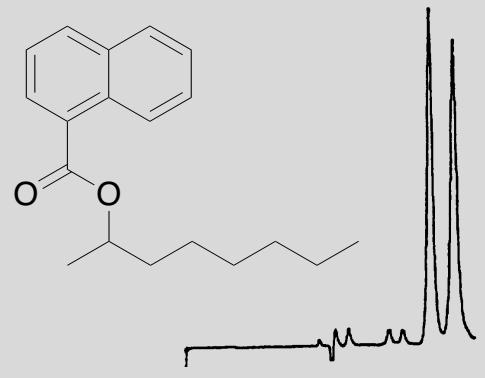
1 ml/min; 254 nm

4.6 mm x 25 cm

Whelk-O 1

 $k'_1 = 2.27$ $\alpha = 1.11$

reference 26



2-Methyl-1-Tetralone

2-Methyl-1-Tetralone

99:1 hexane/IPA

1 ml/min; 254 nm

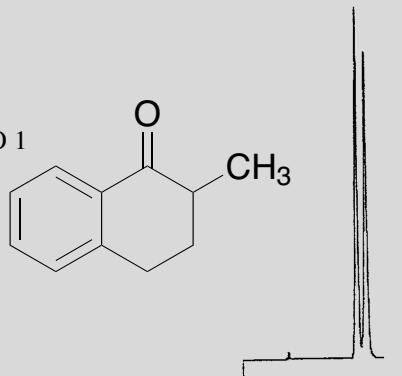
Run Time = 12 min

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 2.76$

$\alpha = 1.11$

reference 18



1,3,5-Triphenylpent-4-yn-1-one

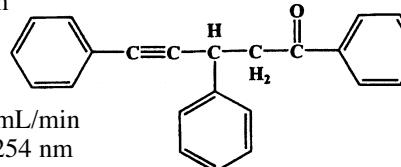
1,3,5-Triphenylpent-4-yn-1-one

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase =

Hexane +
0.5% IPA



Flow Rate = 1.0 mL/min

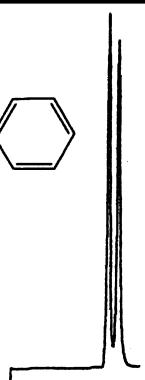
Detection = UV 254 nm

Run Time = 6.5 min

$k'_1 = 1.19$

$\alpha = 1.19$

reference 46



Column = (S,S)-
Whelk-O 1

25 cm x 4.6 mm

Mobile Phase =
(98/2)

Hexane/IPA

Flow Rate = 1.0 mL/min

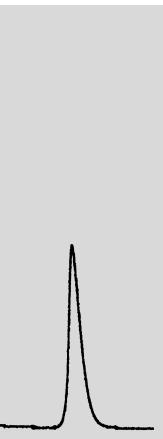
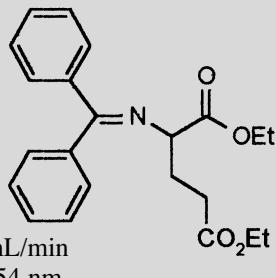
Detection = UV 254 nm

Run Time = 34.0 min

$k'_1 = 8.00$

$\alpha = 1.44$

reference 51



Column = (S,S)- β -Gem 1

25 cm x 4.6 mm

Mobile Phase = (99.5/0.5)
Hexane/IPA

Flow Rate = 1.0 mL/min

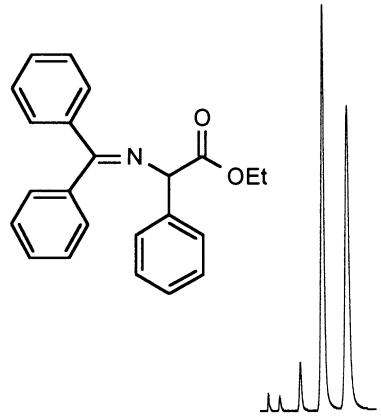
Detection = UV 254 nm

Run Time = 14.5 min

$k'_1 = 2.67$

$\alpha = 1.43$

reference 57



Diperodon

Diperodon

Column = (R)- α -Burke 2 25 cm x 4.6 mm

Mobile Phase = (48/48/4) CH₂Cl₂/Hexane/Ethanol
+ 1.5 mM Ammonium Acetate

Flow Rate = 1.0 mL/min

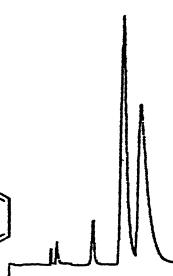
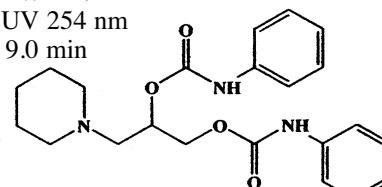
Detection = UV 254 nm

Run Time = 9.0 min

$k'_1 = 1.7$

$\alpha = 1.25$

reference 46



10% IPA/hexane

1 ml/min; 254 nm

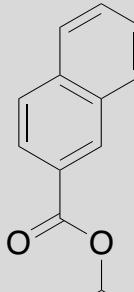
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 0.67$

$\alpha = 1.16$

reference 26



Column = (S,S)-
Whelk-O 1

25 cm x 4.6 mm

Mobile Phase =
(98/2)

Hexane/IPA

Flow Rate = 1.0 mL/min

Detection =

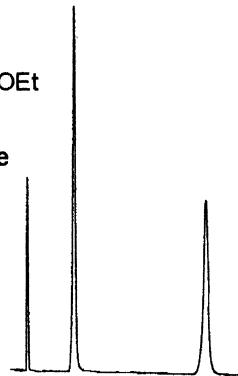
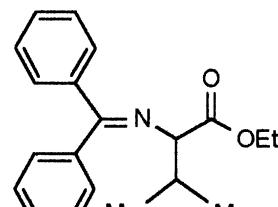
UV 254 nm

Run Time = 20.5 min

$k'_1 = 1.62$

$\alpha = 4.18$

reference 51



Column = D-Phenylglycine

25 cm x 4.6 mm

Mobile Phase = 99/1)

Hexane/IPA

Flow Rate = 1.0 mL/min

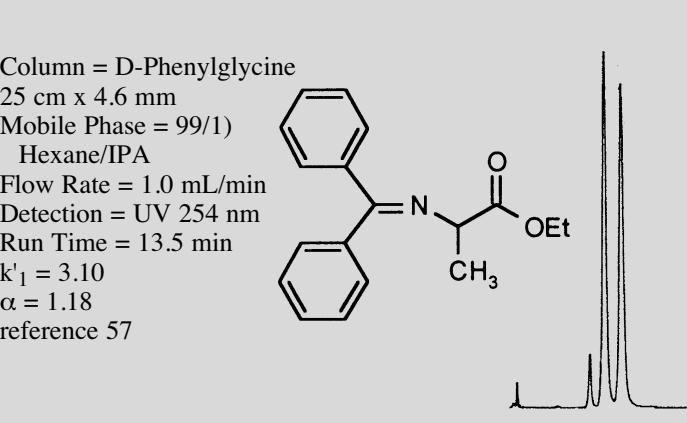
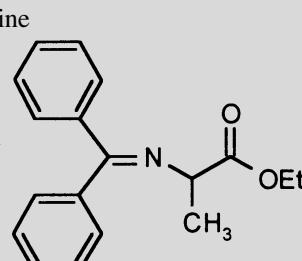
Detection = UV 254 nm

Run Time = 13.5 min

$k'_1 = 3.10$

$\alpha = 1.18$

reference 57



Buckminsterfullerene-Enone [2+2] Photoadducts

Semi-prep separation on analytical column

2:1 toluene/hexane

1 ml/min; 400 nm

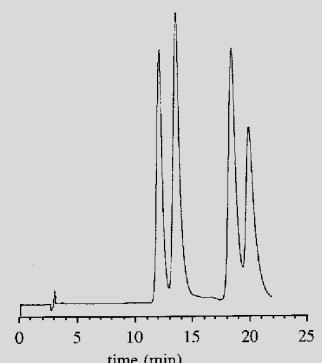
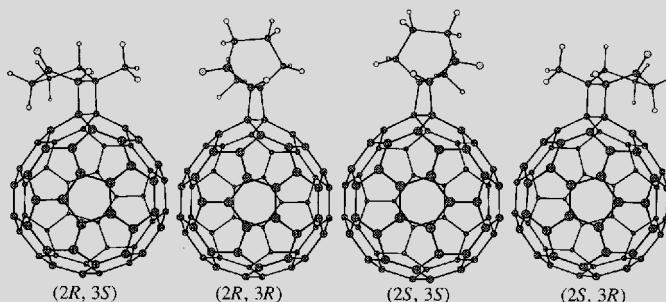
Run Time = 22 min

Sample: 100 μ l of

5 mg/ml solution (0.5 mg)

4.6 mm x 25 cm Whelk-O 1

reference 8

**Ethyl-2-(p-Hydroxyphenoxy) Propionate**

Ethyl-2-(p-Hydroxyphenoxy) Propionate

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (98/2)

Hexane/Ethanol

Flow Rate = 2.0 mL/min

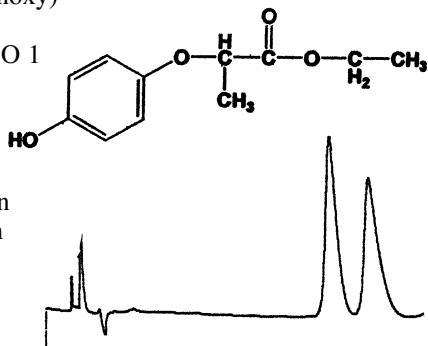
Detection = UV 254 nm

Run Time = 21.1 min

k'_1 = 12.72

α = 1.15

reference 46

**Tert-butyl-2-(benzamido) cyclopentyl carbamate**

Tert-butyl-2-(benzamido) cyclopentyl carbamate

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (95/5)

Hexane/IPA

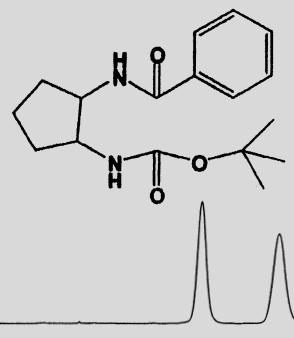
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

k'_1 = 3.65

α = 1.46

reference 46

**1'-Acetoxychavicol Acetate**

1'-Acetoxychavicol Acetate

Column = (R,R)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (90/10)

Hexane/IPA

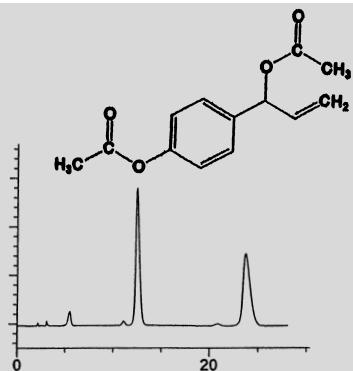
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

k'_1 = 5.94

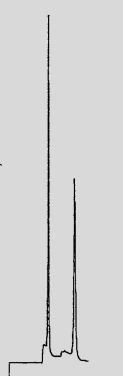
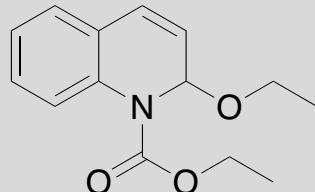
α = 2.05

reference 46

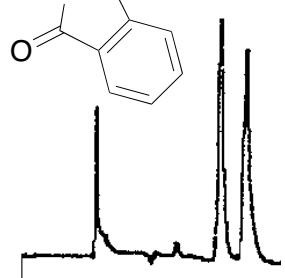
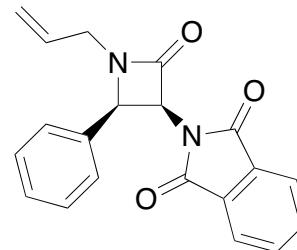


EEDQ

EEDQ
90:10 hexane/IPA
1 ml/min; 254 nm
Run Time = 25 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.53$
 $\alpha = 2.13$
reference 18

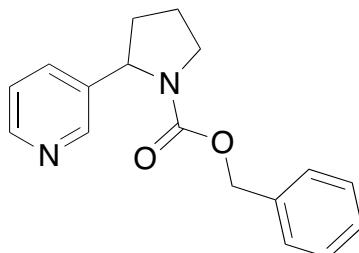


methanol
2 ml/min; 254 nm
Run Time = 6 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.75$
 $\alpha = 1.20$
reference 7

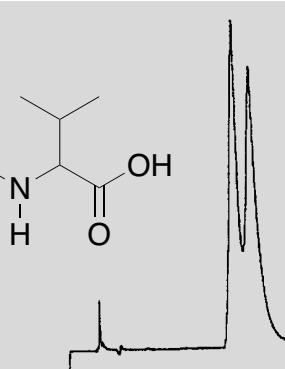
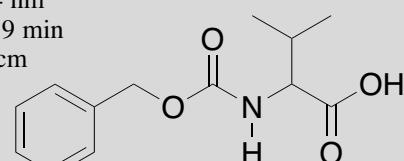


CBZ nornicotine

CBZ nornicotine
1:3 MeOH/dichloromethane
1 ml/min; 254 nm
Run Time = 5 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.37$
 $\alpha = 1.38$
reference 7

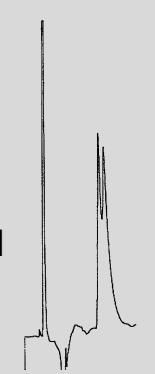
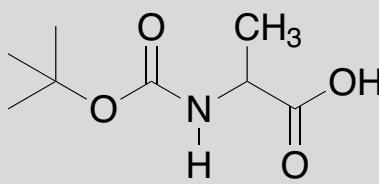


CBZ-Val
95:5:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 19 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 5.49$
 $\alpha = 1.13$
reference 18

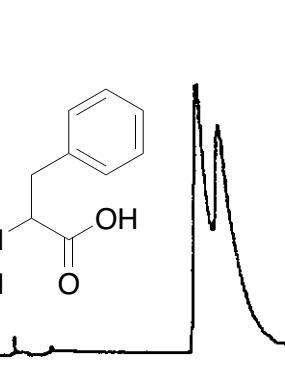
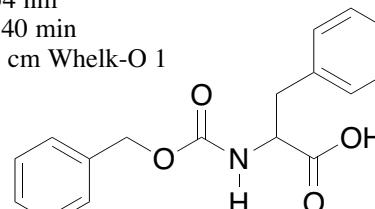


BOC-Ala

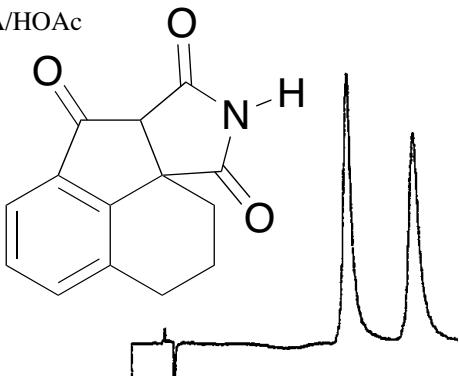
BOC-Ala
98:2:0.2 hexane/IPA/HOAc
1 ml/min; 220 nm
Run Time = 17 min
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.43$
 $\alpha = 1.09$
reference 18



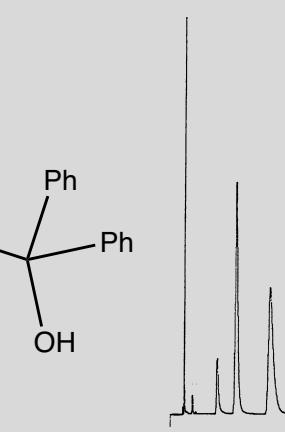
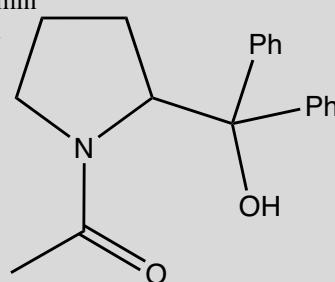
CBZ-Phe
95:5:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 40 min
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 10.2$
 $\alpha = 1.20$
reference 18

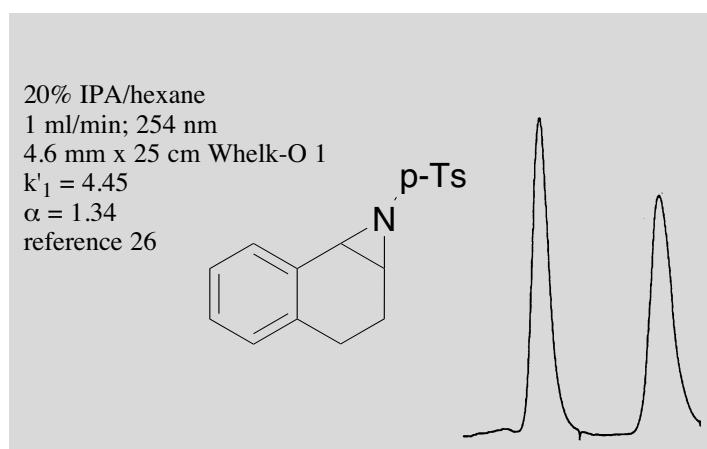
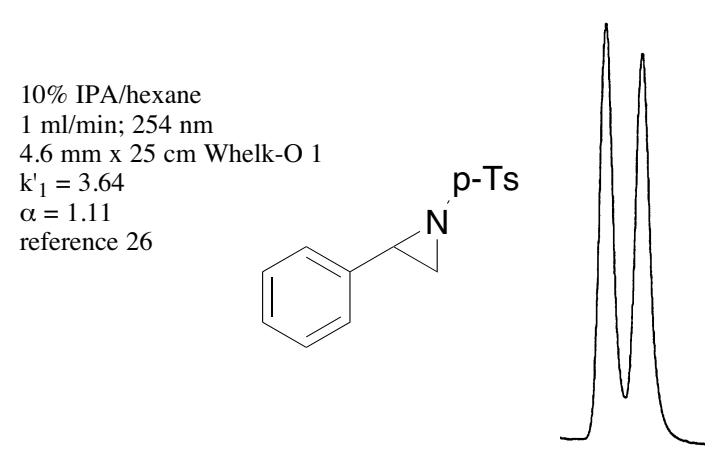
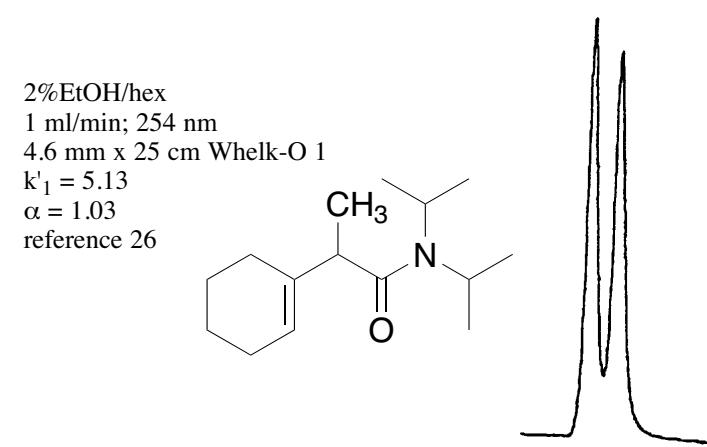
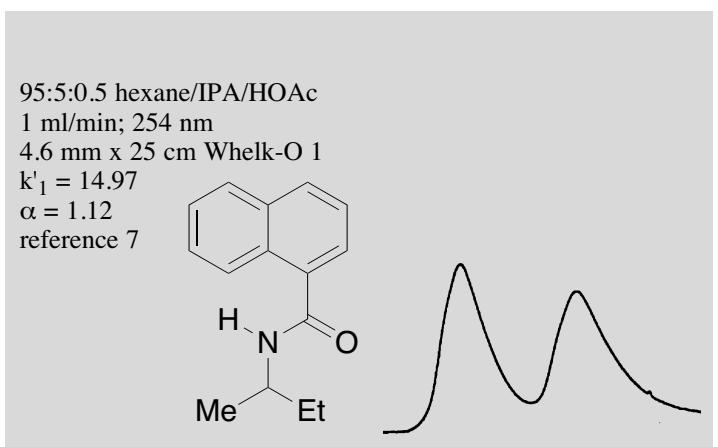
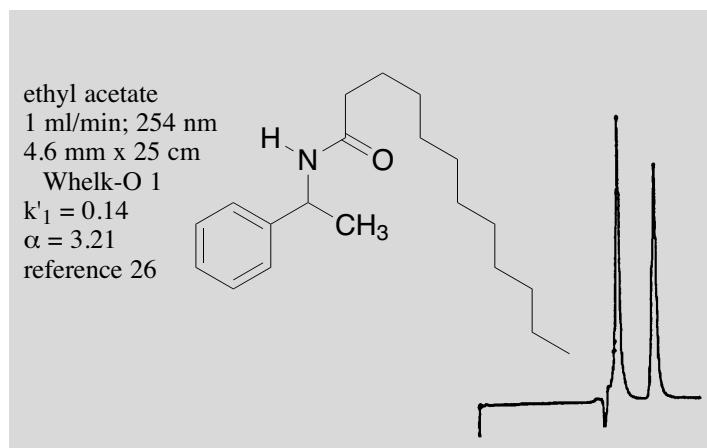
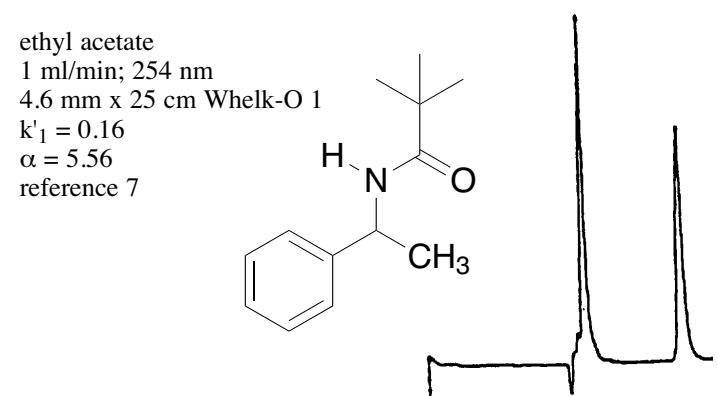
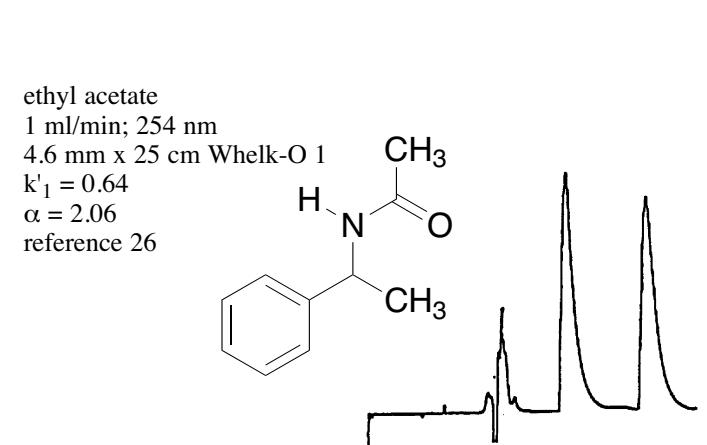
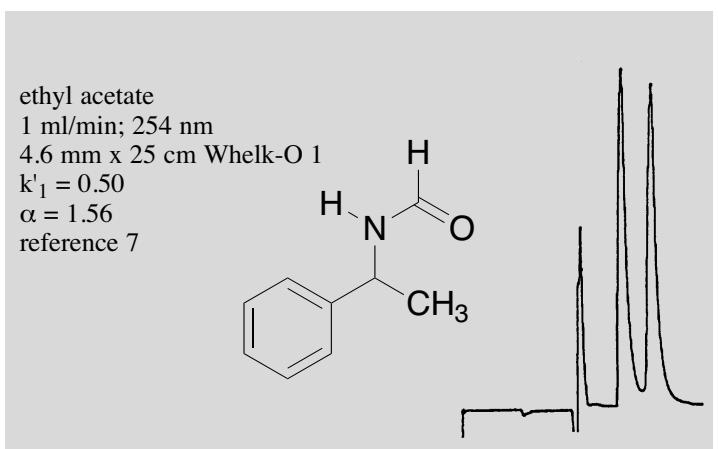


80:20:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 25 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 5.97$
 $\alpha = 1.36$
reference 18



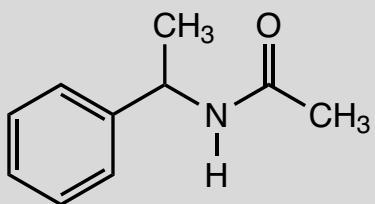
15% EtOH/hexane
1 ml/min; 254 nm
Run Time = 16 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 3.79$
 $\alpha = 1.66$
reference 18



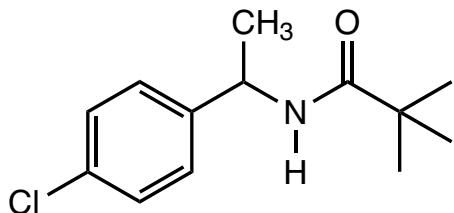


REGIS Amides, Imides, Carbamates, etc.

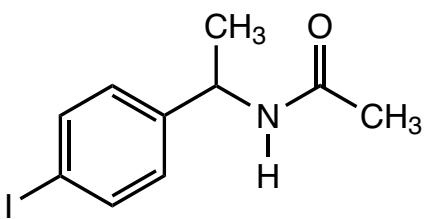
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 3.72$
 $\alpha = 3.17$
reference 38



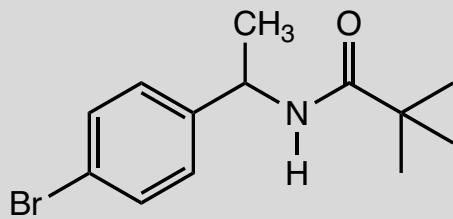
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.48$
 $\alpha = 11.6$
reference 38



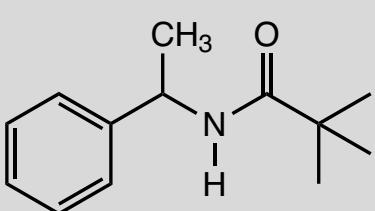
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 4.10$
 $\alpha = 5.12$
reference 38



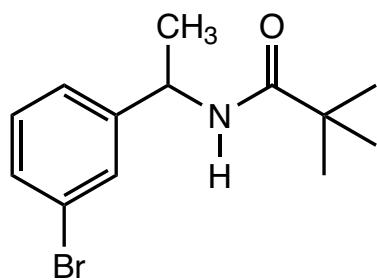
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.61$
 $\alpha = 12.8$
reference 38



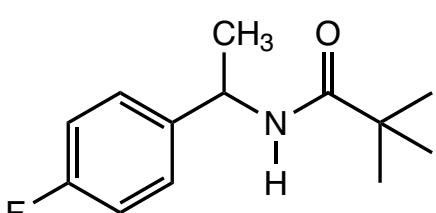
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.39$
 $\alpha = 6.74$
reference 38



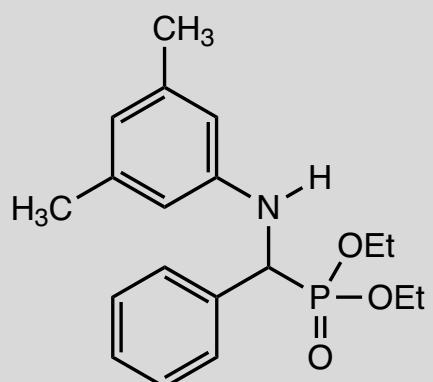
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.75$
 $\alpha = 13.7$
reference 38



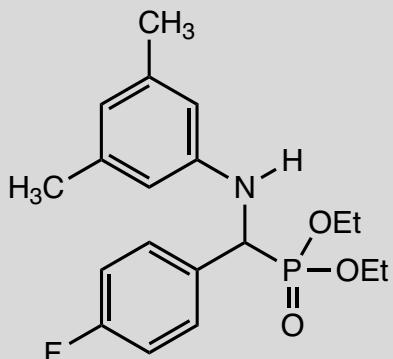
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.17$
 $\alpha = 7.29$
reference 38



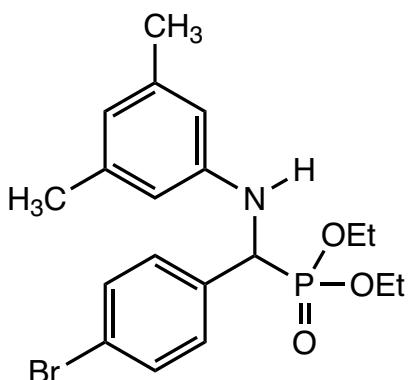
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 10.87$
 $\alpha = 1.29$
reference 38



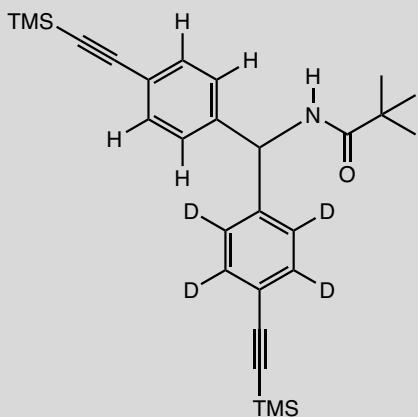
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 0.83$
 $\alpha = 1.39$
reference 38



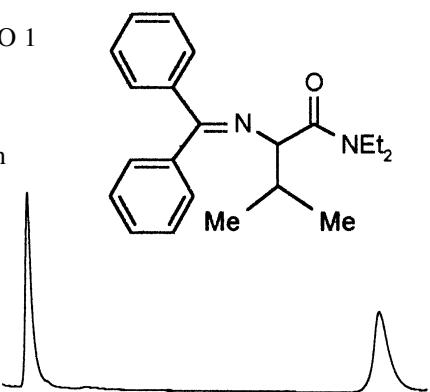
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 0.86$
 $\alpha = 1.66$
reference 38



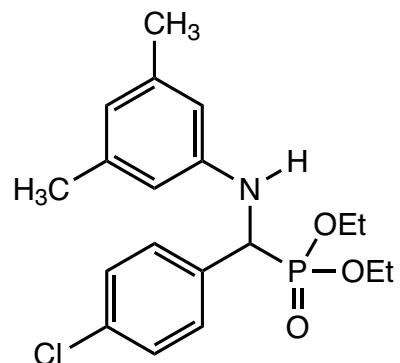
10% acetonitrile
in CO_2
(*S,S*) Whelk-O 1
 $k'_1 = 25$
 $\alpha = 1.025$
reference 39



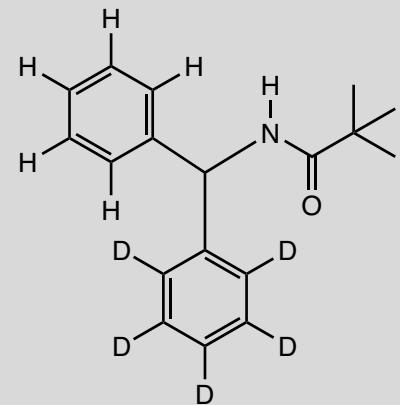
Column = (*S,S*)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (90/10)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 46.0 min
 $k'_1 = 2.70$
 $\alpha = 6.02$
reference 51



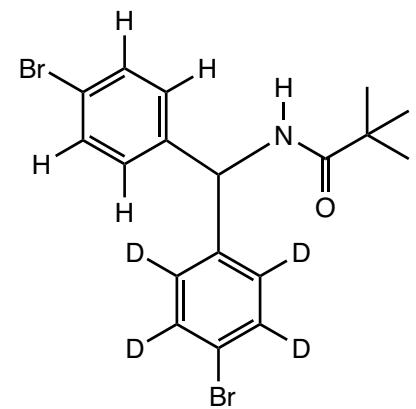
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 0.84$
 $\alpha = 1.55$
reference 38



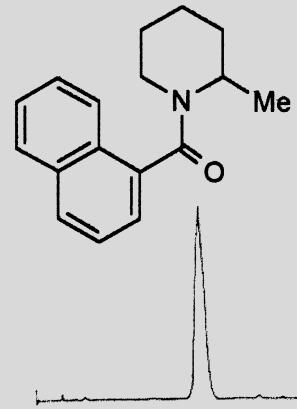
10% acetonitrile
in CO_2
(*S,S*) Whelk-O 1
 $k'_1 = 8.5$
 $\alpha = 1.025$
reference 39



10% acetonitrile
in CO_2
(*S,S*) Whelk-O 1
 $k'_1 = 19.7$
 $\alpha = 1.025$
reference 39



Column = (*R,R*)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (80/20)
Hexane/IPA
Flow Rate = 2.0
mL/min
Detection = UV 254
nm
Run Time = 19.0 min
 $k'_1 = 7.53$
 $\alpha = 1.77$
reference 52



β -Lactam

β -Lactam

Column: (S,S)-DACH-DNB
25 cm x 4.6 mm

Mobile Phase: (48/48/2)
Hex/CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

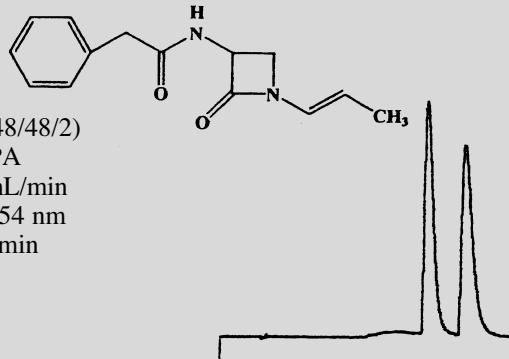
Detection: UV 254 nm

Run Time: 14.0 min

k' : 3.40

α : 1.33

reference 59



Cyclopentyl Benzoyl-Diamide

Cyclopentyl Benzoyl-Diamide

Column: (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase: (90/10)

Hexane/IPA

Flow Rate: 1.5 mL/min

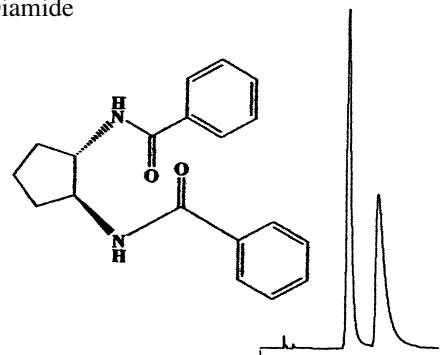
Detection: UV 254 nm

Run Time: 8.7 min

k' : 2.62

α : 1.47

reference 46



REGIS Epoxides

Styrene Oxide

Styrene Oxide

1% IPA/hexane

1 ml/min; 254 nm

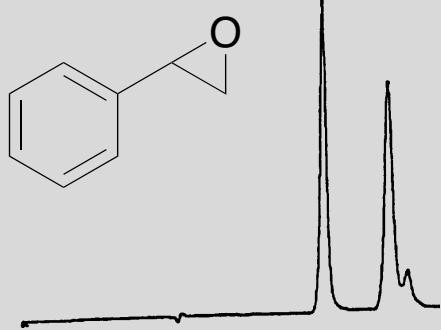
4.6 mm x 25 cm

Whelk-O 1

k' = 1.37

α = 1.37

reference 18



Stilbene Oxide

Stilbene Oxide

10% IPA/hexane

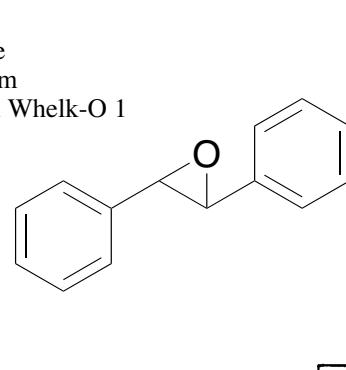
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

k' = 0.45

α = 2.00

reference 18



0.1% HOAc in hexane

1 ml/min; 254 nm

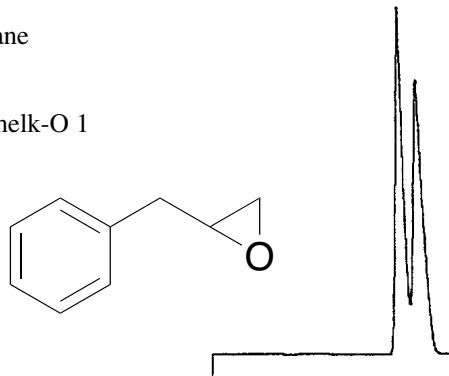
Run Time = 20 min

4.6 mm x 25 cm Whelk-O 1

k' = 5.92

α = 1.12

reference 18



m-Cl Styrene Oxide

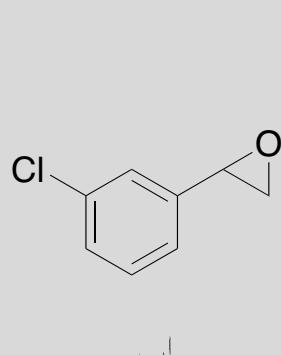
hexane

1 ml/min; 220 nm

4.6 mm x 25 cm

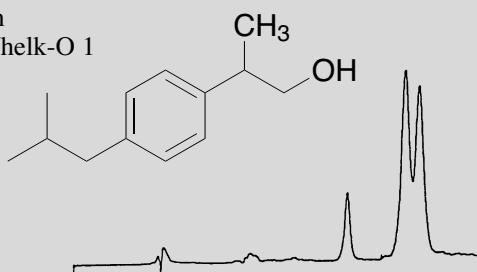
Whelk-O 1

reference 30

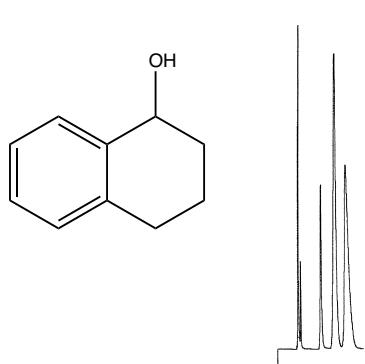


Ibuprofenol

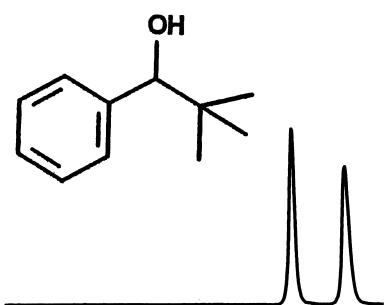
Ibuprofenol
99:1 hexane/IPA
1 ml/min; 254 nm
Run Time = 14 min
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.38$
 $\alpha = 1.05$
reference 26

**1,2,3,4-Tetrahydro-1-Naphthol**

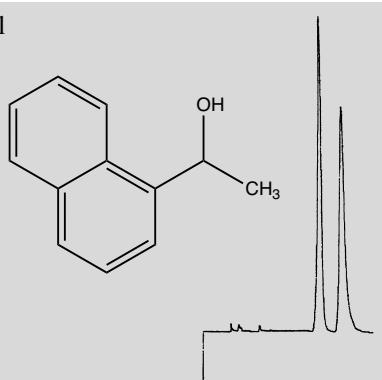
1,2,3,4-Tetrahydro-1-Naphthol
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 10.5 min
 $k'_1 = 2.17$
 $\alpha = 1.30$
reference 46

**Tert Butyl Phenyl Carbinol**

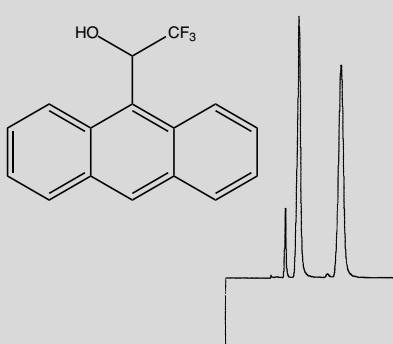
Tert Butyl Phenyl Carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 6.0 min
 $k'_1 = 4.60$
 $\alpha = 1.46$
reference 46

 **α -Naphthyl Methyl Carbinol**

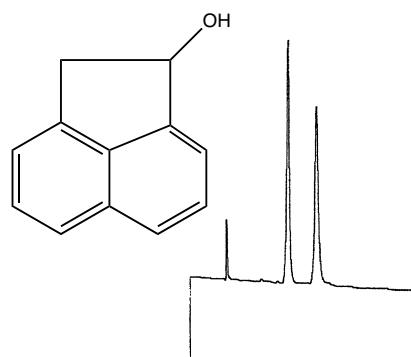
α -Naphthyl Methyl Carbinol
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 14.5 min
 $k'_1 = 3.49$
 $\alpha = 1.25$
reference 46

**9-Anthryl Trifluoromethyl Carbinol**

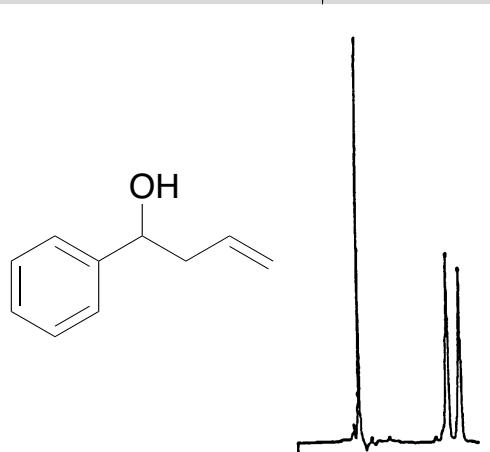
9-Anthryl Trifluoromethyl Carbinol
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 10 min
 $k'_1 = 1.36$
 $\alpha = 2.02$
reference 46

**Acenaphthenol**

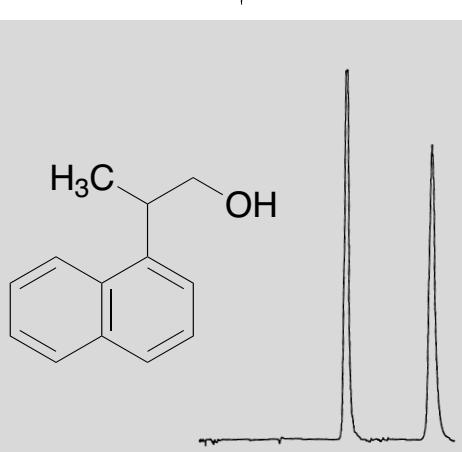
Acenaphthenol
Column: (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase: (95/5)
Hexane/IPA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 10 min
 $k'_1: 1.68$
 $\alpha: 1.46$
reference 46



2% IPA/hexane
1 ml/min; 220 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.76$
 $\alpha = 1.13$
reference 18



80:20 hexane/IPA
1 ml/min; 254 nm
run time = 10 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.22$
 $\alpha = 2.08$
reference 26



Beta Naphthyl Methyl Carbinol

Beta Naphthyl Methyl Carbinol

Column: (R,R)-ULMO

25 cm x 4.6 mm

Mobile Phase: (97/3) Hexane/IPA

Flow Rate: 1.0 mL/min

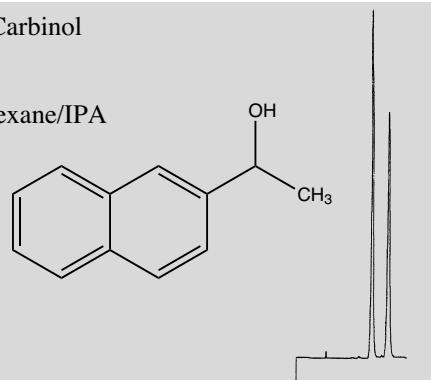
Detection: UV 254 nm

Run Time: 9 min

k'_1 : 1.64

α : 1.34

reference 46



Tetrahydrobenzopyrene-7-ol

Tetrahydrobenzopyrene-7-ol

80:20 hexane/IPA

1 ml/min; 254 nm

run time = 22 min

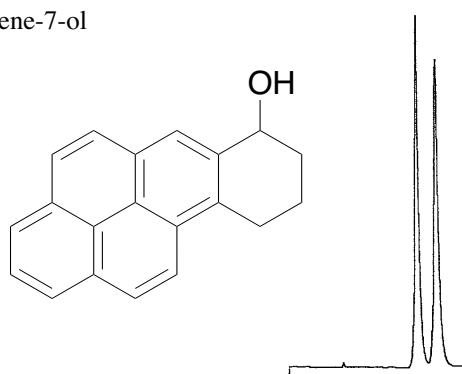
4.6 mm x 25 cm

Whelk-O 1

k'_1 = 6.10

α = 1.18

reference 18



1-Naphthyl-2-butanol

1-Naphthyl-2-butanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (95/5)

Heptane/IPA

Flow Rate = 1.0 mL/min

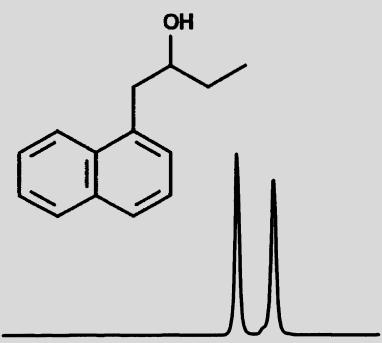
Detection = UV 215 nm

Run Time = 6 min

k'_1 = 0.80

α = 1.35

reference 48



1% IPA/hexane

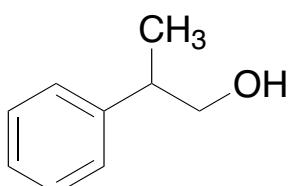
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

k'_1 = 3.38

α = 1.05

reference 7



1,1'-Bi-2-Naphthol

1,1'-Bi-2-Naphthol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98/2)

Hexane/IPA + 0.1% TFA

Flow Rate = 1.0 mL/min

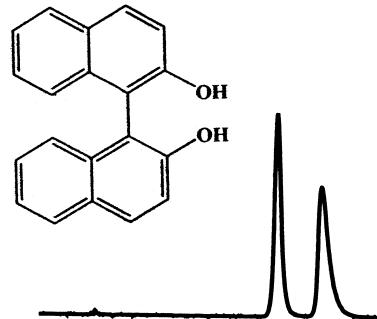
Detection = UV 254 nm

Run Time = 18.0 min

k'_1 = 4.84

α = 1.24

reference 48



9-Anthrylethanol

9-Anthrylethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (95/5) Heptane/IPA

Flow Rate = 1.0 mL/min

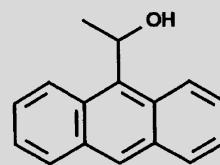
Detection = UV 215 nm

Run Time = 12 min

k'_1 = 1.82

α = 1.74

reference 48



2-Naphthyl-2-butanol

2-Naphthyl-2-butanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (95/5)

Heptane/IPA

Flow Rate = 1.0 mL/min

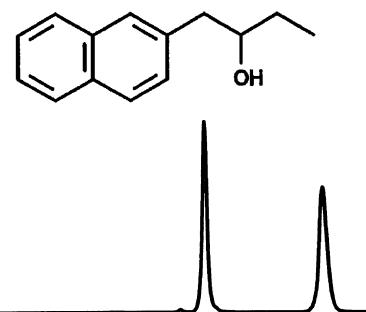
Detection = UV 215 nm

Run Time = 8 min

k'_1 = 1.00

α = 1.93

reference 48



Column = (S,S)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (99/1)

Hexane/IPA

Flow Rate = 1.0 mL/min

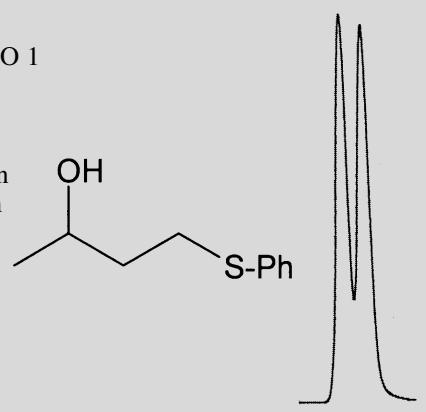
Detection = UV 254 nm

Run Time = 18.5 min

k'_1 = 5.59

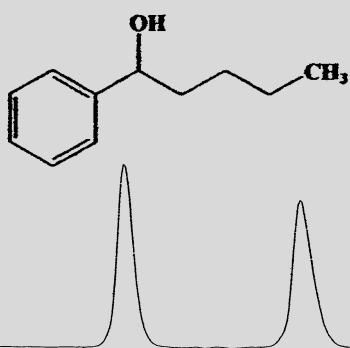
α = 1.09

reference 55

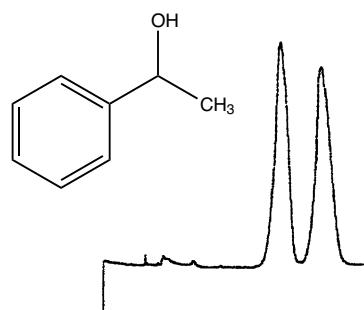


1-Phenylpentanol

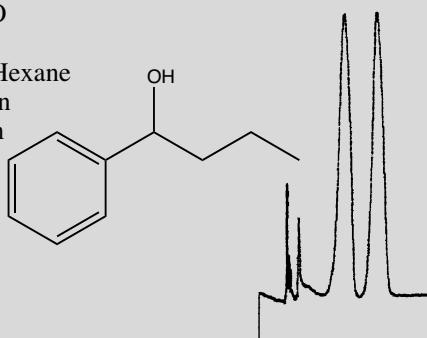
1-Phenylpentanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 7.0 min
 $k'_1 = 1.65$
 $\alpha = 1.45$
 reference 60

**Phenyl Methyl Carbinol**

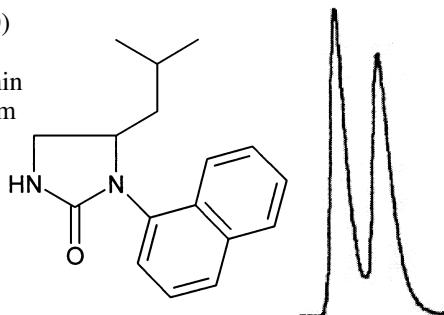
Phenyl Methyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = 100% Hexane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14 min
 $k'_1 = 3.11$
 $\alpha = 1.30$
 reference 46

**Phenyl Propyl Carbinol**

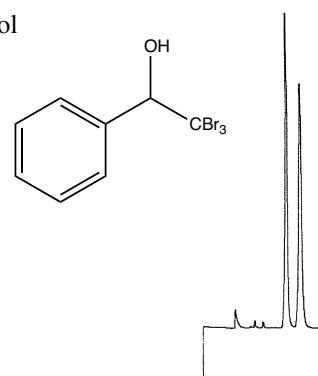
Phenyl Propyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = 100% Hexane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 12 min
 $k'_1 = 2.25$
 $\alpha = 1.56$
 reference 46



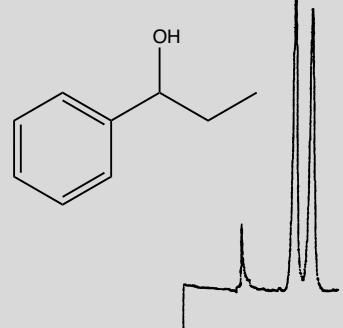
Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (80/20)
 Hexane/IPA
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 24.0 min
 $k'_1 = 13.30$
 $\alpha = 1.11$
 reference 55

**Phenyl Tribromomethyl Carbinol**

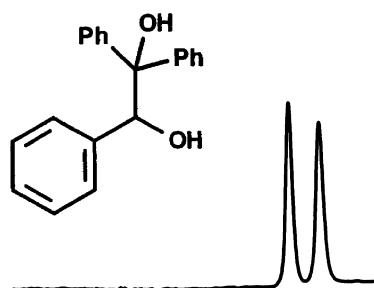
Phenyl Tribromomethyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 9 min
 $k'_1 = 1.87$
 $\alpha = 1.25$
 reference 46

**Phenyl Ethyl Carbinol**

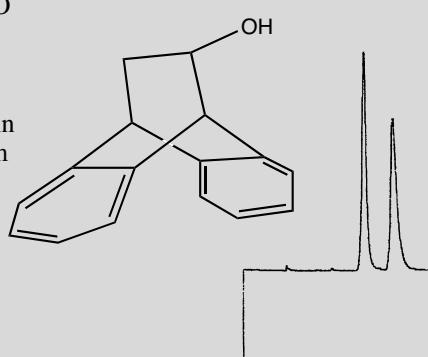
Phenyl Ethyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 6.5 min
 $k'_1 = 1.06$
 $\alpha = 1.30$
 reference 46

**1,1,2,-Triphenyl-1,2-Ethanediol**

1,1,2,-Triphenyl-1,2-Ethanediol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 13 min
 $k'_1 = 2.59$
 $\alpha = 1.14$
 reference 48

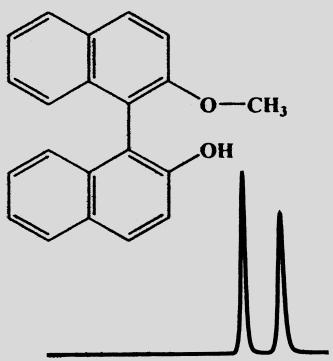


Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10 min
 $k'_1 = 1.97$
 $\alpha = 1.37$
 reference 48



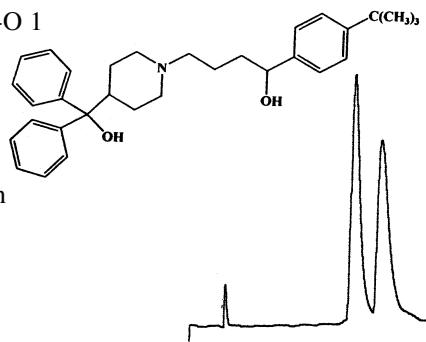
1,1'-Binaphthol Monomethylether

1,1'-Binaphthol
Monomethylether
Column: (S,S)-ULMO 25 cm x 4.6 mm
Mobile Phase: (98/2) Hexane/IPA + 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 11.0 min
 $k'_1 = 2.23$
 $\alpha = 1.28$
reference 48



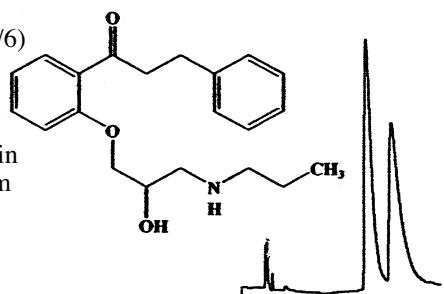
Terfenadine

Terfenadine
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (97/3)
Hexane/Ethanol + 0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 5.91$
 $\alpha = 1.20$
reference 46



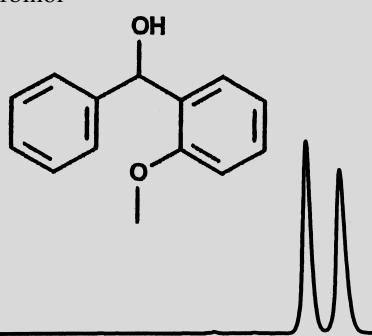
Propafenone

Propafenone
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (47/47/6)
CH₂Cl₂/Hexane/Ethanol + 0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 11.0 min
 $k'_1 = 3.99$
 $\alpha = 1.25$
reference 48



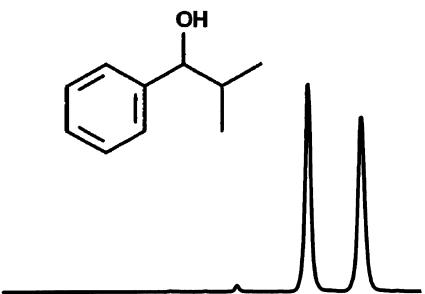
2-Methoxyphenyl Phenyl Carbinol

2-Methoxyphenyl Phenyl Carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 12.0 min
 $k'_1 = 2.92$
 $\alpha = 1.13$
reference 48



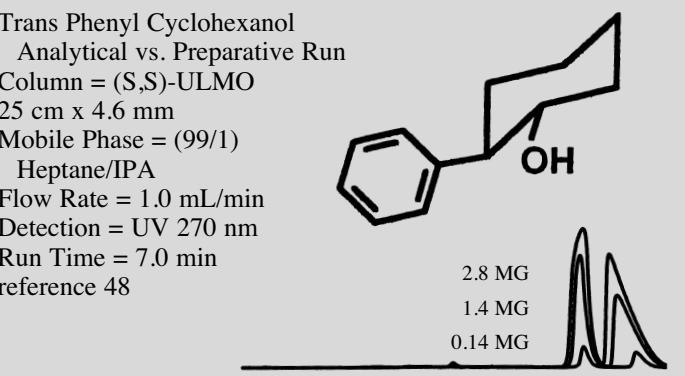
Phenyl isopropyl carbinol

Phenyl isopropyl carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time: 6 min
 $k'_1 = 0.86$
 $\alpha = 1.38$
reference 48



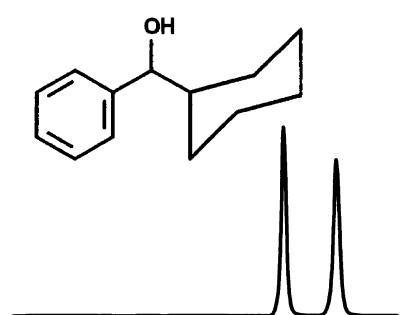
Trans Phenyl Cyclohexanol Analytical vs. Preparative Run

Trans Phenyl Cyclohexanol
Analytical vs. Preparative Run
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 270 nm
Run Time = 7.0 min
reference 48



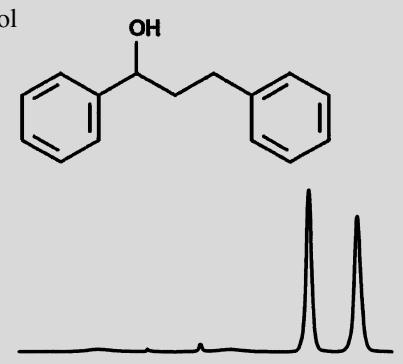
Phenyl cyclohexyl carbinol

Phenyl cyclohexyl carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase: (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 6.5 min
 $k'_1 = 0.97$
 $\alpha = 1.39$
reference 48



Phenyl phenylethyl carbinol

Phenyl phenylethyl carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 9.5 min
 $k'_1 = 1.81$
 $\alpha = 1.30$
reference 48



Methyl 3-phenyl-3azido-2hydroxypropanoate (Erythro-diastereomer)

Methyl 3-phenyl-3azido-2hydroxypropanoate
(Erythro-diastereomer)

Column = (S,S)-ULMO 25 cm x 4.6 mm

Mobile Phase = (97/3)

Heptane/Glyme

Flow Rate = 1.0 mL/min

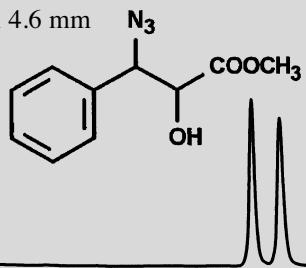
Detection = UV 215 nm

Run Time = 10.5 min

$k'_1 = 2.34$

$\alpha = 1.16$

reference 60



1-(4-Methoxyphenyl)-2-propanol

1-(4-Methoxyphenyl)-2-propanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

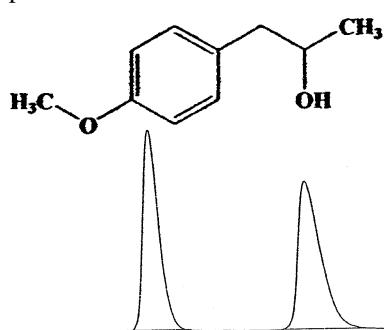
Detection = UV 254 nm

Run Time = 17.5 min

$k'_1 = 5.33$

$\alpha = 1.28$

reference 60



2-Thiopheneethanol

2-Thiopheneethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

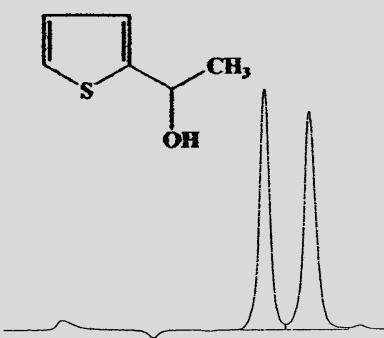
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 2.21$

$\alpha = 1.12$

reference 60



1-(4-Hydroxyphenyl) Ethanol

1-(4-Hydroxyphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (95/5)

n-Heptane/IPA + 0.1% TFA

Flow Rate = 1.0 mL/min

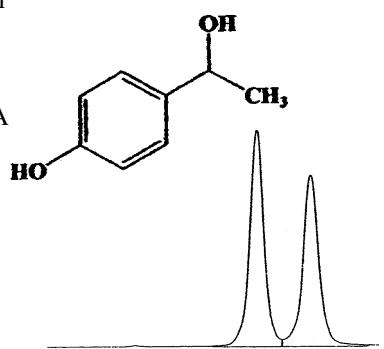
Detection = UV 254 nm

Run Time = 8.5 min

$k'_1 = 1.491$

$\alpha = 1.16$

reference 60



1-(4-Methoxyphenyl)-2-butanol

1-(4-Methoxyphenyl)-2-butanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

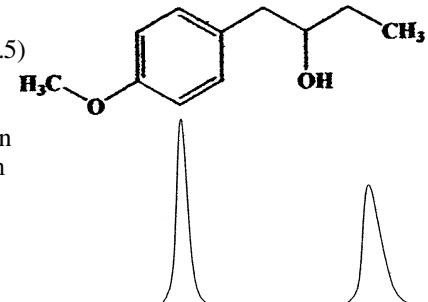
Detection = UV 254 nm

Run Time = 12.0 min

$k'_1 = 2.04$

$\alpha = 1.49$

reference 60



1-Phenyl-2-propanol

1-Phenyl-2-propanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.5 mL/min

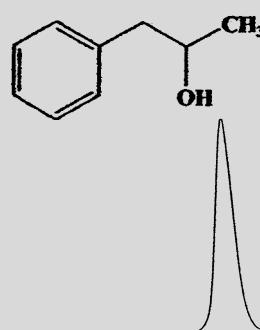
Detection = UV 254 nm

Run Time = 6.5 min

$k'_1 = 1.72$

$\alpha = 1.19$

reference 60



3-Thiopheneethanol

3-Thiopheneethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

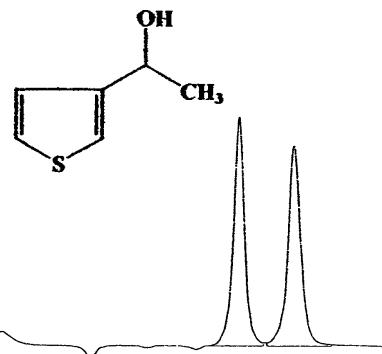
Detection = UV 254 nm

Run Time = 11.5 min

$k'_1 = 2.42$

$\alpha = 1.13$

reference 60



1-(o-Methoxyphenyl) Ethanol

1-(o-Methoxyphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.5 mL/min

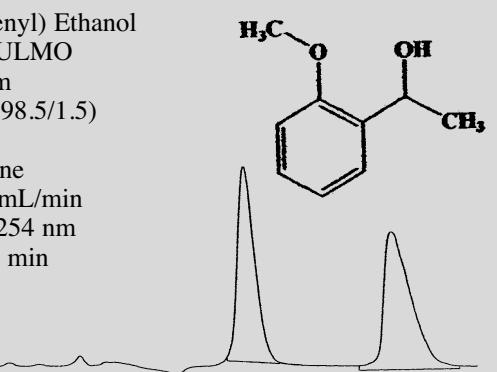
Detection = UV 254 nm

Run Time = 11.0 min

$k'_1 = 3.27$

$\alpha = 1.29$

reference 60



1-[(4-Phenyl) phenyl] Ethanol

1-[(4-Phenyl) phenyl] Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 2.0 mL/min

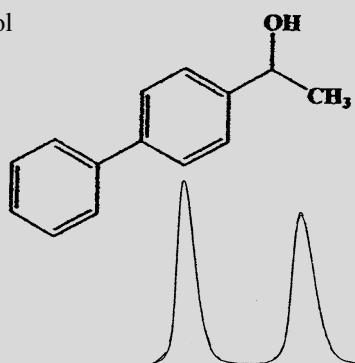
Detection = UV 254 nm

Run Time = 8.5 min

$k'_1 = 3.76$

$\alpha = 1.21$

reference 60



1-(4-Benzyl) phenyl Ethanol

1-(4-Benzyl) phenyl Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 2.0 mL/min

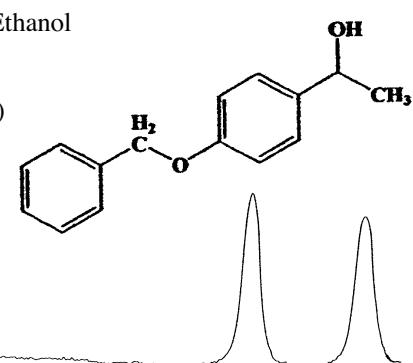
Detection = UV 254 nm

Run Time = 11.0 min

$k'_1 = 5.21$

$\alpha = 1.21$

reference 60



1-(p-Bromophenyl) Ethanol

1-(p-Bromophenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

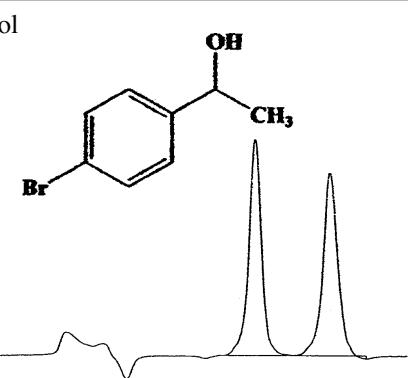
Detection = UV 254 nm

Run Time = 11.5 min

$k'_1 = 2.39$

$\alpha = 1.17$

reference 60



1-(p-Fluorophenyl) Ethanol

1-(p-Fluorophenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

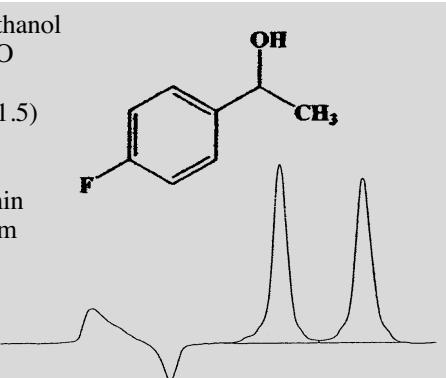
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 2.13$

$\alpha = 1.16$

reference 60



1-(m-Trifluoromethylphenyl) Ethanol

1-(m-Trifluoromethylphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

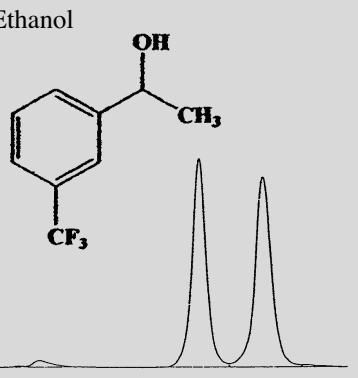
Detection = UV 254 nm

Run Time = 9.0 min

$k'_1 = 1.66$

$\alpha = 1.14$

reference 60



1-(p-Methylphenyl) Ethanol

1-(p-Methylphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

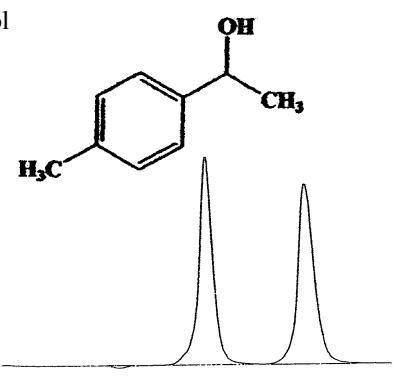
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 2.06$

$\alpha = 1.21$

reference 60



1-(m-Methylphenyl) Ethanol

1-(m-Methylphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

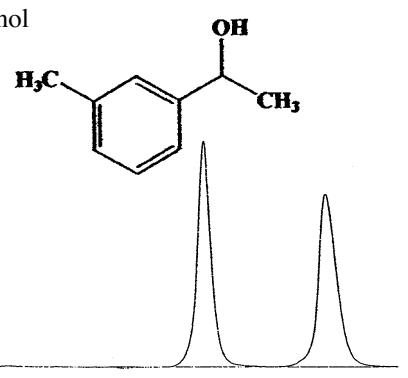
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 1.94$

$\alpha = 1.26$

reference 60



1-(o-Methylphenyl) Ethanol

1-(o-Methylphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-Dimethoxyethane

Flow Rate = 1.0 mL/min

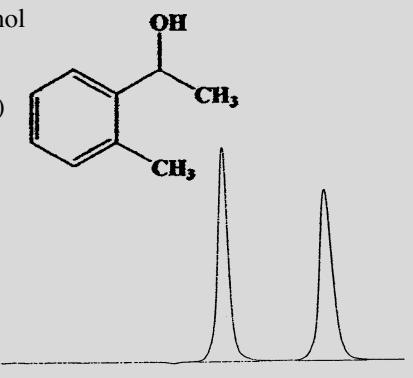
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 1.88$

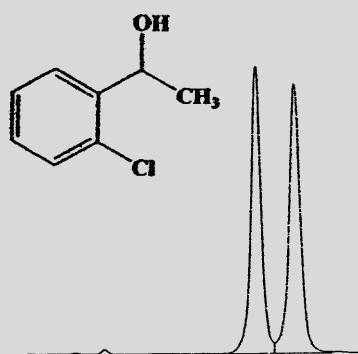
$\alpha = 1.29$

reference 60

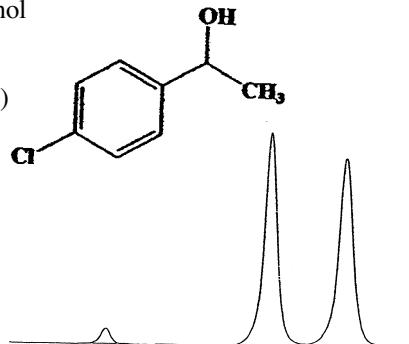


1-(o-Chlorophenyl) Ethanol

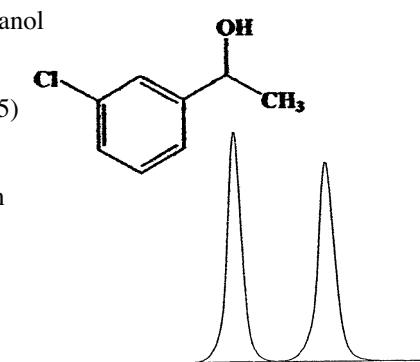
1-(o-Chlorophenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 8.5 min
 $k'_1 = 1.58$
 $\alpha = 1.12$
 reference 60

**1-(p-Chlorophenyl) Ethanol**

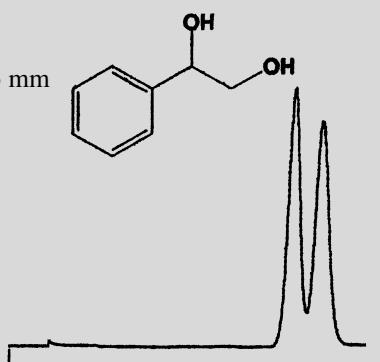
1-(p-Chlorophenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 2.18$
 $\alpha = 1.15$
 reference 60

**1-(m-Chlorophenyl) Ethanol**

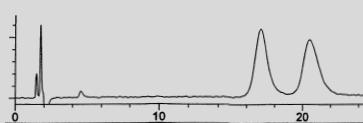
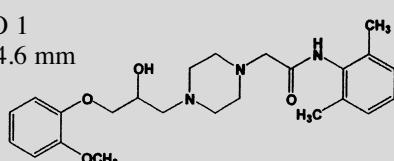
1-(m-Chlorophenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 2.13$
 $\alpha = 1.17$
 reference 60

**Phenylethylene Glycol**

Phenylethylene Glycol
 Column = (S,S)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/Ethanol
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 18.7 min
 $k'_1 = 11.62$
 $\alpha = 1.11$
 reference 46

**Ranolazine**

Ranolazine
 Column = (R,R)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (65/35)
 Hexane/IPA + 35 mM Ammonium Acetate
 Flow Rate = 2.0 mL/min
 Detection = UV 220 nm
 $k'_1 = 11.51$
 $\alpha = 1.23$
 reference 46



Hydrobenzoin

Hydrobenzoin

95:5 hexane/IPA

1 ml/min; 254 nm

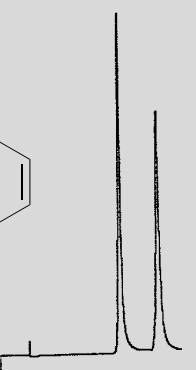
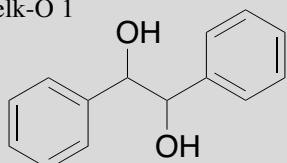
Run Time = 18 min

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 1.14$

$\alpha = 1.40$

reference 18



Benzoin

Benzoin

80:20:0.5 hexane/IPA/HOAc

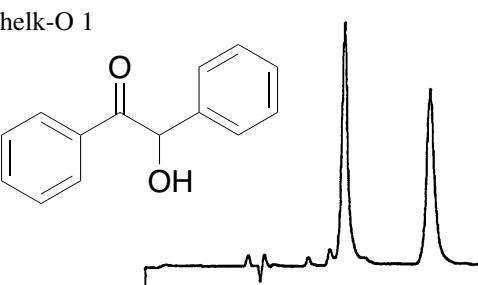
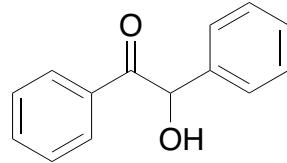
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 0.86$

$\alpha = 1.97$

reference 7



Anisoin

Anisoin

80:20:0.5

hexane/IPA/HOAc

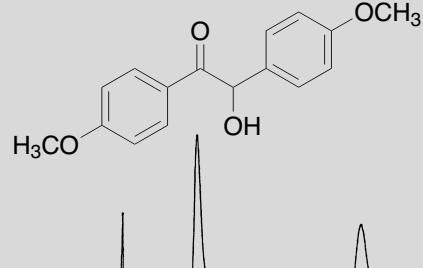
1 ml/min; 254 nm

4.6 mm x 25 cm
Whelk-O 1

$k'_1 = 3.07$

$\alpha = 2.34$

reference 26



Ipsdienol

Ipsdienol

2% IPA/hexane

1 ml/min; 254 nm

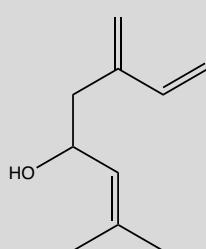
Run Time = 8 min

4.6 mm x 25 cm
Whelk-O 1

$k'_1 = 0.95$

$\alpha = 1.21$

reference 18



60:40 hexane/EtOH

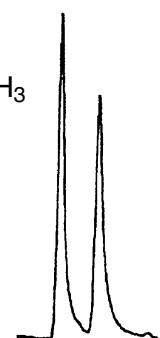
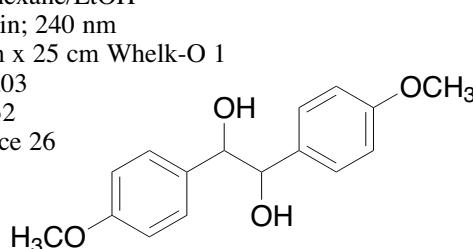
1 ml/min; 240 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 2.03$

$\alpha = 1.32$

reference 26



98:2:0.5 hexane/IPA/HOAc

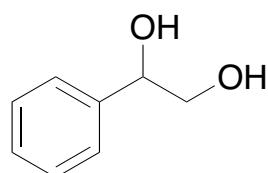
1 ml/min; 254 nm

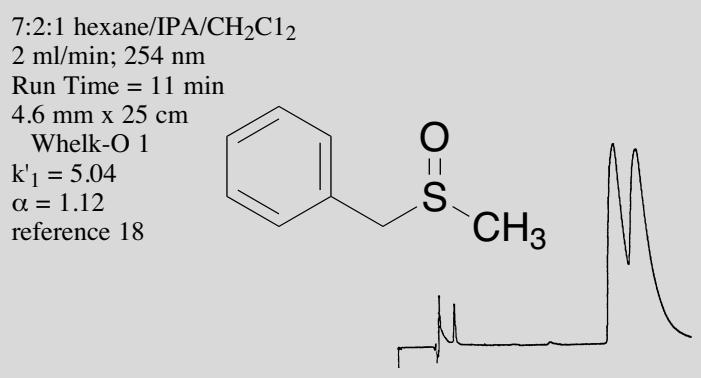
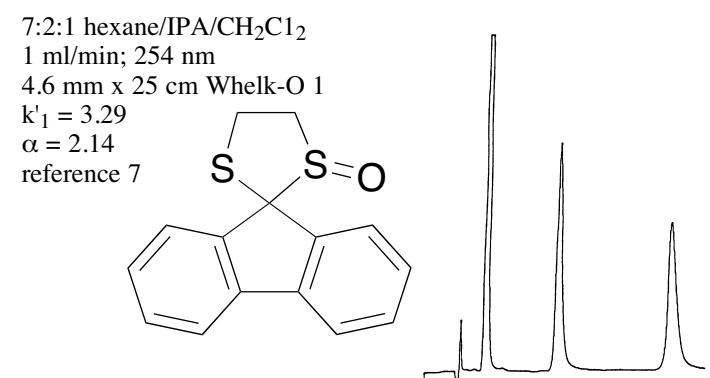
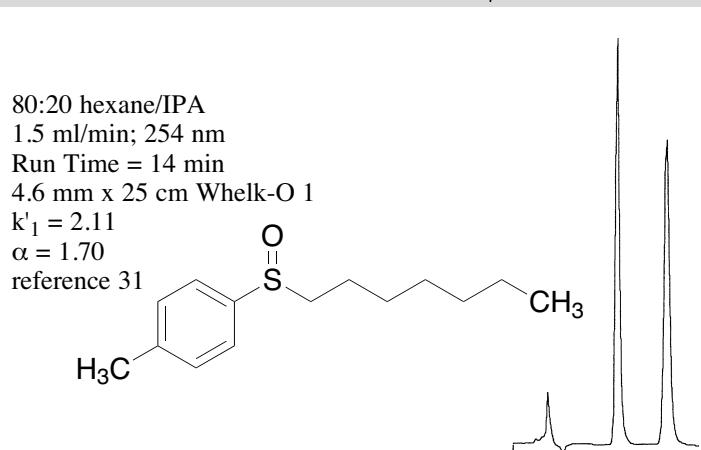
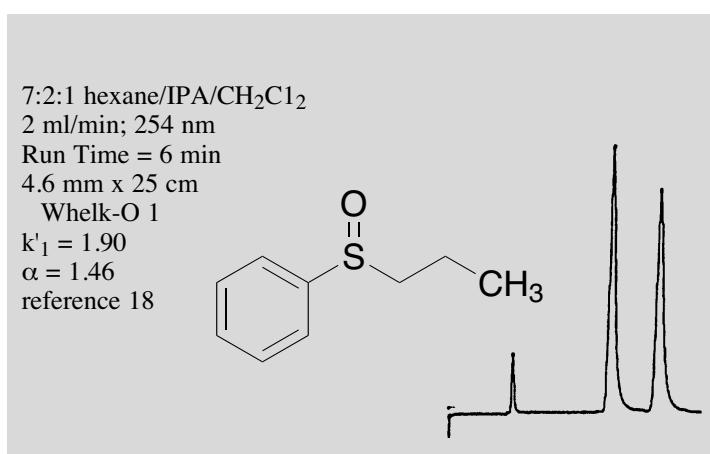
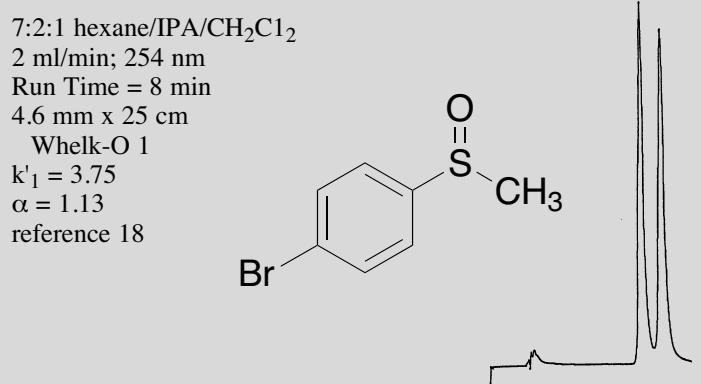
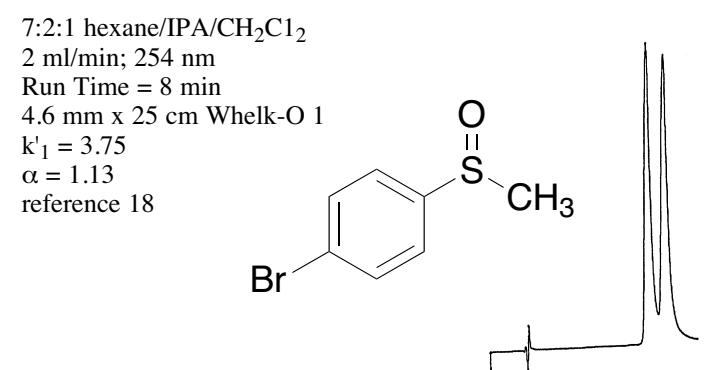
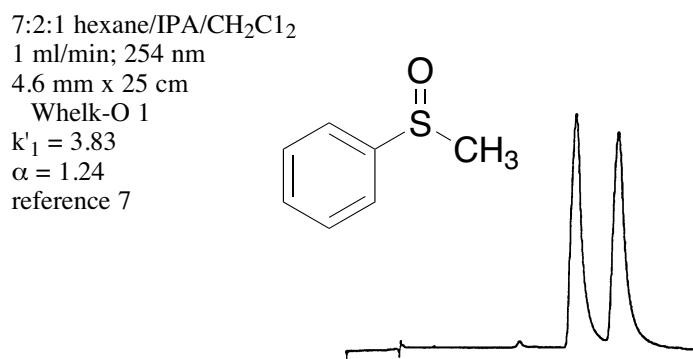
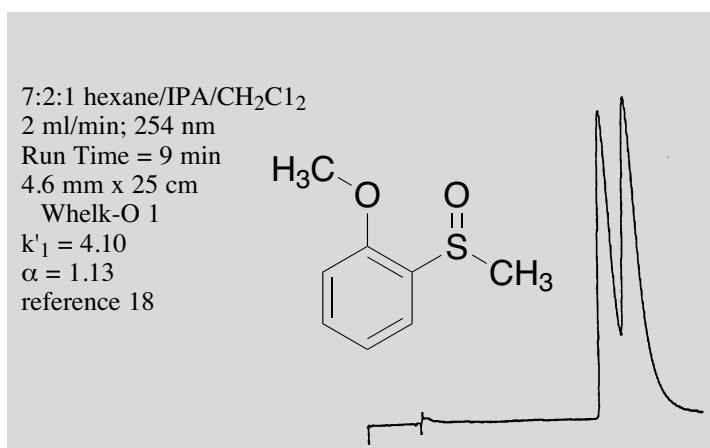
4.6 mm x 25 cm Whelk-O 1

$k'_1 = 7.54$

$\alpha = 1.08$

reference 7





REGIS Sulfoxides

7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

run time = 11 min

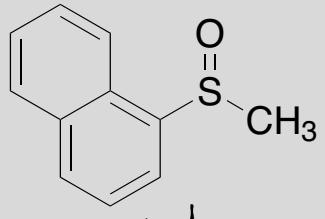
4.6 mm x 25 cm

Whelk-O 1

k' ₁ = 5.02

α = 1.21

reference 18



7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

run time = 6 min

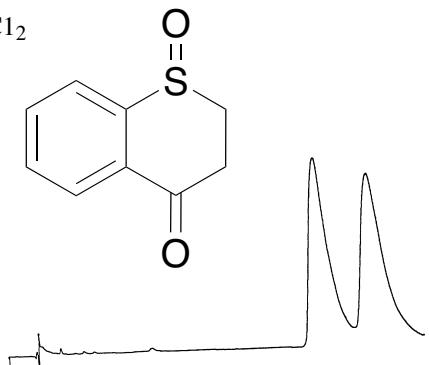
4.6 mm x 25 cm

Whelk-O 1

k' ₁ = 10.72

α = 1.19

reference 18



Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (27.5/27.5/45)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

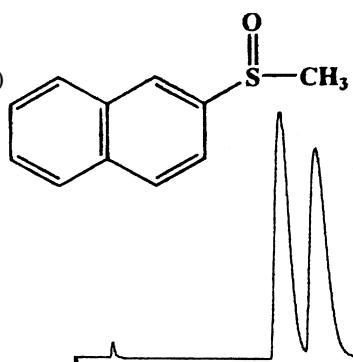
Detection: UV 254 nm

Run Time: 35.0 min

k' ₁: 10.30

α : 1.15

reference 59



Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (98/2)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

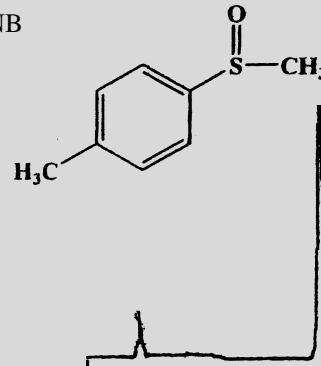
Detection: UV 254 nm

Run Time: 13.0 min

k' ₁: 3.08

α : 1.26

reference 59



Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:

(98/2)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

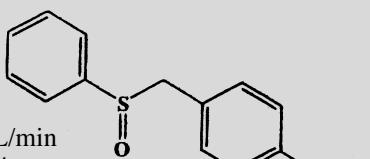
Detection: UV 254 nm

Run Time: 17.0 min

k' ₁: 3.33

α : 1.63

reference 59



Sulfinpyrazone

Sulfinpyrazone

Column = (R,R)-Whelk-O 1 25 cm x 4.6 mm

Mobile Phase = (75/25) Hexane/Ethanol

+ 15 mM Ammonium Acetate

Flow Rate = 1.5 mL/min

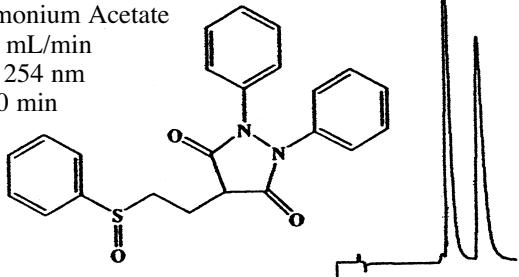
Detection = UV 254 nm

Run Time = 11.0 min

k' ₁ = 3.74

α = 1.35

reference 46



Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:

(98/2)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

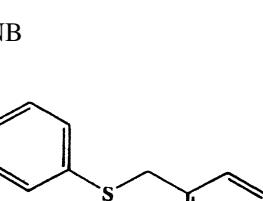
Detection: UV 254 nm

Run Time: 16.0 min

k' ₁: 2.34

α : 2.07

reference 59



Omeprazole

Omeprazole

Column = (S)- α -Burke 2 25 cm x 4.6 mm

Mobile Phase = (95/5) CH₂Cl₂/CH₃OH

Flow Rate = 1.0 mL/min

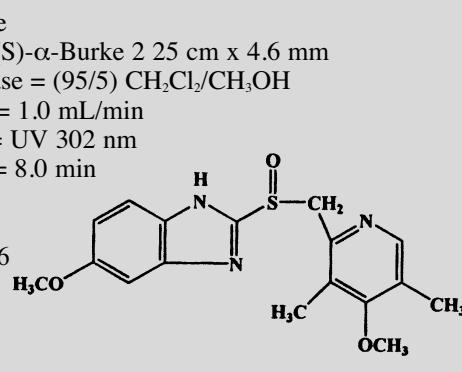
Detection = UV 302 nm

Run Time = 8.0 min

k' ₁ = 0.64

α = 3.04

reference 46



Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (40/40/20)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

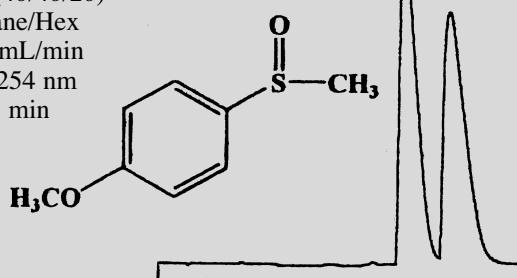
Detection: UV 254 nm

Run Time: 27.0 min

k'₁: 7.51

α : 1.21

reference 59



Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (27.5/27.5/45)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

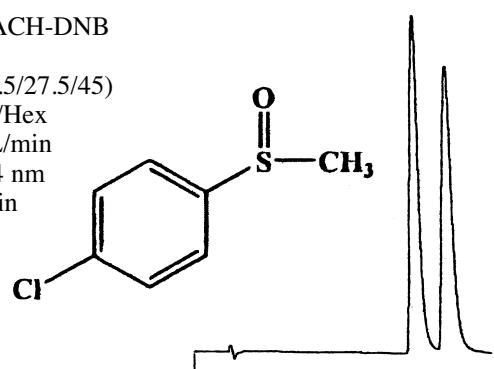
Detection: UV 254 nm

Run Time: 18.0 min

k'₁: 4.77

α : 1.18

reference 59

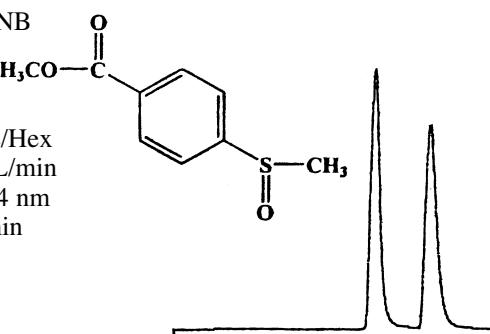


Column:

(R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:



(27.5/27.5/45)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

Detection: UV 254 nm

Run Time: 20.0 min

k'₁: 5.16

α : 1.26

reference 59

Column:

(R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

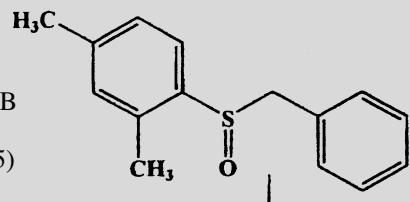
Detection: UV 254 nm

Run Time: 15.0 min

k'₁: 2.15

α : 2.05

reference 59



Pantoprazole

Pantoprazole

Column = (R)- α -Burke 2

25 cm x 4.6 mm

Mobile Phase = (48/48/4)

CH₂Cl₂/Hexane/Ethanol

+ 4 mM Ammonium Acetate

Flow Rate = 1.5 mL/min

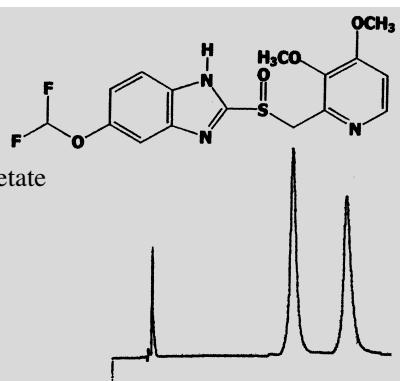
Detection = UV 280 nm

Run Time = 12.0 min

k'₁ = 4.07

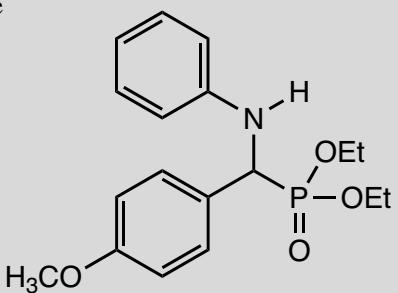
α = 1.38

reference 46

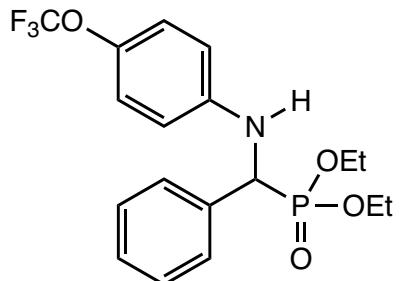


REGIS Phosphorous Compounds

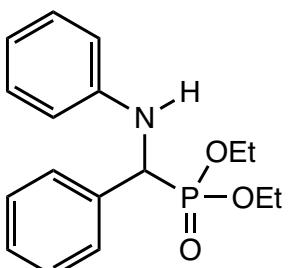
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.26$
 $\alpha = 1.50$
reference 40



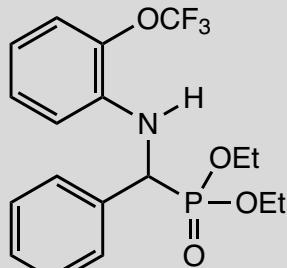
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.72$
 $\alpha = 1.26$
reference 40



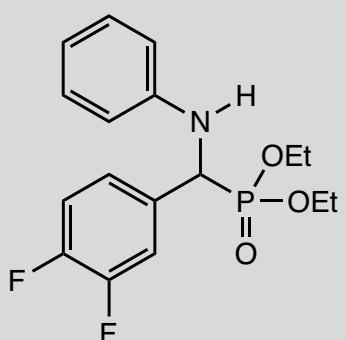
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.26$
 $\alpha = 1.26$
reference 40



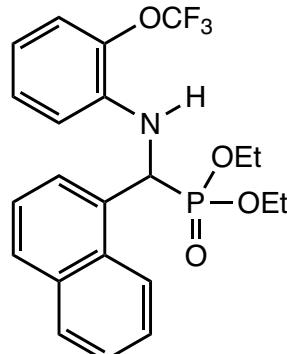
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.10$
 $\alpha = 2.08$
reference 40



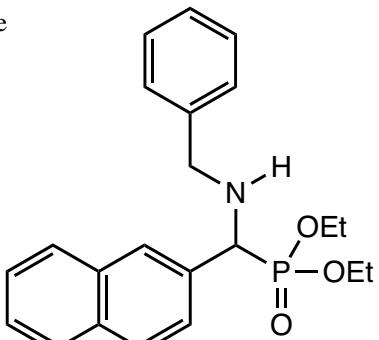
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.09$
 $\alpha = 1.31$
reference 40



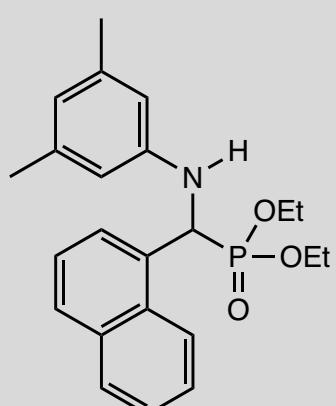
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.05$
 $\alpha = 1.63$
reference 40



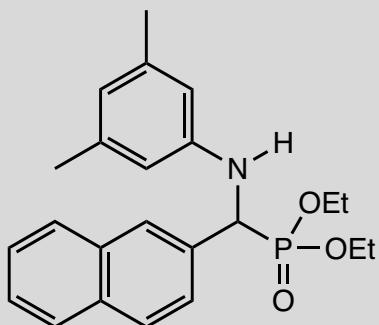
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 9.61$
 $\alpha = 1.75$
reference 40



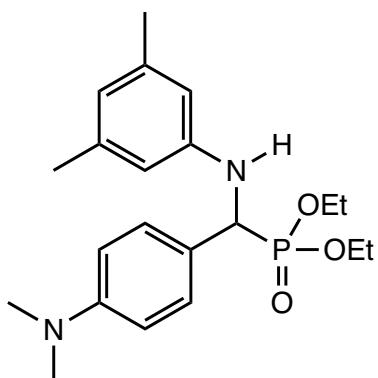
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.58$
 $\alpha = 1.23$
reference 40



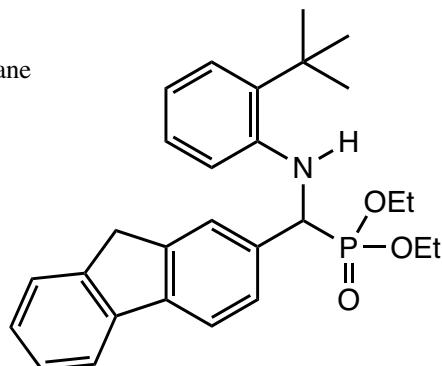
5% 2-propanol in hexane
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 7.35$
 $\alpha = 2.54$
reference 40



5% 2-propanol in hexane
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 12.30$
 $\alpha = 2.00$
reference 40

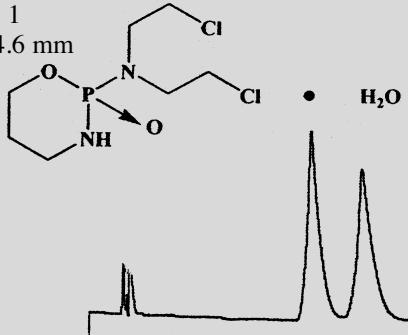


5% 2-propanol in hexane
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 5.87$
 $\alpha = 5.12$
reference 40



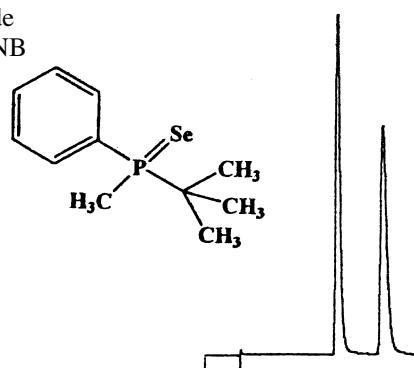
Cyclophosphamide

Cyclophosphamide
Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/Ethanol
Flow Rate = 1.5 mL/min
Detection = UV 195 nm
Run Time = 16.0 min
 $k'_1 = 6.31$
 $\alpha = 1.27$
reference 46

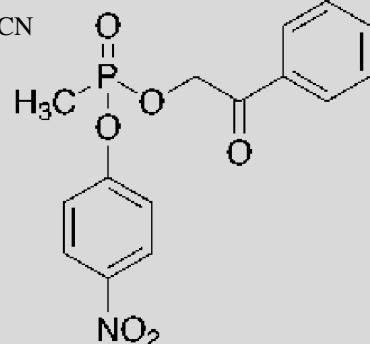


Phosphine Selenium Oxide

Phosphine Selenium Oxide
Column: (S,S)-DACH-DNB
25 cm x 4.6 mm
Mobile Phase: (70/30)
Hex/CH₂Cl₂
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 13.0 min
 $k'_1: 2.49$
 $\alpha: 1.48$
reference 59

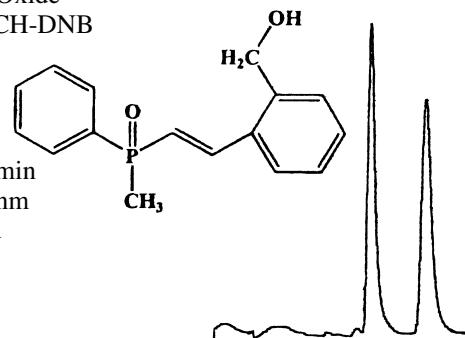


5:4:1 hexane/CH₂Cl₂/CH₃CN
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.11$
 $\alpha = 1.15$
reference 7



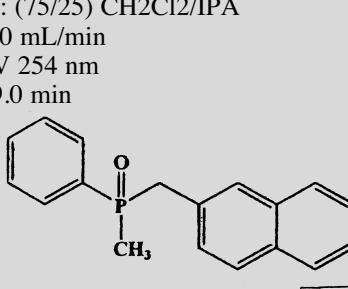
Tertiary Phosphine Oxide

Tertiary Phosphine Oxide
Column: (R,R)-DACH-DNB
25 cm x 4.6 mm
Mobile Phase:
(37.5/37.5/25)
Hex/Dioxane/IPA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 14.0 min
 $k'_1: 2.19$
 $\alpha: 1.48$
reference 59



Secondary Phosphine Oxide

Secondary Phosphine Oxide
Column: (S,S)-DACH-DNB 25 cm x 4.6 mm
Mobile Phase: (75/25) CH₂Cl₂/IPA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 19.0 min
 $k'_1: 1.49$
 $\alpha: 4.11$
reference 59



Secondary Phosphine Oxide

Secondary Phosphine Oxide

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (90/10)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

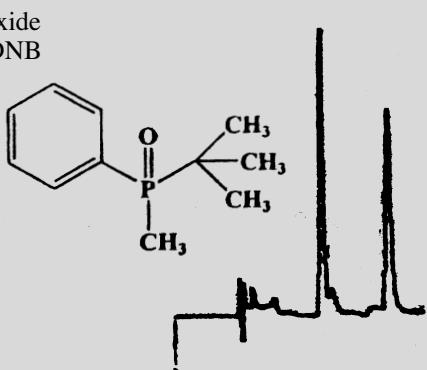
Detection: UV 254 nm

Run Time: 8.0 min

k'_1 : 1.23

α : 1.81

reference 59



Secondary Phosphine Oxide

Secondary Phosphine Oxide

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:

(90/10)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

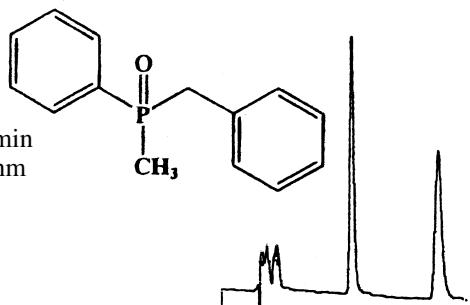
Detection: UV 254 nm

Run Time: 14.5 min

k'_1 : 2.20

α : 1.97

reference 59



Tertiary Phosphine Oxide

Tertiary Phosphine Oxide

Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:

(42.5/42.5/15)

Hex/Dioxane/IPA

Flow Rate: 1.0 mL/min

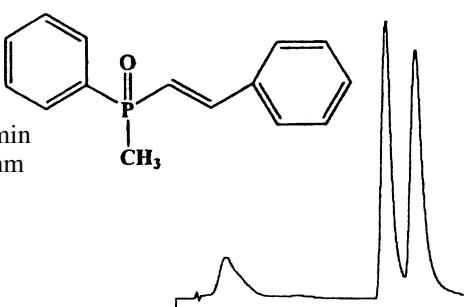
Detection: UV 254 nm

Run Time: 28.0 min

k'_1 : 8.11

α : 1.17

reference 59



Tertiary Phosphine Oxide

Tertiary Phosphine Oxide

Column:

(R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (40/40/20)

Hex/Dioxane/IPA

Flow Rate: 1.0 mL/min

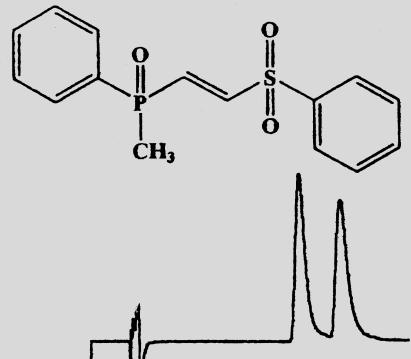
Detection: UV 254 nm

Run Time: 14.0 min

k'_1 : 4.19

α : 1.25

reference 59



10% IPA/hex

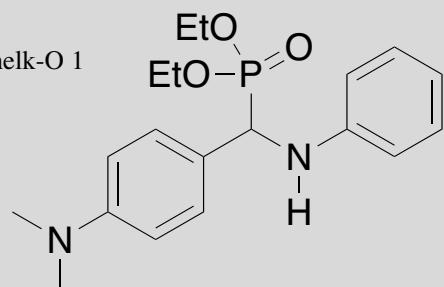
2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

k'_1 = 1.35

α = 3.53

reference 7



10% EtOH/hexane

1 ml/min; 254 nm

run time = 18 min

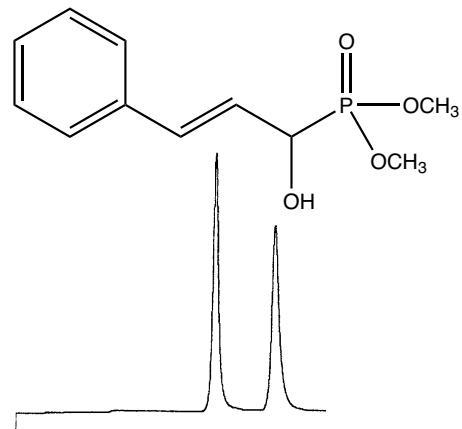
4.6 mm x 25 cm

Whelk-O 1

k'_1 = 3.75

α = 1.38

reference 18



10% EtOH/hexane

1 ml/min; 254 nm

run time = 13 min

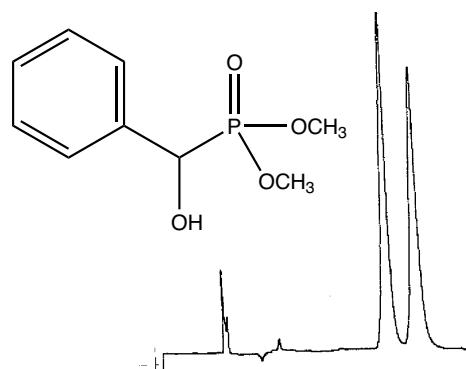
4.6 mm x 25 cm

Whelk-O 1

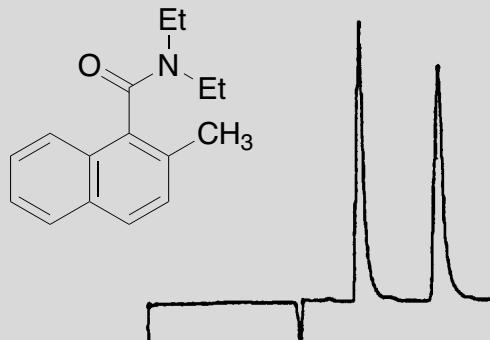
k'_1 = 3.07

α = 1.17

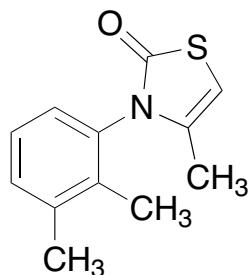
reference 18



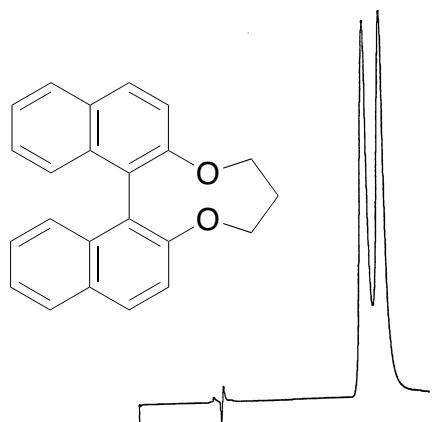
EtOAc
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.46$
 $\alpha = 2.17$
reference 7



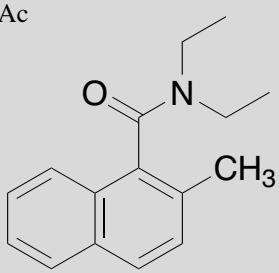
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.73$
 $\alpha = 1.64$
reference 7



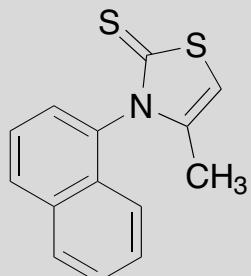
10% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.26$
 $\alpha = 1.11$
reference 7



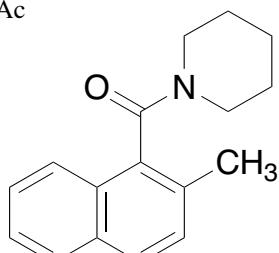
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.29$
 $\alpha = 2.46$
reference 10



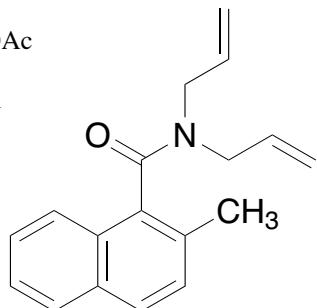
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.91$
 $\alpha = 2.13$
reference 7



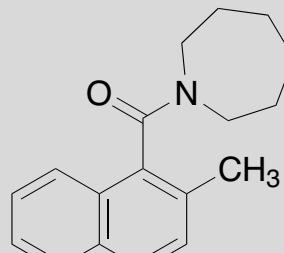
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O-1
 $k'_1 = 6.24$
 $\alpha = 2.63$
reference 10



80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O-1
 $k'_1 = 3.23$
 $\alpha = 2.66$
reference 10

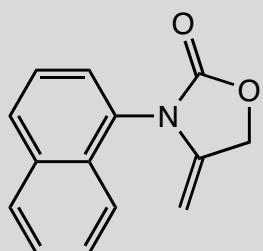


80:20:0.1%
hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O-1
 $k'_1 = 4.46$
 $\alpha = 2.08$
reference 10

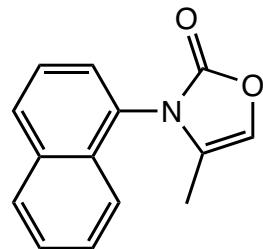


REGIS Atropisomers

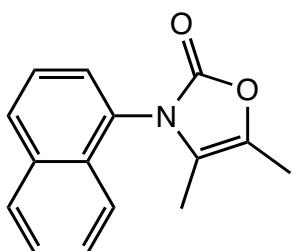
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 10.06$
 $\alpha = 2.37$
reference 41



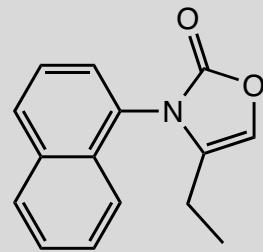
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 13.00$
 $\alpha = 2.57$
reference 41



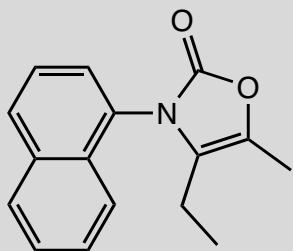
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 14.18$
 $\alpha = 2.78$
reference 41



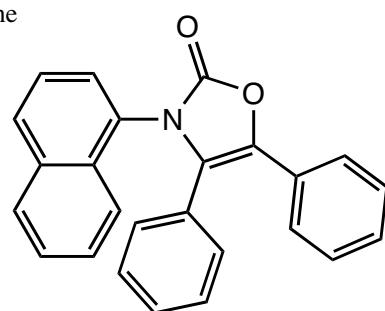
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 12.41$
 $\alpha = 2.74$
reference 41



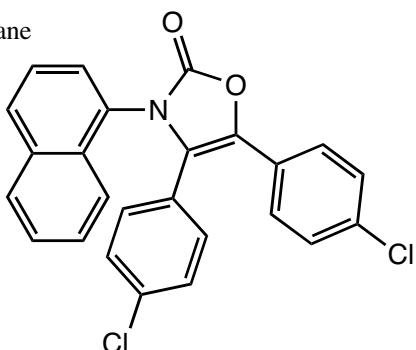
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 16.29$
 $\alpha = 3.15$
reference 41



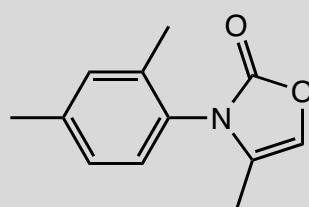
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 15.59$
 $\alpha = 3.74$
reference 41



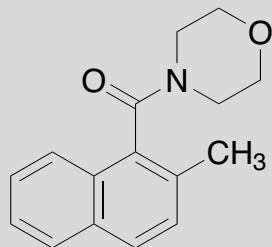
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.06$
 $\alpha = 2.22$
reference 41



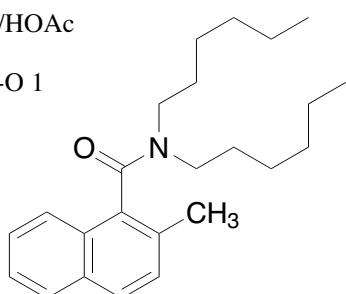
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.29$
 $\alpha = 2.25$
reference 41



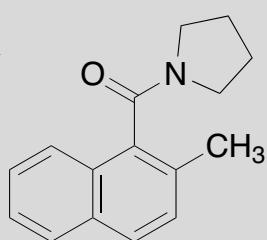
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 7.95$
 $\alpha = 2.43$
reference 10



80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.05$
 $\alpha = 3.00$
reference 10

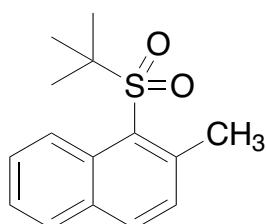


80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 5.11$
 $\alpha = 2.04$
reference 10

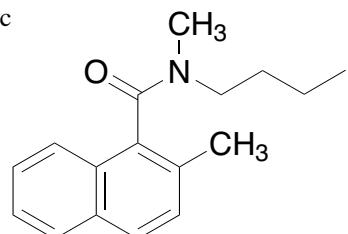


sulfone atropisomer

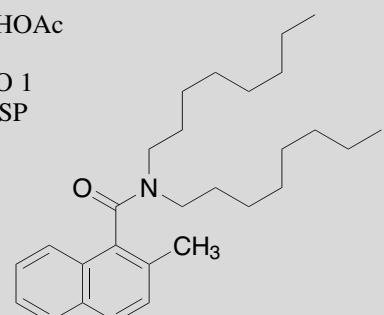
sulfone atropisomer
2% MeOH in CH_2Cl_2
2 ml/min; 300 nm, -80°C
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.54$
 $\alpha = 5.79$
reference 21



mixture of stereoisomers
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 10

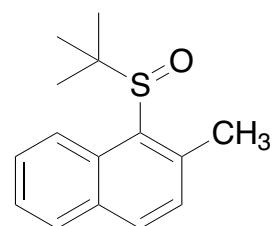


80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
(S)(-) retained on (S,S) CSP
 $k'_1 = 1.71$
 $\alpha = 3.09$
reference 10



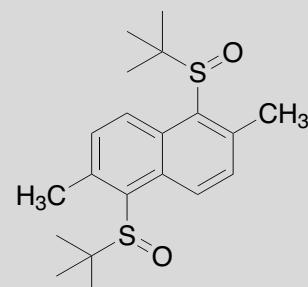
sulfoxide atropisomer

sulfoxide atropisomer
Z diastereomer
2% MeOH in CH_2Cl_2
2 ml/min; 300 nm, -40°C
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.32$
 $\alpha = 4.06$
reference 21



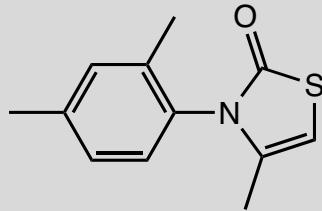
sulfone atropisomer

sulfone atropisomer
-80°C
reference 22

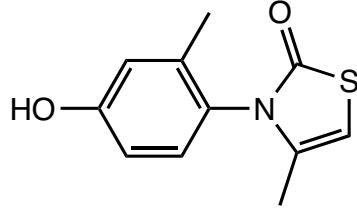


REGIS Atropisomers

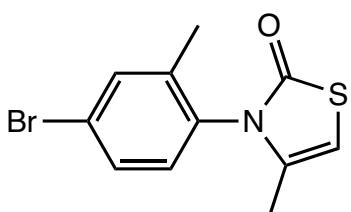
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.06$
 $\alpha = 2.48$
reference 41



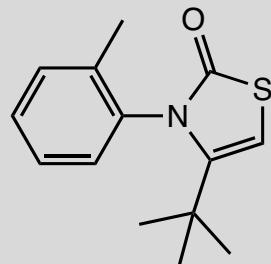
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.00$
 $\alpha = 3.43$
reference 41



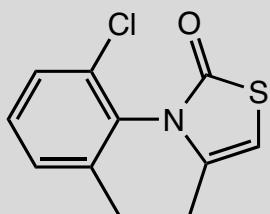
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.25$
 $\alpha = 3.20$
reference 41



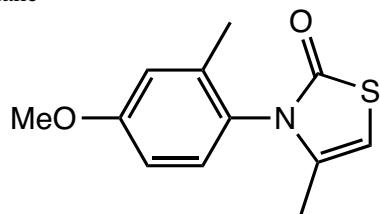
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.06$
 $\alpha = 4.34$
reference 41



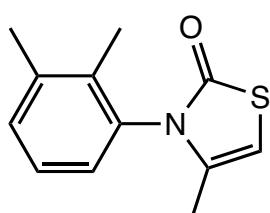
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.94$
 $\alpha = 1.12$
reference 41



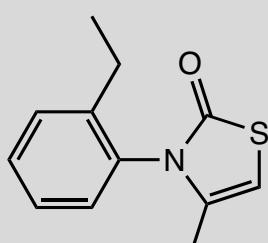
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.65$
 $\alpha = 3.63$
reference 41



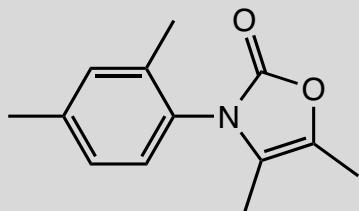
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 1.80$
reference 41



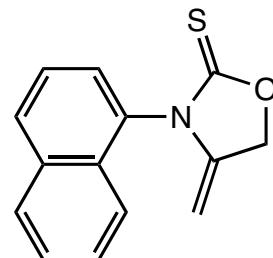
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.78$
 $\alpha = 1.90$
reference 41



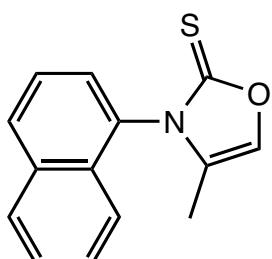
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.12$
 $\alpha = 2.40$
reference 41



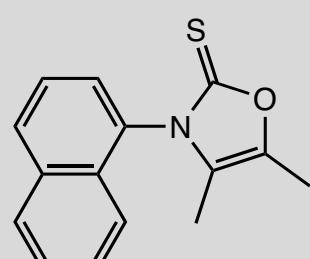
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O CSP
 $k'_1 = 6.94$
 $\alpha = 1.36$
reference 41



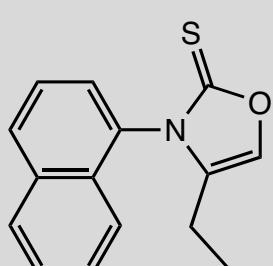
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.65$
 $\alpha = 1.35$
reference 41



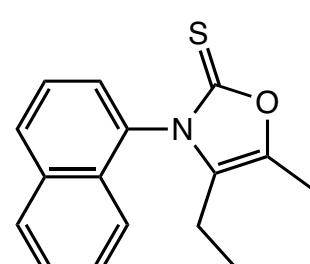
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.41$
 $\alpha = 1.48$
reference 41



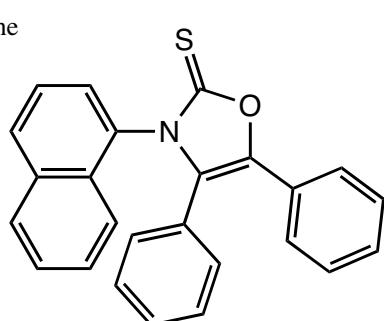
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.12$
 $\alpha = 1.36$
reference 41



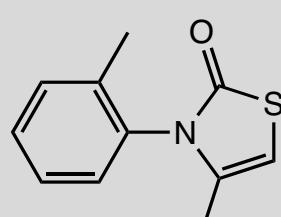
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.65$
 $\alpha = 1.50$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.65$
 $\alpha = 1.64$
reference 41

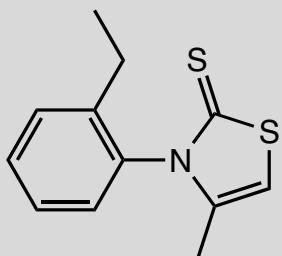


20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.06$
 $\alpha = 2.21$
reference 41

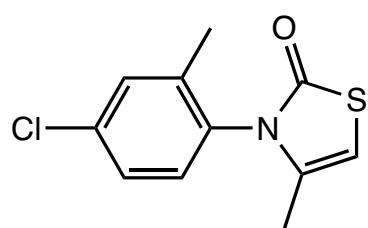


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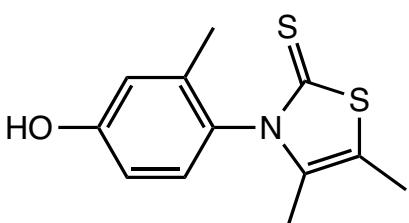
20% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 1.94$
 $\alpha = 1.85$
 reference 41



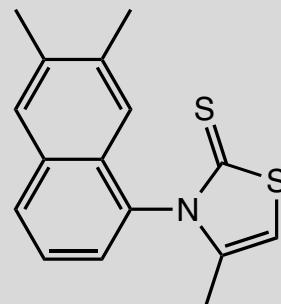
20% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 2.24$
 $\alpha = 1.45$
 reference 41



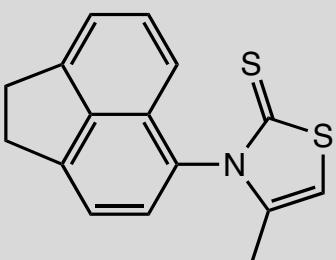
20% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 2.59$
 $\alpha = 1.70$
 reference 41



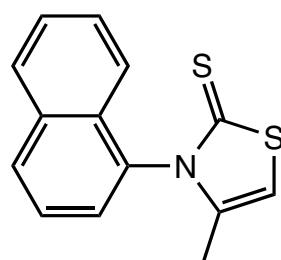
20% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 3.12$
 $\alpha = 2.39$
 reference 41



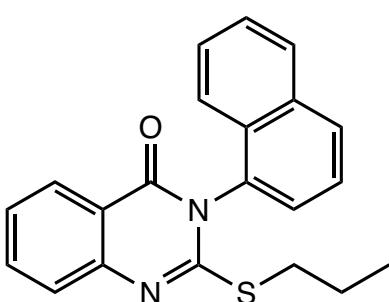
20% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 4.12$
 $\alpha = 1.62$
 reference 41



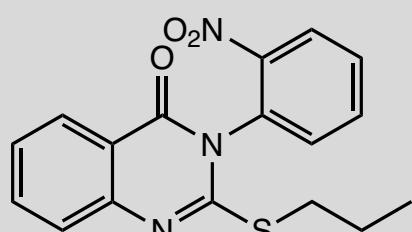
20% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 3.82$
 $\alpha = 1.62$
 reference 41



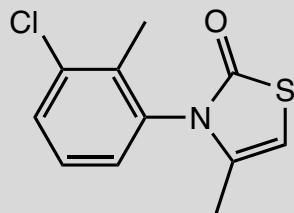
15% IPA/hexane
 1 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 2.14$
 $\alpha = 5.56$
 reference 42



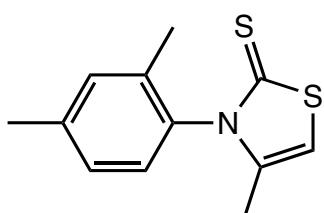
15% IPA/hexane
 1 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 2.86$
 $\alpha = 1.86$
 reference 42



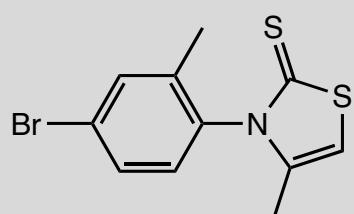
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.12$
 $\alpha = 2.21$
reference 41



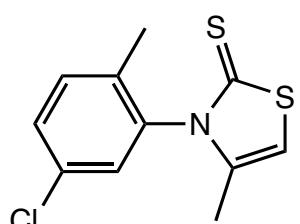
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.82$
 $\alpha = 1.75$
reference 41



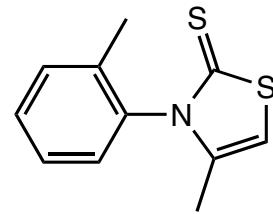
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.24$
 $\alpha = 2.02$
reference 41



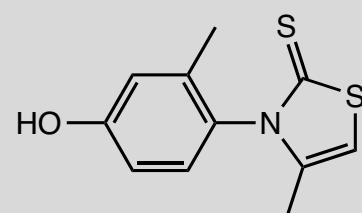
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.18$
 $\alpha = 1.46$
reference 41



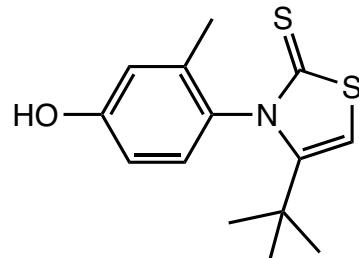
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 1.66$
reference 41



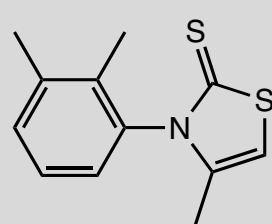
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 2.61$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.53$
 $\alpha = 1.84$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.29$
 $\alpha = 1.80$
reference 41

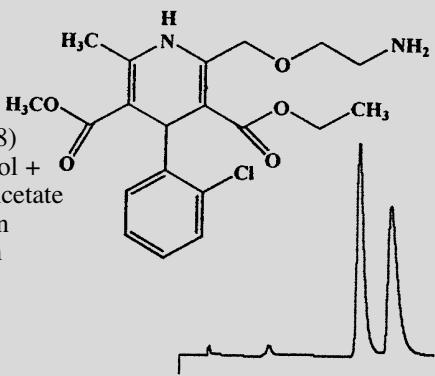


REGIS Atropisomers

Amlodipine

Amlodipine
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

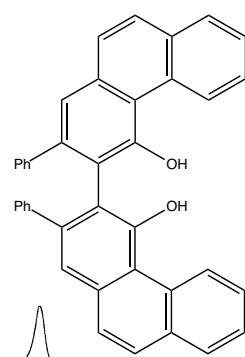
Mobile Phase = (46/46/8)
 $\text{CH}_2\text{Cl}_2/\text{Hexane/Ethanol} + 0.01 \text{ M Ammonium Acetate}$
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 13.0 min
 $k'_1 = 5.13$
 $\alpha = 1.22$
reference 46



Vapol

Vapol
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase =

100% Methanol
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 13 min
 $k'_1 = 1.74$
 $\alpha = 3.37$
reference 48

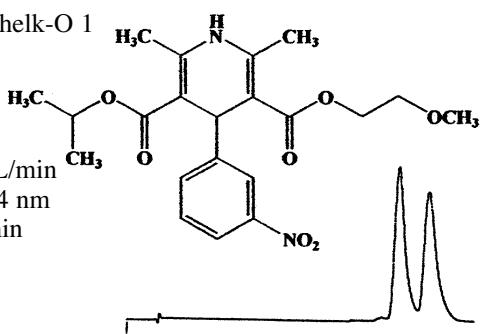


Nimodipine

Nimodipine

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase =
(65/35)
Methanol/H₂O
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 31.0 min
 $k'_1 = 9.25$
 $\alpha = 1.13$
reference 46

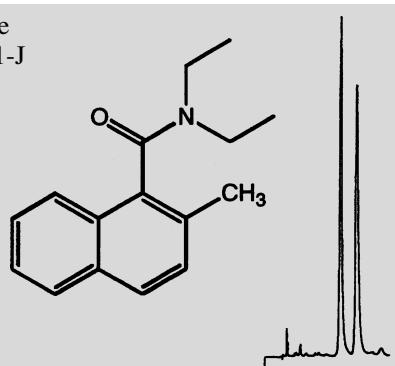


Adam's Acid Diethylamide

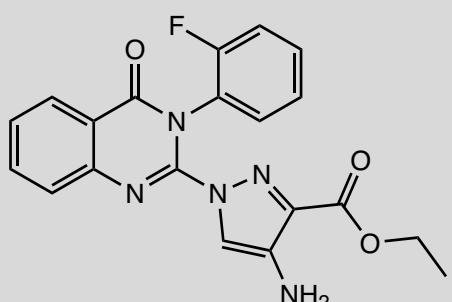
Adam's Acid Diethylamide

Column = (3R,4S)-Pirkle 1-J
25 cm x 4.6 mm

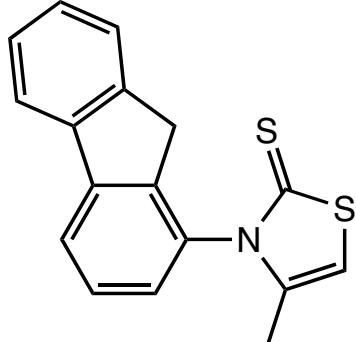
Mobile Phase = (70/30)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 17.0 min
 $k'_1 = 4.11$
 $\alpha = 1.27$
reference 46



15% IPA/hexane
1 mL/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 11.8$
 $\alpha = 1.58$
reference 42

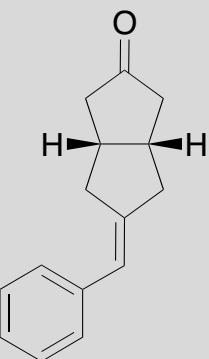


20% 2-propanol in hexane
2 mL/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 4.00$
 $\alpha = 2.25$
reference 41

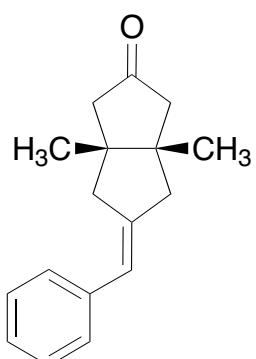


Phototrigger 1

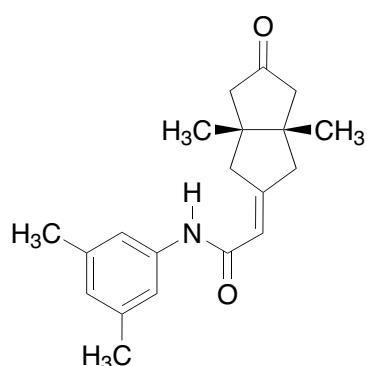
Phototrigger 1
8% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24

**Phototrigger 2**

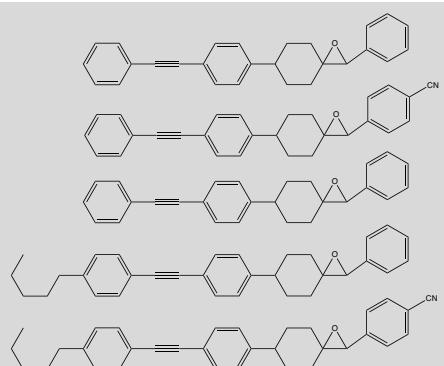
Phototrigger 2
8% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24

**Phototrigger 4**

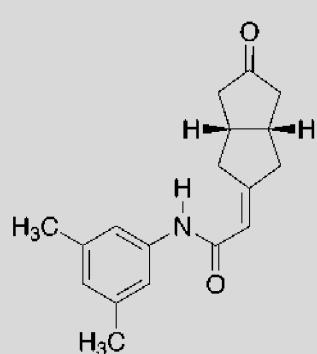
Phototrigger 4
40% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24

**Mesogens**

Mesogens
Schuster's candidate
photoresolvable
mesogensepoxyde
derivatives
reference 13

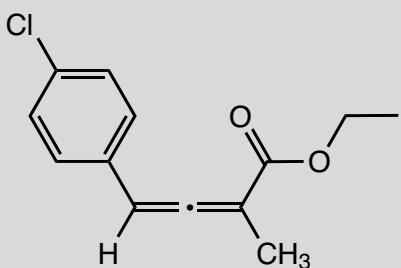
**Phototrigger 3**

Phototrigger 3
40% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24

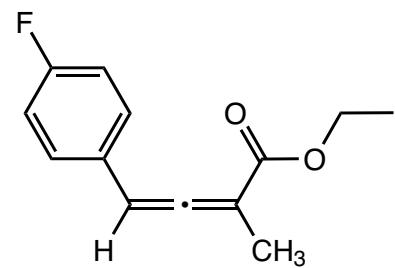


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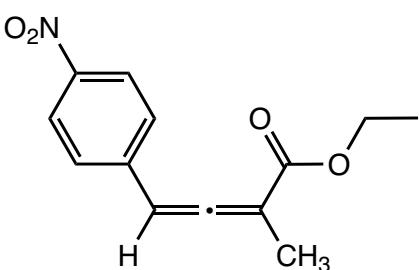
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.74$
 $\alpha = 2.38$
reference 42



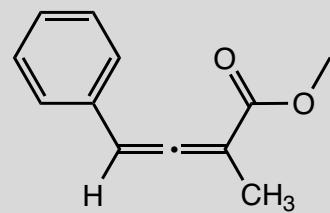
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.72$
 $\alpha = 2.33$
reference 42



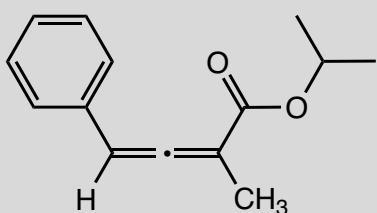
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.20$
 $\alpha = 1.59$
reference 42



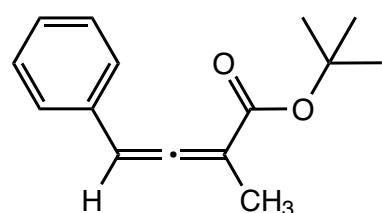
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.87$
 $\alpha = 2.90$
reference 42



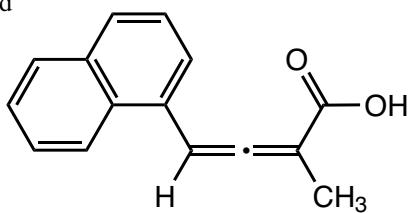
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.69$
 $\alpha = 3.70$
reference 42



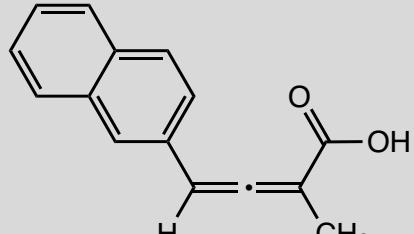
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.43$
 $\alpha = 3.23$
reference 42



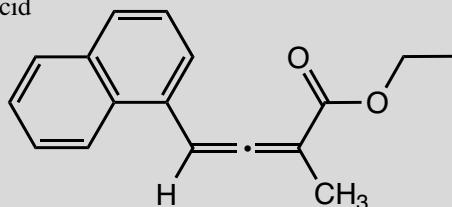
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.99$
 $\alpha = 7.49$
reference 42



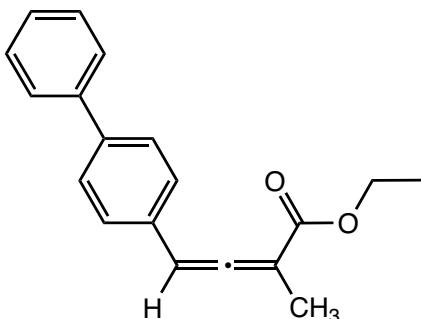
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.95$
 $\alpha = 4.19$
reference 42



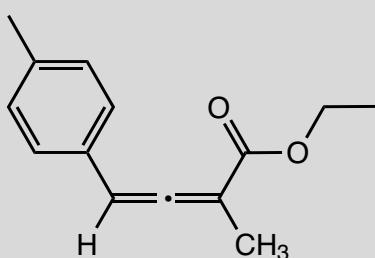
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 1.84$
 $\alpha = 5.68$
reference 42



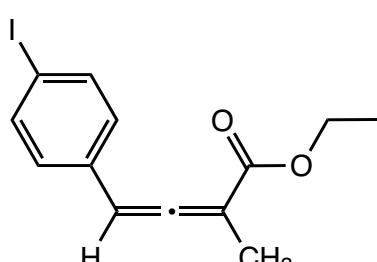
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 1.33$
 $\alpha = 3.13$
reference 42



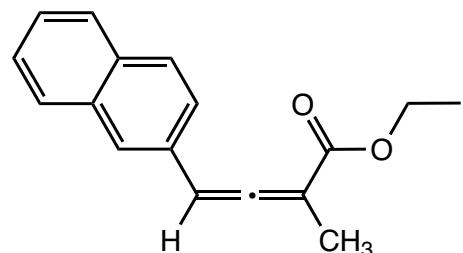
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 0.93$
 $\alpha = 3.85$
reference 42



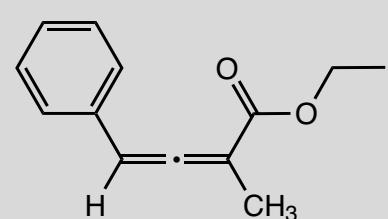
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 0.85$
 $\alpha = 2.48$
reference 42



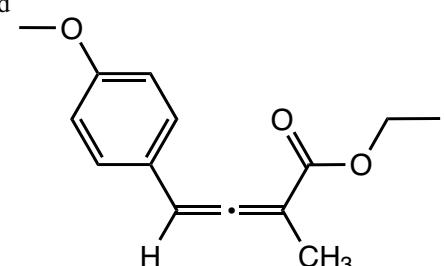
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 1.84$
 $\alpha = 3.46$
reference 42



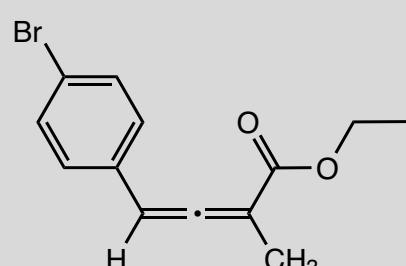
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 0.79$
 $\alpha = 3.23$
reference 42



95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 1.64$
 $\alpha = 3.29$
reference 42

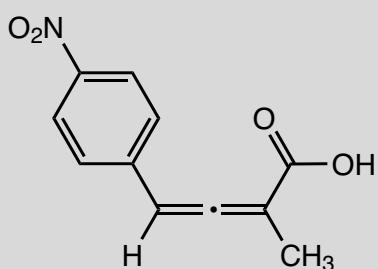


95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 0.79$
 $\alpha = 2.41$
reference 42

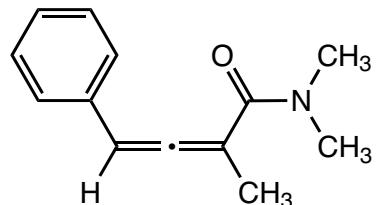


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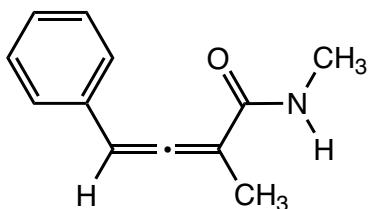
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 2.94$
 $\alpha = 1.74$
reference 42



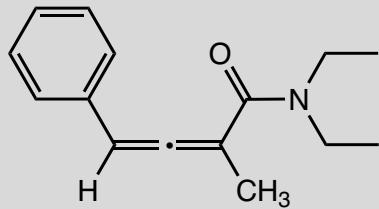
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 7.96$
 $\alpha = 1.03$
reference 42



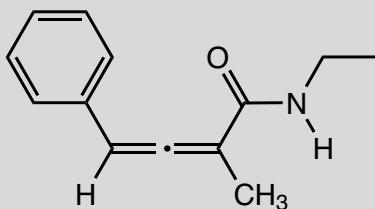
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 10.07$
 $\alpha = 1.45$
reference 42



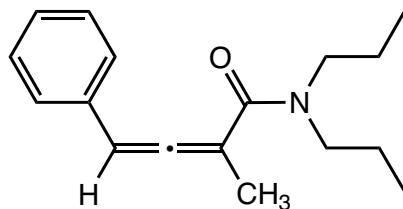
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 5.00$
 $\alpha = 1.14$
reference 42



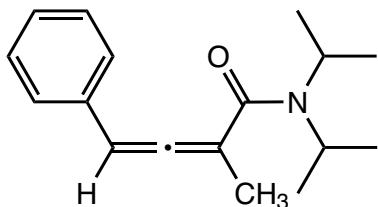
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 7.10$
 $\alpha = 1.44$
reference 42



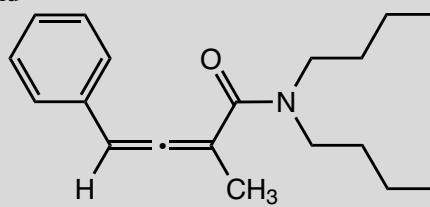
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 4.47$
 $\alpha = 0.09$
reference 42



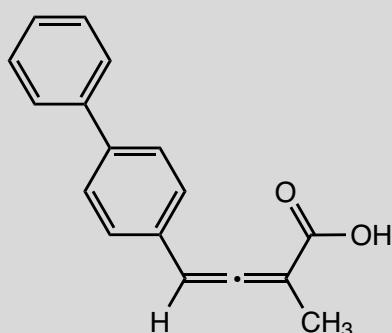
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 2.76$
 $\alpha = 1.13$
reference 42



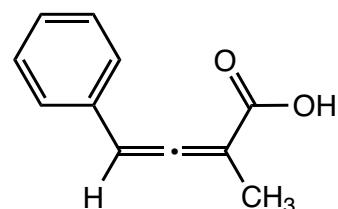
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 4.14$
 $\alpha = 1.08$
reference 42



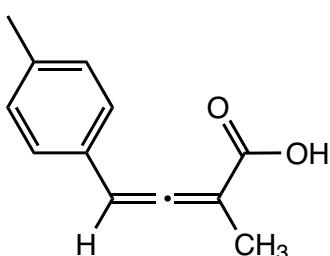
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.53$
 $\alpha = 3.56$
reference 42



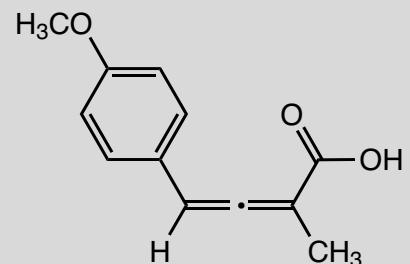
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.90$
 $\alpha = 3.92$
reference 42



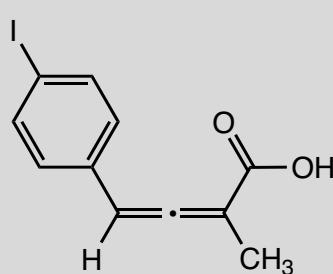
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.04$
 $\alpha = 4.28$
reference 42



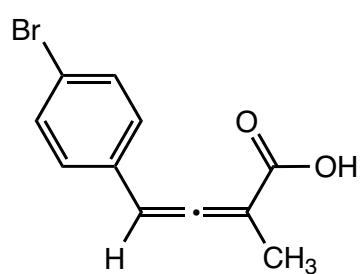
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.88$
 $\alpha = 3.62$
reference 42



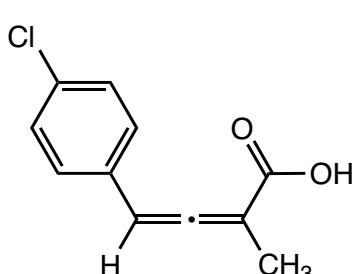
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.07$
 $\alpha = 2.84$
reference 42



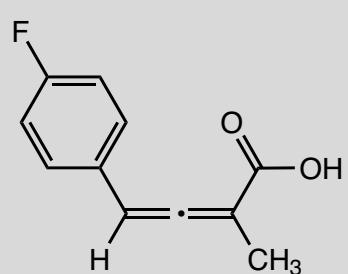
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.01$
 $\alpha = 2.67$
reference 42



$k'_1 = 0.92$
 $\alpha = 2.67$
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
reference 42

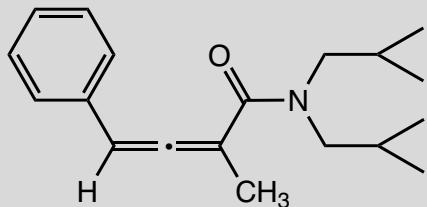


$k'_1 = 0.90$
 $\alpha = 2.57$
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
reference 42

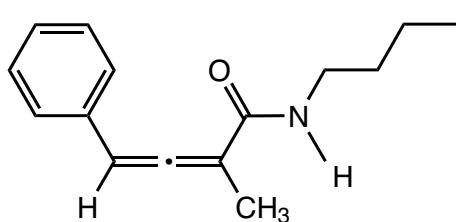


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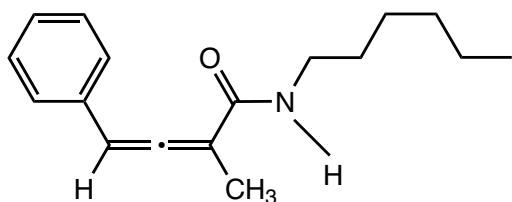
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 3.10$
 $\alpha = 1.18$
reference 42



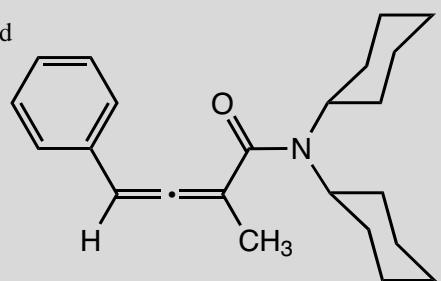
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 5.46$
 $\alpha = 1.34$
reference 42



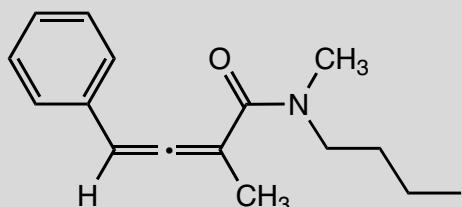
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 5.21$
 $\alpha = 1.33$
reference 42



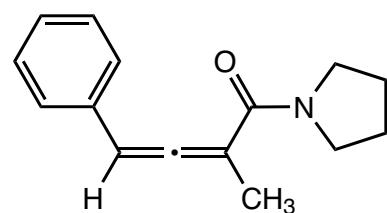
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 3.47$
 $\alpha = 1.14$
reference 42



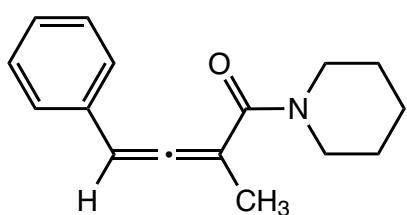
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 5.30$
 $\alpha = 1.06$
reference 42



95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 10.30$
 $\alpha = 1.11$
reference 42



95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(S,S) Whelk-O 1
 $k'_1 = 6.07$
 $\alpha = 1.12$
reference 42



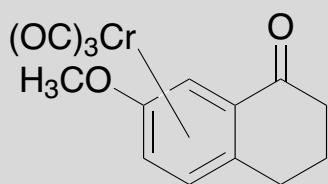
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 3.82$ $\alpha = 1.07$

reference 20



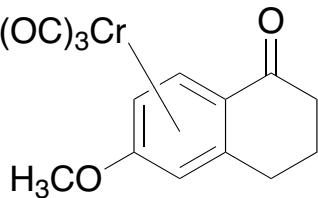
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 5.93$ $\alpha = 1.18$

reference 20



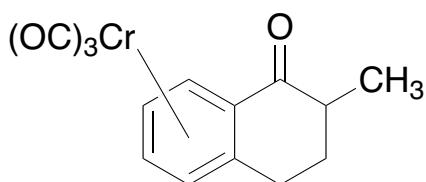
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 2.25$ $\alpha = 1.19$

reference 20



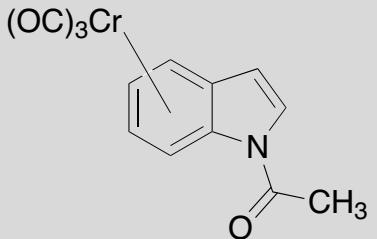
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 6.79$ $\alpha = 1.04$

reference 20



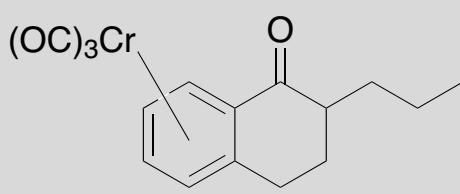
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 1.48$ $\alpha = 1.23$

reference 20



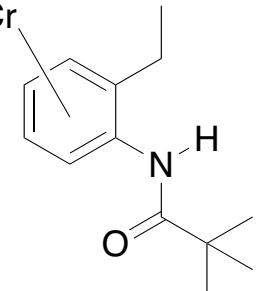
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 1.71$ $\alpha = 1.75$

reference 20



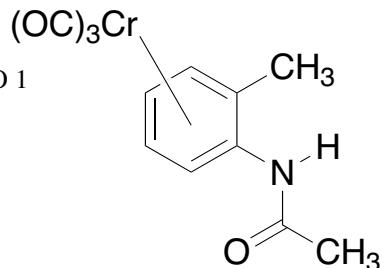
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 4.93$ $\alpha = 1.62$

reference 20



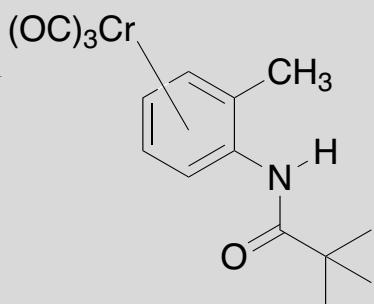
20% IPA in hexane

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

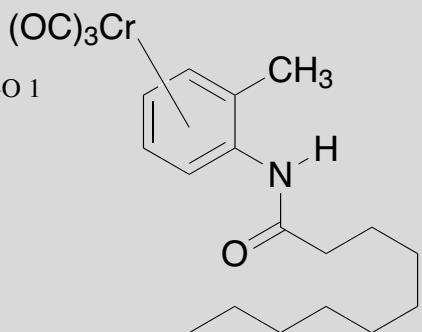
 $k'_1 = 2.44$ $\alpha = 1.75$

reference 20

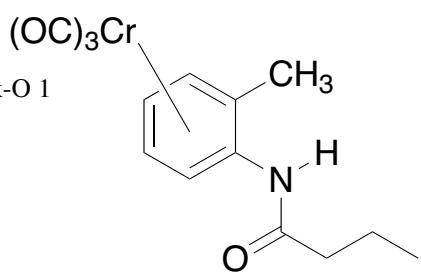


REGIS Organometallic Compounds

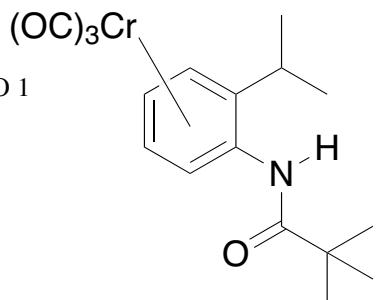
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.79$
 $\alpha = 1.99$
reference 20



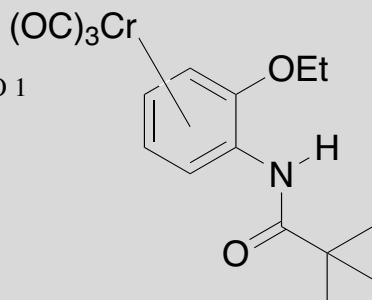
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.14$
 $\alpha = 1.86$
reference 20



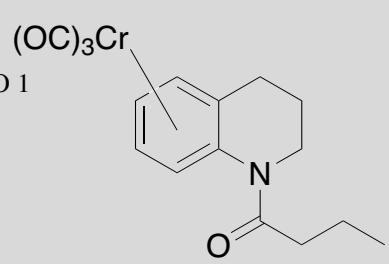
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.14$
 $\alpha = 1.75$
reference 20



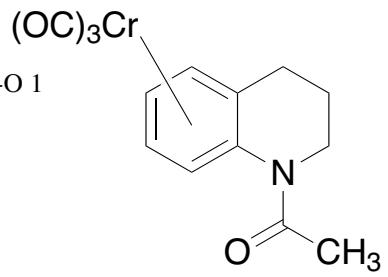
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.86$
 $\alpha = 1.69$
reference 20



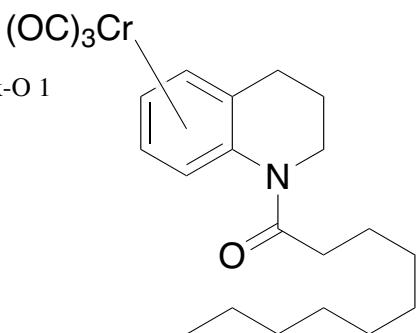
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.71$
 $\alpha = 2.50$
reference 20



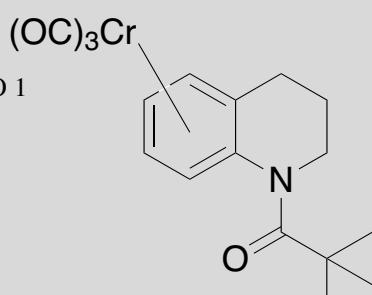
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 11.86$
 $\alpha = 2.08$
reference 20



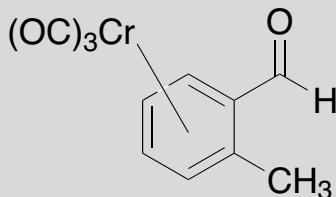
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.71$
 $\alpha = 2.50$
reference 20



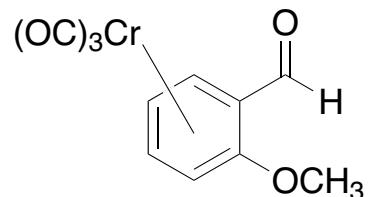
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.29$
 $\alpha = 2.46$
reference 20



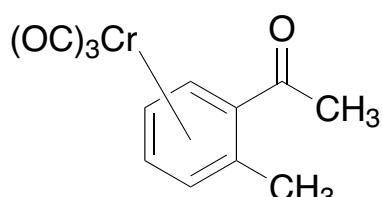
30% CH₂Cl₂ in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.28$
 $\alpha = 1.07$
reference 20



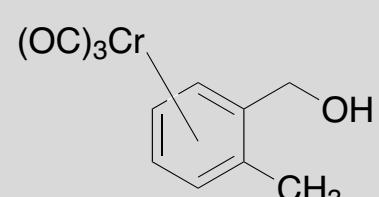
30% CH₂Cl₂ in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 7.57$
 $\alpha = 1.09$
reference 20



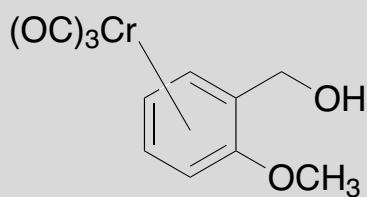
30% CH₂Cl₂ in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.57$
 $\alpha = 1.06$
reference 20



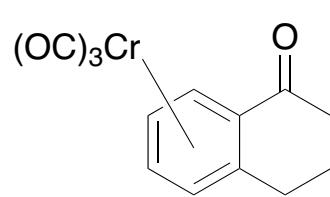
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.77$
 $\alpha = 1.11$
reference 20



20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.22$
 $\alpha = 1.15$
reference 20

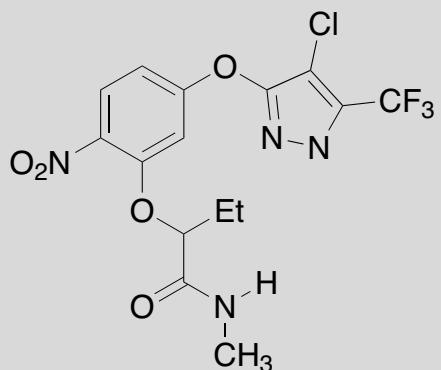


20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.48$
 $\alpha = 1.08$
reference 20



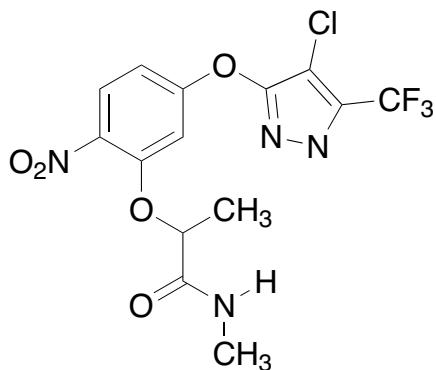
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 5.2$
 $\alpha = 1.32$
reference 23



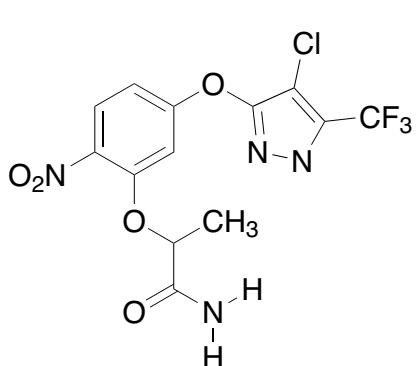
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 7.5$
 $\alpha = 1.29$
reference 23



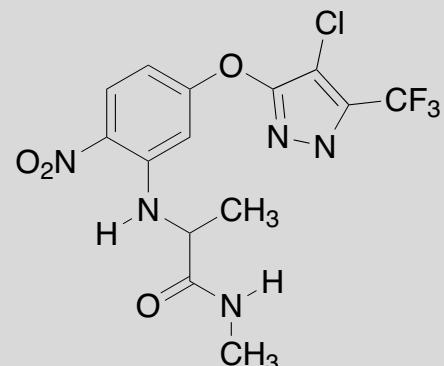
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 8.0$
 $\alpha = 1.22$
reference 23



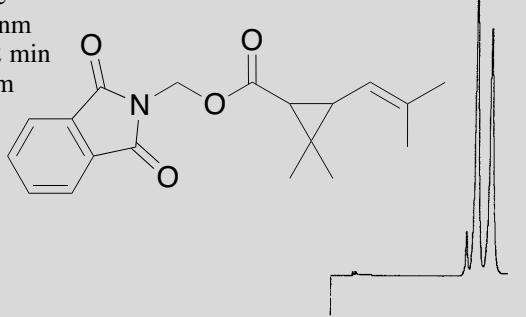
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 15.1$
 $\alpha = 1.04$
reference 23



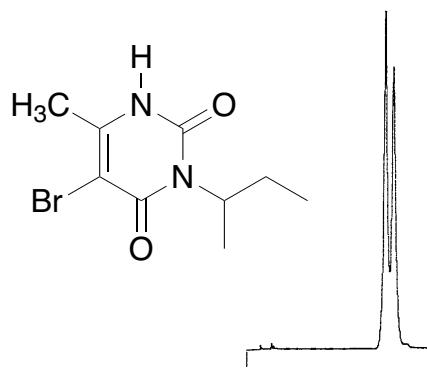
Tetramethrin

Tetramethrin (insecticide)
2% IPA/hexane
1 ml/min; 254 nm
Run Time = 22 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 11.77$
 $\alpha = 1.12$
reference 43



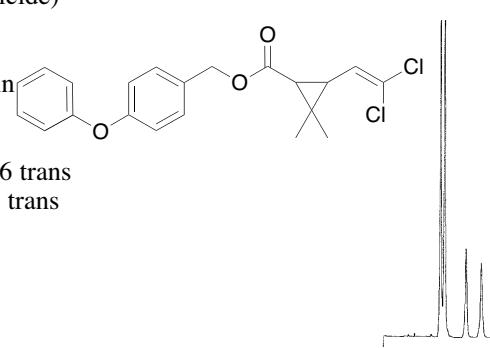
Bromacil

Bromacil (insecticide)
2% IPA/hexane
1 ml/min; 254 nm
Run Time = 38 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 21.43$
 $\alpha = 1.07$
reference 43



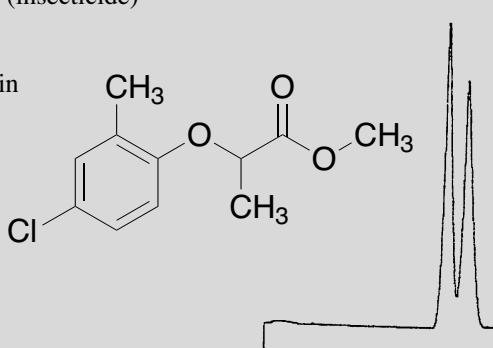
Permethrin

Permethrin (insecticide)
0.2% IPA/hexane
1 ml/min; 254 nm
Run Time = 16 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 4.83$ cis; 7.46 trans
 $\alpha = 1.11$ cis; 1.24 trans
reference 43



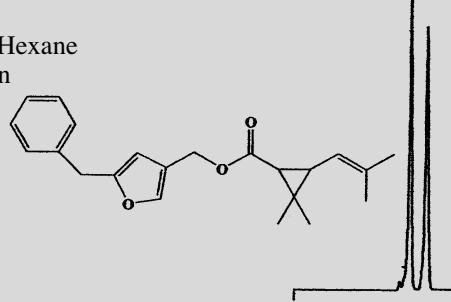
Mecoprop Methyl

Mecoprop Methyl (insecticide)
hexane
1 ml/min; 254 nm
Run Time = 15 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 6.92$
 $\alpha = 1.15$
reference 43

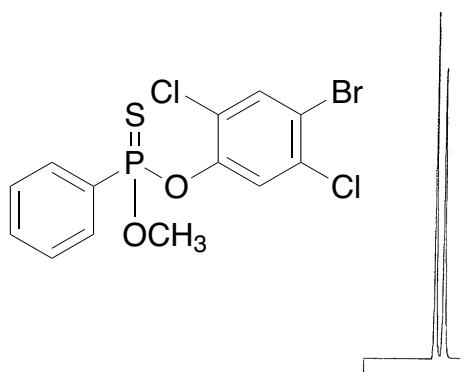


Resmethrin

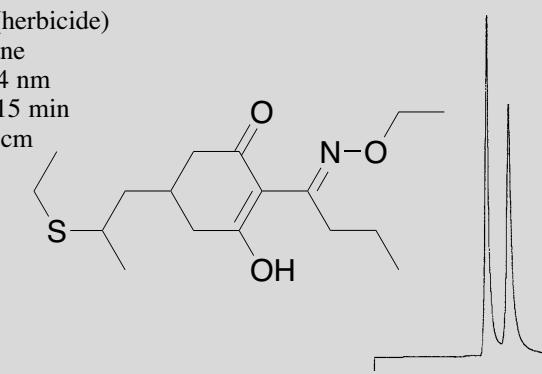
Resmethrin
 Column: (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase: 100% Hexane
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 15.0 min
 $k'_1 = 6.30$
 $\alpha = 1.19$
 reference 46

**Leptophos, Phosvel**

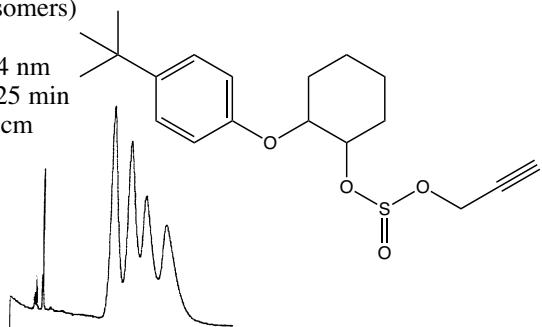
Leptophos, Phosvel
 (insecticide)
 hexane
 1 ml/min; 254 nm
 Run Time = 10 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 4.11$
 $\alpha = 1.18$
 reference 43

**Sethoxydim**

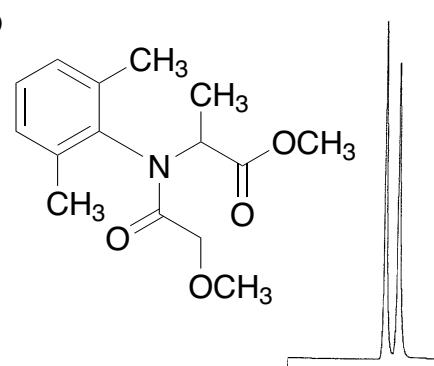
Sethoxydim (herbicide)
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 15 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.77$
 $\alpha = 1.26$
 reference 43

**Omite**

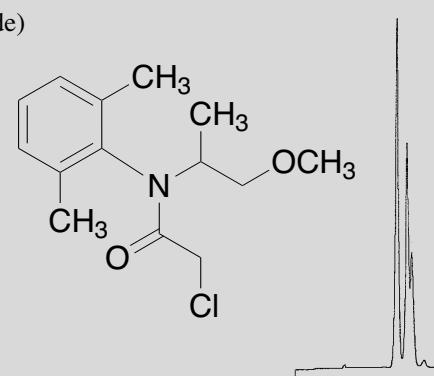
Omite (acaricide)
 (mixture of isomers)
 hexane
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 24 cm
 Whelk-O 1
 reference 43

**Metalaxyl**

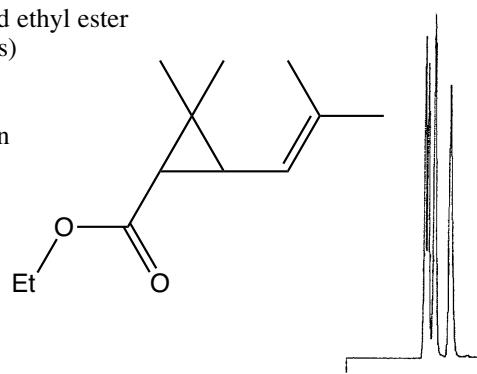
Metalaxyl (herbicide)
 70:30 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 13 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.54$
 $\alpha = 1.13$
 reference 43

**Metolachlor**

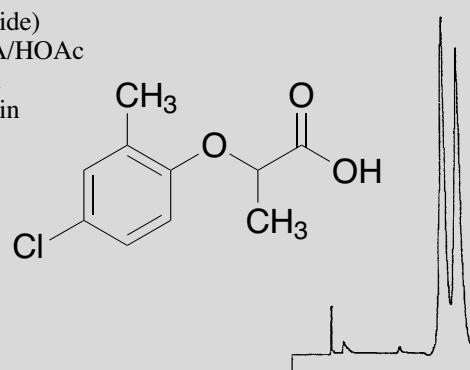
Metolachlor (herbicide)
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 25 cm
 Whelk-O 1
 reference 43

**Chrysanthemic Acid-Ethyl Ester**

Chrysanthemic acid ethyl ester
 (mixture of isomers)
 hexane
 1 ml/min; 254 nm
 Run Time = 10 min
 4.6 mm x 24 cm
 Whelk-O 1
 reference 43

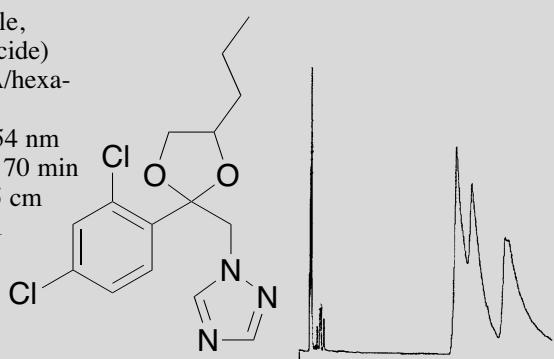
**Mecoprop**

Mecoprop (herbicide)
 99:1:0.1 HEX/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 15 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.54$
 $\alpha = 1.13$
 reference 43



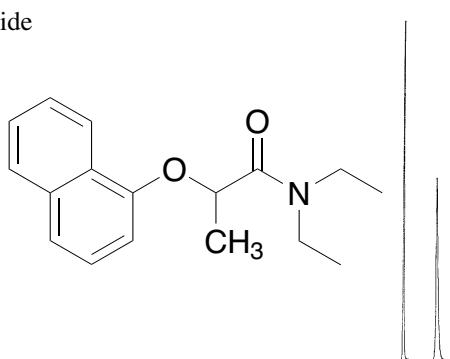
Propiconazole, Tilt

Propiconazole,
Tilt (fungicide)
99:1:0.1 IPA/hexane/HOAc
1 ml/min; 254 nm
Run Time = 70 min
4.6 mm x 25 cm
Whelk-O 1
reference 43



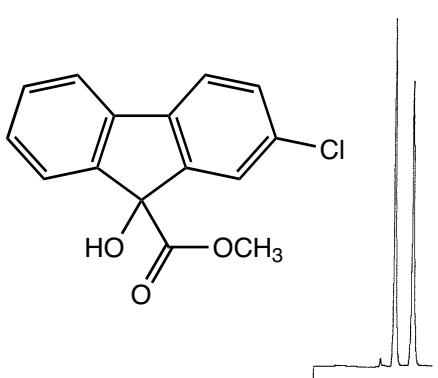
Devrinol, Napropamide

Devrinol, Napropamide
(herbicide)
1:1 IPA/hexane
1 ml/min; 254 nm
Run Time = 15 min
4.6 mm x 25 cm
Whelk-O 1
 k' = 3.17
 α = 3.00
reference 43



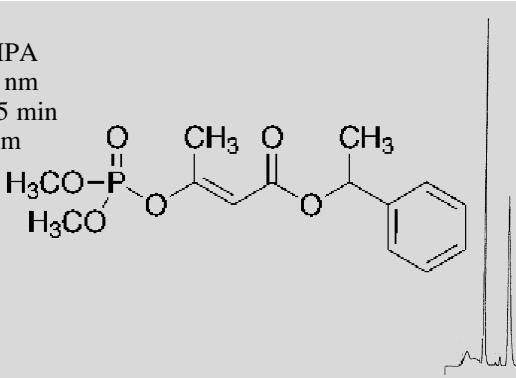
Chlorflurecol Methyl

Chlorflurecol Methyl
(herbicide)
2% IPA/hexane
1 ml/min; 254 nm
Run Time = 16 min
4.6 mm x 25 cm
Whelk-O 1
 k' = 3.96
 α = 1.28
reference 43



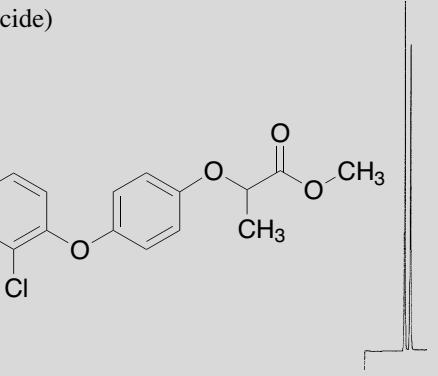
Crotoxyphos

Crotoxyphos
70:30 hexane/IPA
1 ml/min; 254 nm
Run Time = 15 min
4.6 mm x 25 cm
Whelk-O 1
 k' = 4.37
 α = 1.93
reference 43



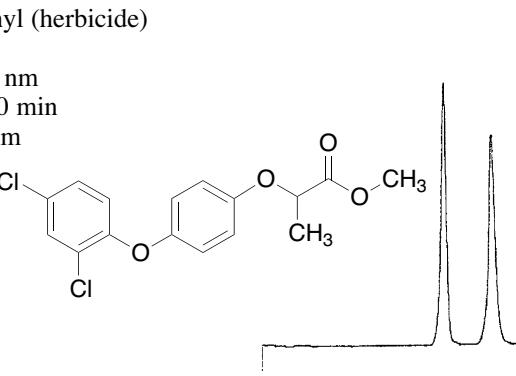
Diclofop Methyl

Diclofop Methyl (herbicide)
1% IPA/hexane
1 ml/min; 254 nm
Run Time = 30 min
4.6 mm x 25 cm
Whelk-O 1
 k' = 4.29
 α = 1.21
reference 43



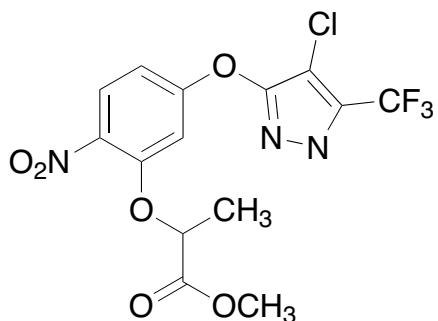
Diclofop Methyl

Diclofop Methyl (herbicide)
hexane
1 ml/min; 254 nm
Run Time = 30 min
4.6 mm x 25 cm
Whelk-O 1
 k' = 14.19
 α = 1.30
reference 43



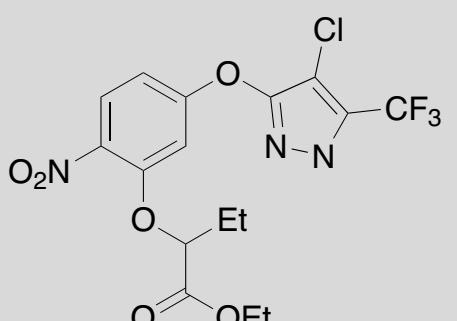
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 k' = 3.9
 α = 1.11
reference 23



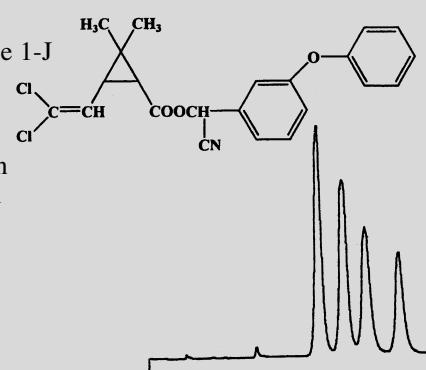
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 k' = 2.4
 α = 1.12
reference 23

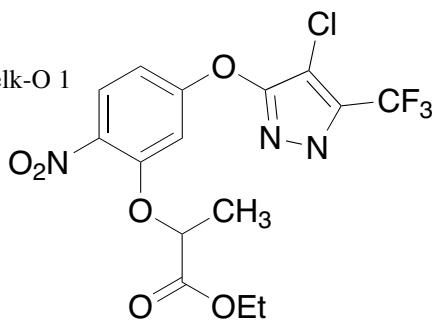


cis:trans Cypermethrin

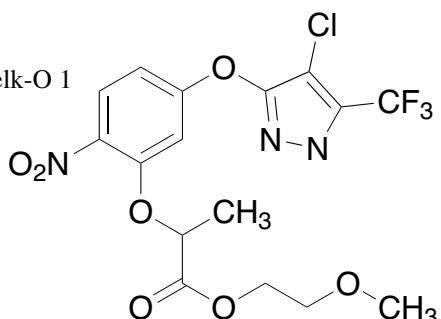
cis:trans Cypermethrin
 Column = (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 22.0 min
 k'_1 (trans) = 4.59
 α (trans) = 1.19
 k'_1 (cis) = 6.19
 α (cis) = 1.18
 reference 46

**PPO Inhibitor**

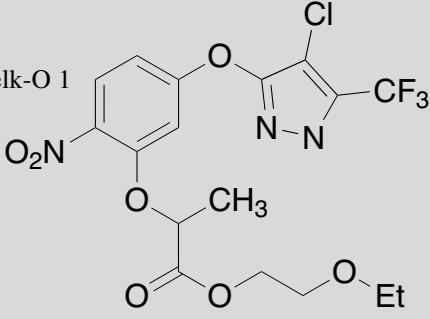
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 3.2
 α = 1.08
 reference 23

**PPO Inhibitor**

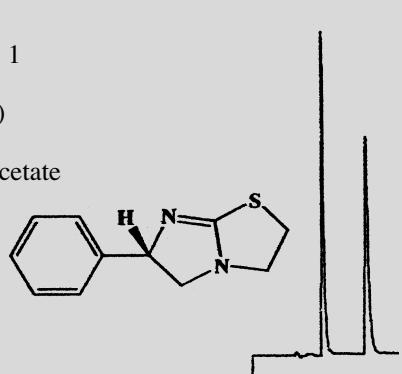
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 6.1
 α = 1.08
 reference 23

**PPO Inhibitor**

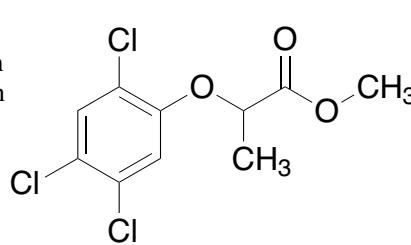
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 4.2
 α = 1.10
 reference 23

**Tetramisole**

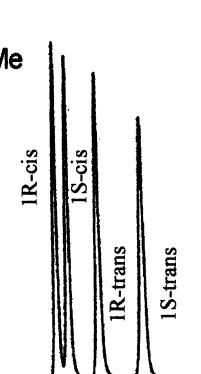
Tetramisole
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (40/40/20)
 CH₂Cl₂/Hexane/Ethanol
 + 0.01 M Ammonium Acetate
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 7.0 min
 k'_1 = 0.52
 α = 2.84
 reference 46

**Silvex Methyl**

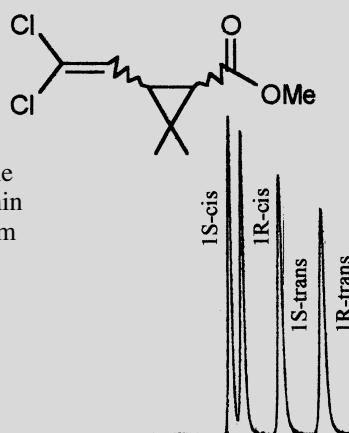
Silvex Methyl
 (herbicide)
 hexane
 1 ml/min; 254 nm
 run time = 15 min
 4.6 mm x 25 cm
 Whelk-O 1
 k'_1 = 6.47
 α = 1.05
 reference 43



Column = (R,R)-
 Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = Hexane
 Flow Rate = 0.5 mL/min
 Detection = UV 254 nm
 Run Time = 15.5 min
 reference 54



Column = (S,S)-
 Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = Hexane
 Flow Rate = 0.5 mL/min
 Detection = UV 254 nm
 Run Time = 18.5 min
 reference 54



Fluazifop-butyl

Fluazifop-butyl

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

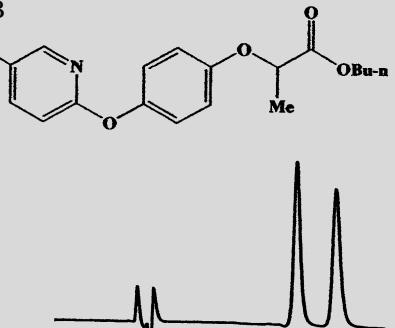
Detection: UV 254 nm

Run Time: 11.5 min

$k'_1 = 2.65$

$\alpha = 1.22$

reference: 59



Haloxyfop-ethoxyethyl

Haloxyfop-ethoxyethyl

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

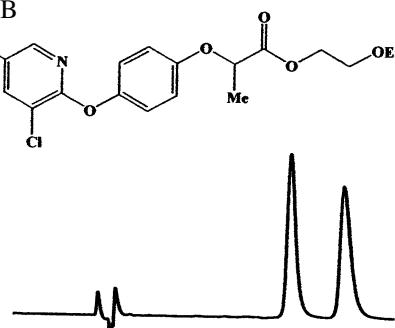
Detection: UV 254 nm

Run Time: 13.0 min

$k'_1 = 3.13$

$\alpha = 1.25$

reference: 59



Fenoxaprop-ethyl

Fenoxaprop-ethyl

Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

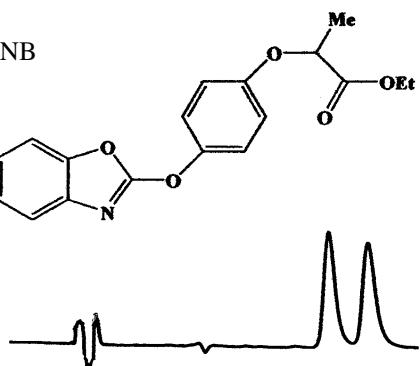
Detection: UV 254 nm

Run Time: 18.0 min

$k'_1 = 4.70$

$\alpha = 1.15$

reference: 59



Quizalofop-ethyl

Quizalofop-ethyl

Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

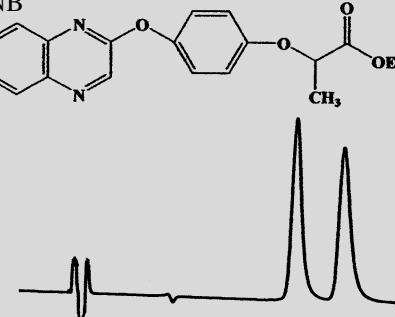
Detection: UV 254 nm

Run Time: 20.0 min

$k'_1 = 5.22$

$\alpha = 1.21$

reference: 59



Dinocap

Dinocap (fungicide) - mixture of isomers

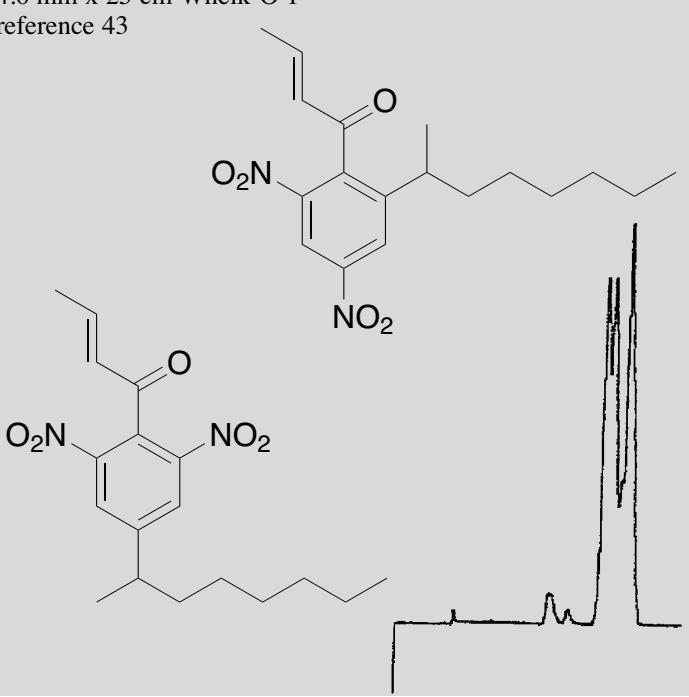
hexane

1 ml/min; 254 nm

Run Time = 15 min

4.6 mm x 25 cm Whelk-O 1

reference 43



Fenvalerate

Fenvalerate

Column: (S,S)-Whelk-O 1

10/100 (FEC) 25 cm x 4.6 mm

Mobile Phase: (99/1)

Hexane/IPA

Flow Rate: 3.0 mL/min

Detection: UV 254 nm

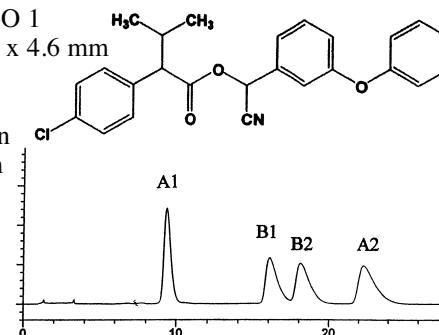
$k'_{\text{A1}} = 9.36$

$\alpha_{(\text{A1},\text{A2})} = 2.54$

$k'_{\text{B1}} = 16.79$

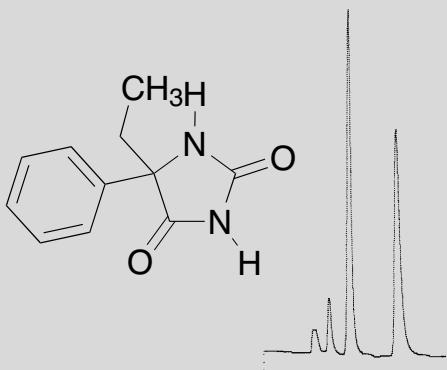
$\alpha_{(\text{B1},\text{B2})} = 1.14$

reference 46

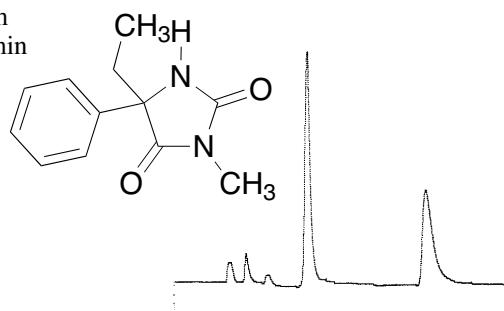


Nirvanol

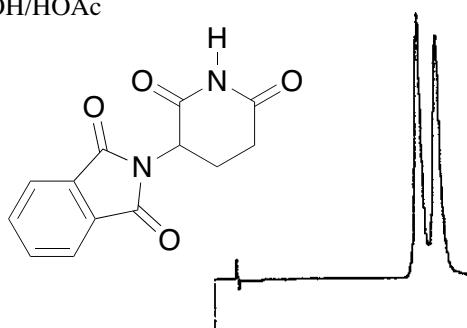
Nirvanol
20% IPA/hexane
1 ml/min; 254 nm
Run Time = 8 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.50$
 $\alpha = 2.57$
reference 31

**Mephenytoin**

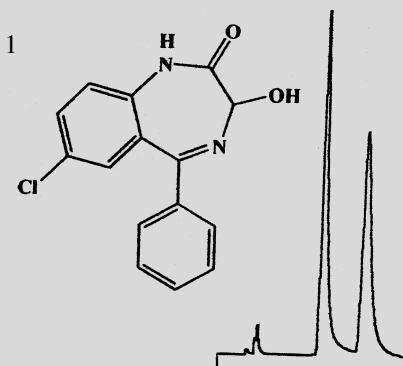
Mephenytoin
20% IPA/hexane
1 ml/min; 254 nm
Run Time = 14 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.57$
 $\alpha = 2.46$
reference 31

**Thalidomide**

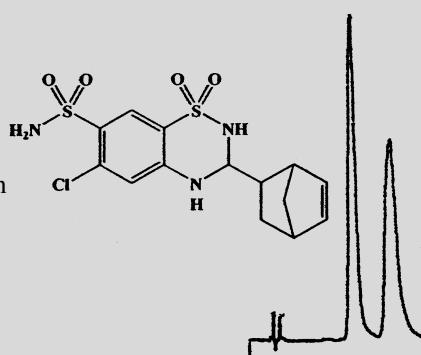
Thalidomide
63:37:0.1 $\text{H}_2\text{O}/\text{MeOH}/\text{HOAc}$
1 ml/min; 254 nm
Run Time = 33 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 10.19$
 $\alpha = 1.10$
reference 18

**Oxazepam**

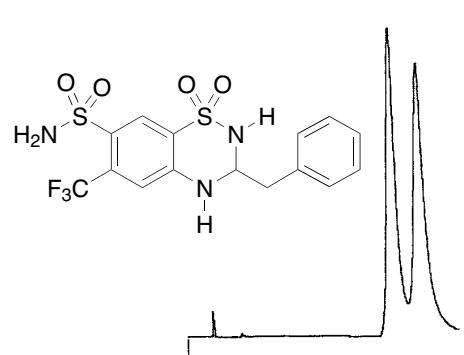
Oxazepam
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA + 0.01 M
Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 9.5 min
 $k'_1 = 2.73$
 $\alpha = 1.56$
reference 46

**Cyclothiazide**

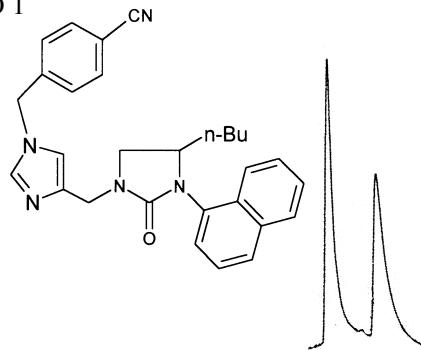
Cyclothiazide
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA + 0.1%
Acetic Acid
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 12.0 min
 $k'_1 = 3.71$
 $\alpha = 1.47$
reference 46

**Bendroflumethiazide**

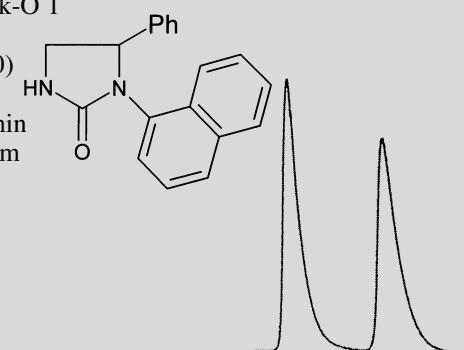
bendroflumethiazide
 $k'_1 = 7.89$
 $\alpha = 1.16$
1:1 hexane/IPA
1 ml/min; 220 nm
run time = 30 min
4.6 mm x 25 cm
Whelk-O 1
reference 18



Column: (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (60/40)
Ethanol/Hexane + 0.1%
Triethylamine
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 32.0 min
 $k'_1 = 3.78$
 $\alpha = 1.66$
reference 55

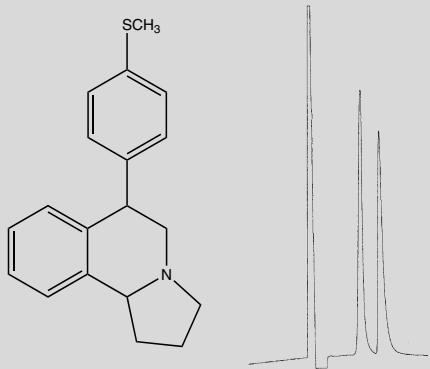


Column: (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (80/20)
Hexane/IPA
Flow Rate: 2.0 mL/min
Detection: UV 254 nm
Run Time: 32.0 min
 $k'_1 = 15.64$
 $\alpha = 1.33$
reference 55



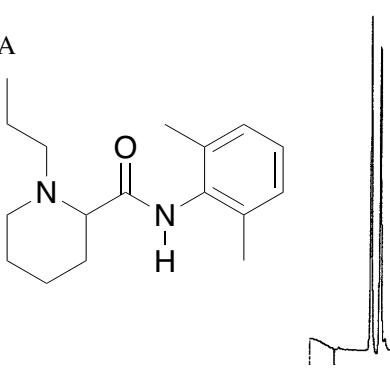
McN 5652

McN 5652
2% IPA/hex w. 0.2% diethylamine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.85$
 $\alpha = 1.36$
reference 32



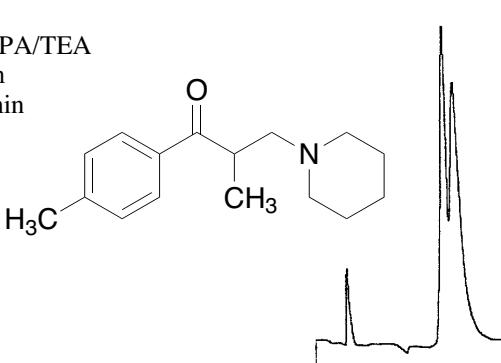
Bupivacaine

Bupivacaine
80:20:0.1 hexane/IPA/TEA
1 ml/min; 254 nm
Run Time = 7-8 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.89$
 $\alpha = 1.25$
reference 18



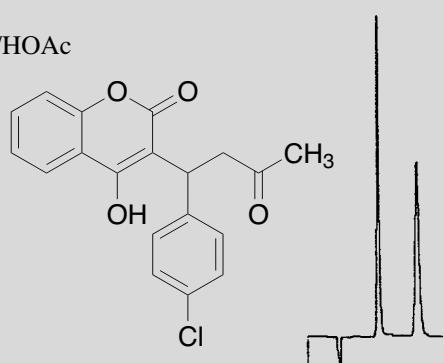
Tolperisone

Tolperisone
99:1:0.1 hexane/IPA/TEA
1 ml/min; 254 nm
Run Time = 18 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 4.81$
 $\alpha = 1.10$
reference 18



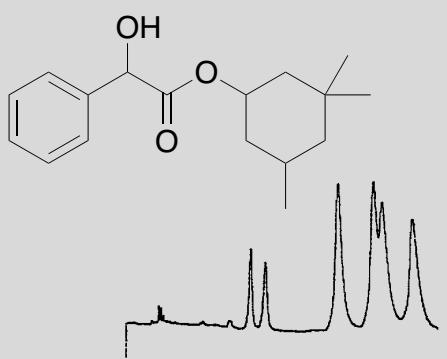
p-Chloro-Warfarin

p-Chloro-Warfarin
85:15:0.1 MeOH/H₂O/HOAc
1 ml/min; 254 nm
Run Time = 12 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.64$
 $\alpha = 1.93$
reference 18



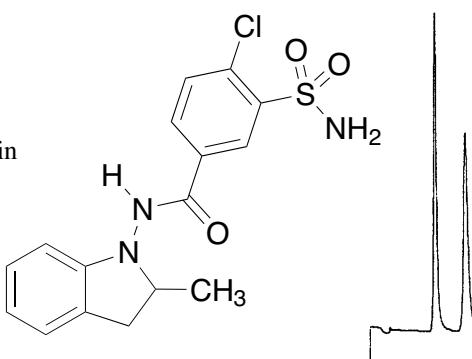
Cyclandelate

Cyclandelate
(mixture of isomers)
hexane
1 ml/min; 254 nm
Run Time = 35 min
4.6 mm x 25 cm
Whelk-O 1
reference 18

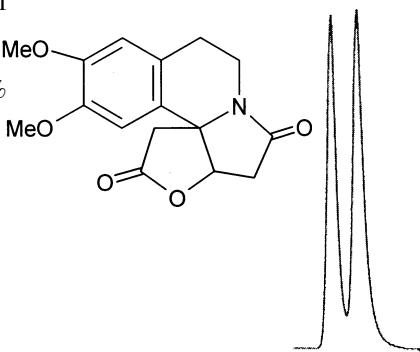


Indapamide

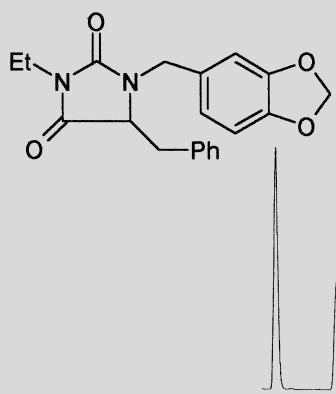
Indapamide
 $k'_1 = 2.46$
 $\alpha = 1.68$
1:1 hexane/IPA
1 ml/min; 220 nm
Run Time = 14 min
4.6 mm x 25 cm
Whelk-O 1
reference 18



Column: (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (60/40)
Ethanol/Hexane + 0.1% Triethylamine
Flow Rate: 1.0 mL/min
Detection: UV 280 nm
Run Time: 17.0 min
 $k'_1 = 3.78$
 $\alpha = 1.14$
reference 56

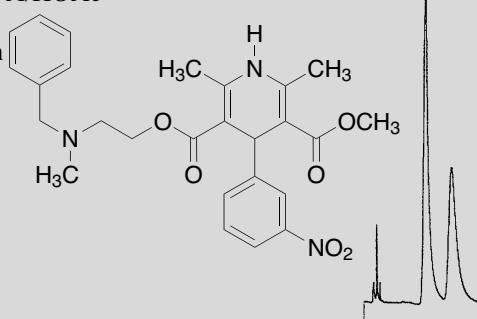


Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 25.0 min
 $k'_1 = 3.45$
 $\alpha = 2.04$
reference 53

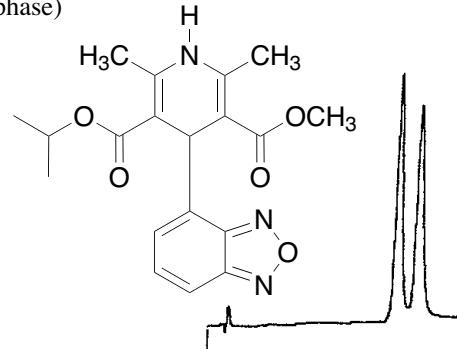


Nicardipine

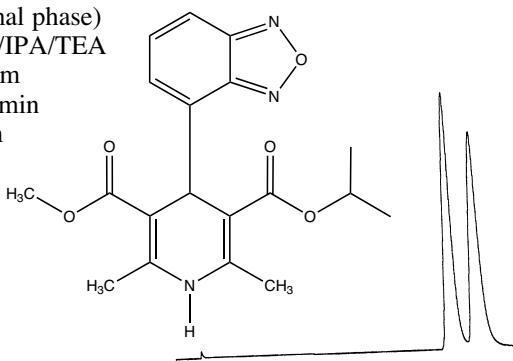
Nicardipine
73:27:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 30 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 6.06$
 $\alpha = 1.52$
reference 18

**Isradipine (reversed phase)**

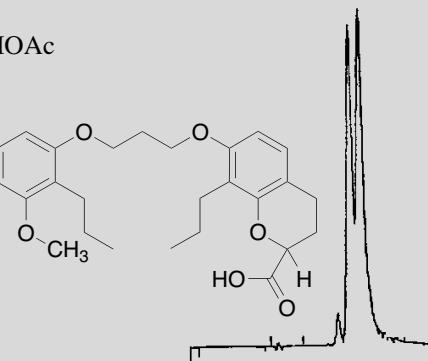
Isradipine (reversed phase)
63/37 MeOH/H₂O
1 ml/min; 254 nm
Run Time = 35 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 11.21$
 $\alpha = 1.12$
reference 18

**Isradipine (normal phase)**

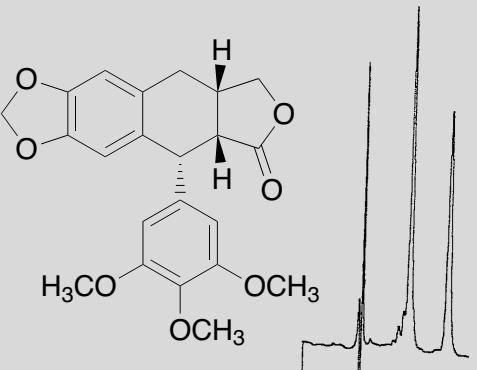
Isradipine (normal phase)
98:2:0.5 hexane/IPA/TEA
1 ml/min; 254 nm
Run Time = 52 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 9.71$
 $\alpha = 1.10$
reference 18

**SC 41930**

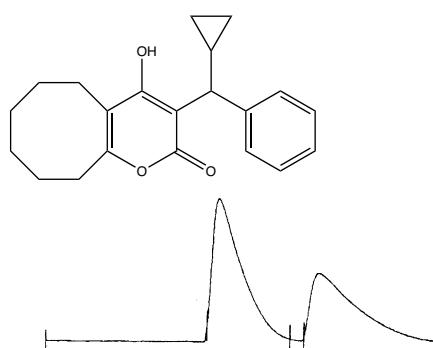
SC 41930
80:20:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 6 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.05$
 $\alpha = 1.12$
reference 7

**Hanessian's Lignan**

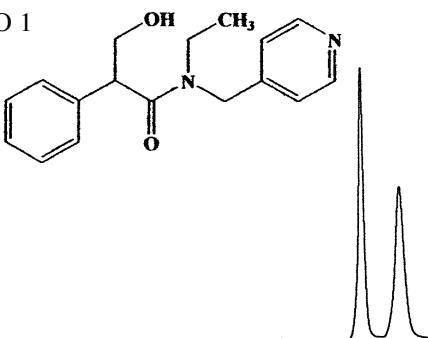
Hanessian's lignan
methanol
1 ml/min; 254 nm
Run Time = 8 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.94$
 $\alpha = 1.69$
reference 7

**U-100057**

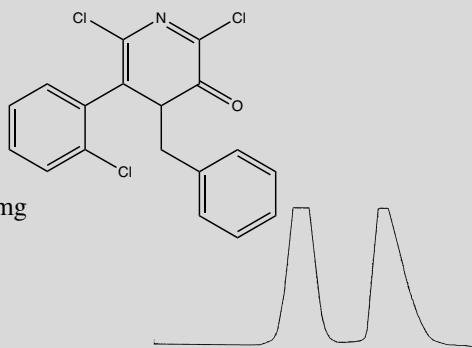
U-100057
65:35 hexane/IPA
90 ml/min to 34 min,
then 120 ml/min
Run Time = 50 min
5.1 cm x 25 cm
Whelk-O 1
Sample Load = 1.9 g
reference 37

**Tropicamide**

Tropicamide
Column: (R,R)-Whelk-O 1
10/100 (FEC)
25 cm x 4.6 mm
Mobile Phase: (75/25)
Hexane/Ethanol
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time = 13.9 min
 $k'_1 = 4.52$
 $\alpha = 1.49$
reference 46

**U-94863**

U-94863
70:30:0.5 hexane/
IPA/HOAC
12 ml/min; 254 nm
2.1 cm x 25 cm
Whelk-O 1
Run Time = 12 min
Sample Load = 40 mg
reference 37



Troglitazone

Troglitazone

Column = (S,S)-Whelk-O 1
10/100 (FEC)
25 cm x 4.6 mm

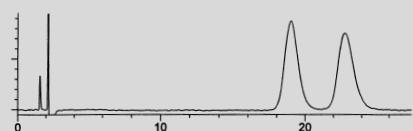
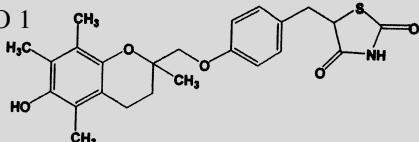
Mobile Phase = (90/10)
Hexane/IPA
+ 0.1% Acetic Acid

Flow Rate = 2.0 mL/min
Detection = UV 220 nm

$k'_1 = 13.05$

$\alpha = 1.22$

reference 46



U-94863

U-94863

40:60:0.5 hexane/IPA/HOAc

1 ml/min; 254 nm

run time = 15 min

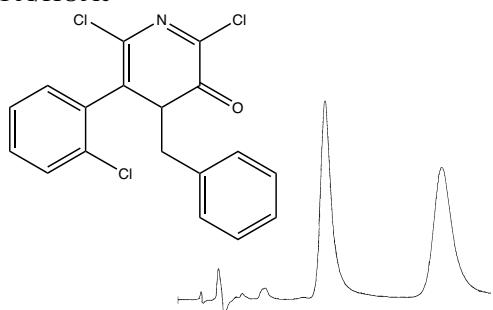
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 2.26$

$\alpha = 1.95$

reference 37



Temazepam

Column: (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase: (97/3)

Hexane/IPA +
0.1% Acetic acid

Flow Rate: 1.5 mL/min

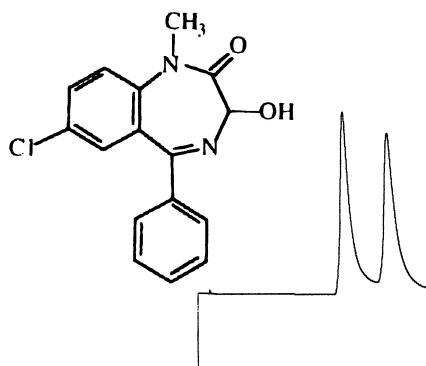
Detection: UV 254 nm

Run Time: 31.0 min

$k'_1 = 12.05$

$\alpha = 1.34$

reference 46



30% IPA/hexane

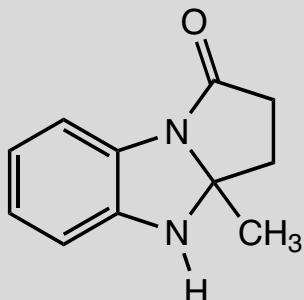
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 1.61$

$\alpha = 1.48$

reference 44



Proglumide

75:25:0.1 hexane/IPA/HOAc

1 ml/min; 254 nm

run time = 10 min

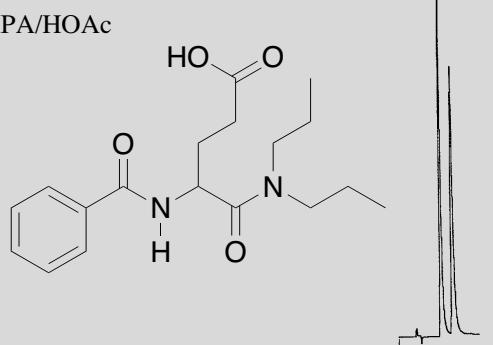
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 1.54$

$\alpha = 1.49$

reference 18



30% IPA/hexane

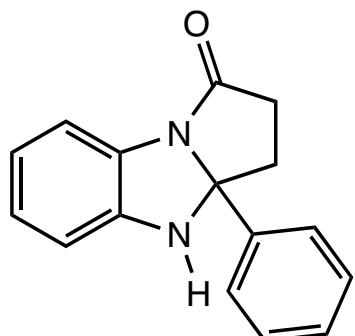
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 1.29$

$\alpha = 1.83$

reference 44



30% IPA/hexane

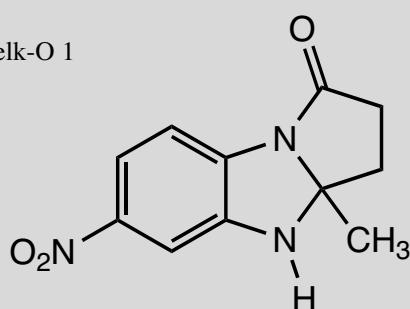
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

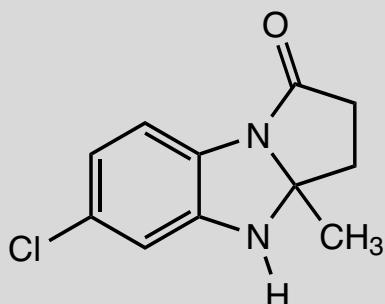
$k'_1 = 2.56$

$\alpha = 1.25$

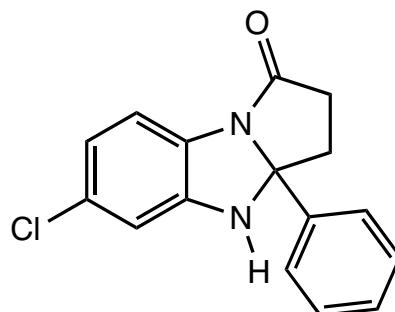
reference 44



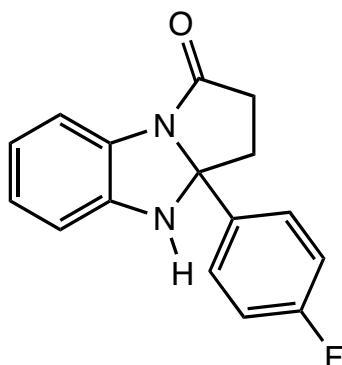
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.38$
 $\alpha = 1.44$
reference 44



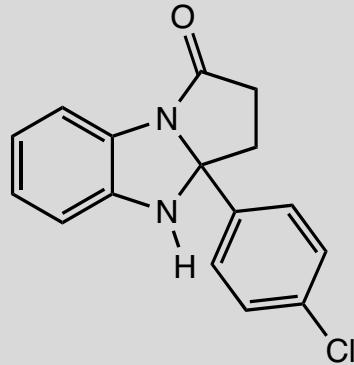
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.34$
 $\alpha = 1.60$
reference 44



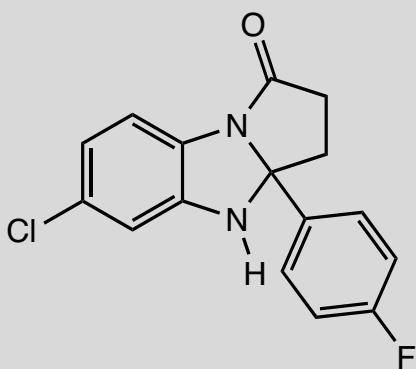
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.29$
 $\alpha = 1.83$
reference 44



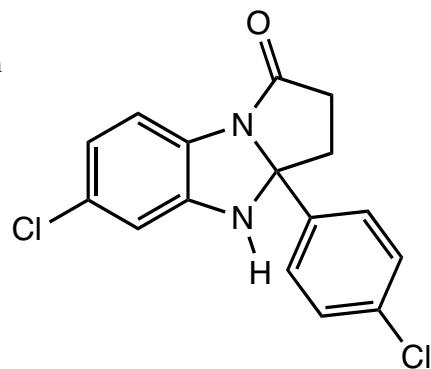
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.37$
 $\alpha = 1.90$
reference 44



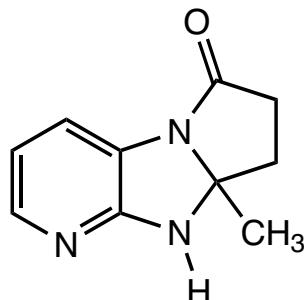
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.18$
 $\alpha = 1.72$
reference 44



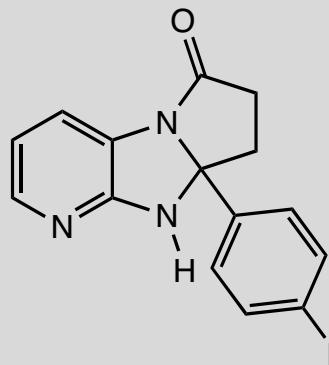
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.34$
 $\alpha = 1.78$
reference 44



70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.36$
 $\alpha = 1.33$
reference 44



70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.70$
 $\alpha = 1.55$
reference 44

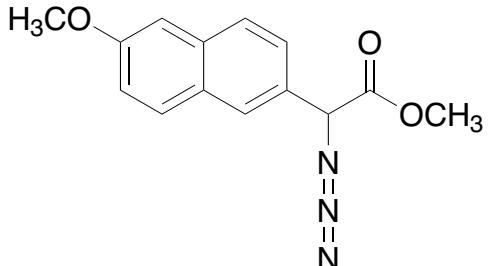


REGIS Miscellaneous Pharmaceuticals

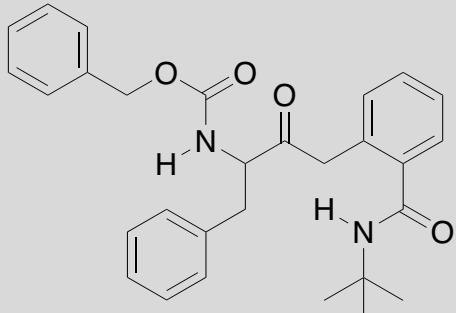
70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.90$
 $\alpha = 1.45$
reference 44



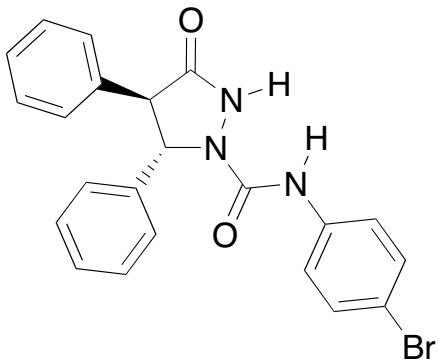
40% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.34$
 $\text{Rs} = 2.10$
reference 45



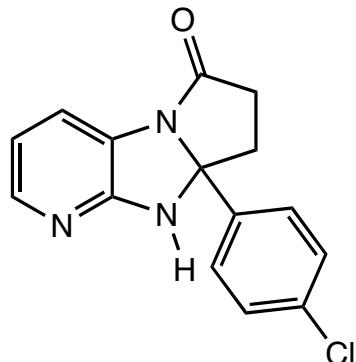
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.10$
 $\text{Rs} = 0.95$
reference 45



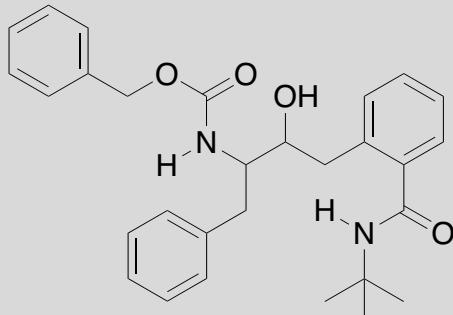
45% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 2.17$
 $\text{Rs} = 2.20$
reference 45



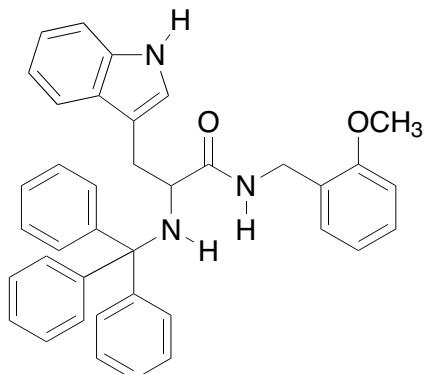
70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.73$
 $\alpha = 1.59$
reference 44



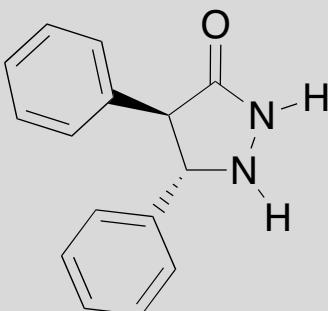
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.29$
 $\text{Rs} = 2.10$
reference 45



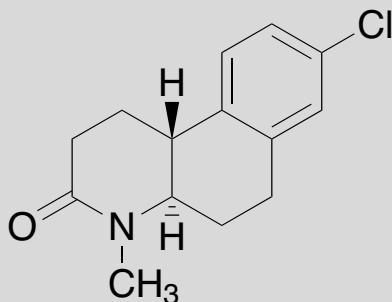
50% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.32$
 $\text{Rs} = 2.10$
reference 45



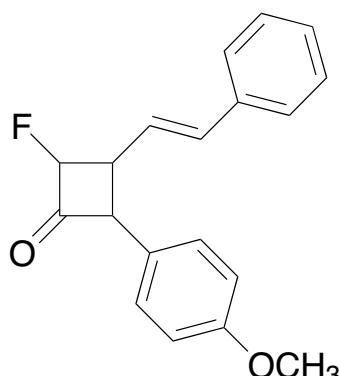
45% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.57$
 $\text{Rs} = 2.20$
reference 45



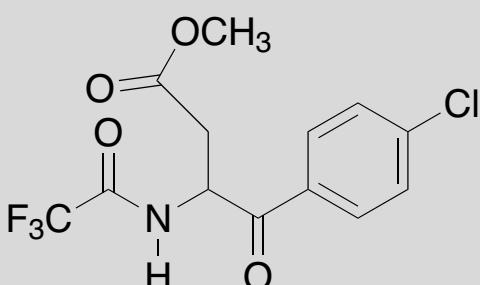
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.04$
Rs = 0.60
reference 45



15% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.13$
Rs = 1.50
reference 45

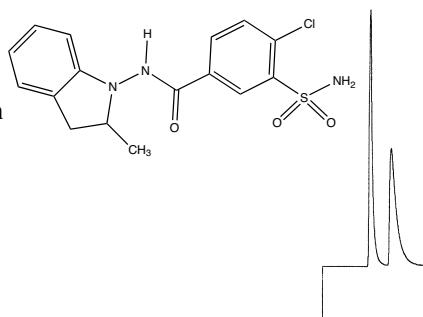


10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.11$
Rs = 1.50
reference 45

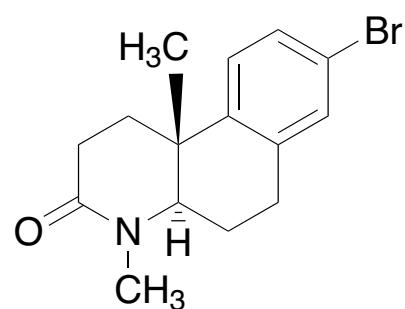


Indapamide

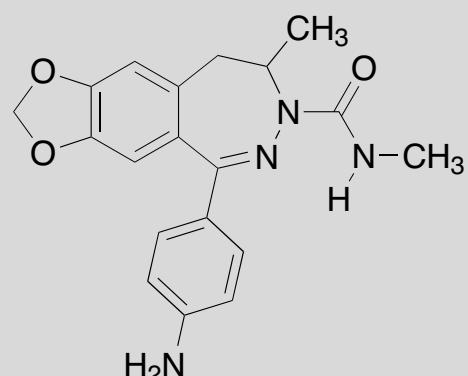
Indapamide
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 16 min
 $k'_1 = 3.09$
 $\alpha = 1.58$
reference 46



10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.04$
Rs = 0.60
reference 45

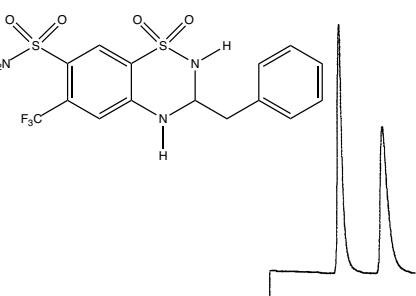


40% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.22$
Rs = 1.50
reference 45



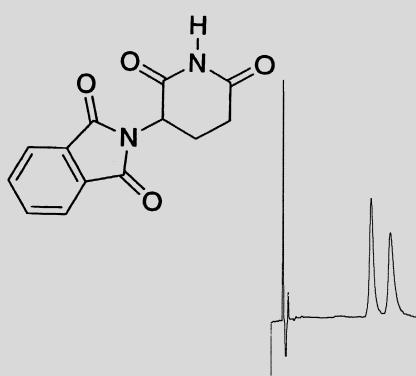
Bendroflumethiazide

Bendroflumethiazide
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 18 min
 $k'_1 = 2.99$
 $\alpha = 1.84$
reference 46



Thalidomide

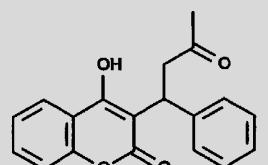
Thalidomide
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA + 0.1%
Acetic acid
Flow Rate = 1.0 mL/min
Detection = UV 220 nm
Run Time = 28.0 min
 $k'_1 = 7.71$
 $\alpha = 1.22$
reference 46



Warfarin

Warfarin

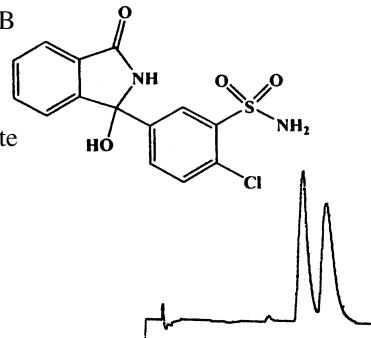
Column: (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase: (70/30)
Heptane/IPA + 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 230 nm
Run Time: 6.5 min
 $k'_1 = 0.89$
 $\alpha = 1.36$
reference 48



Chlorthalidone

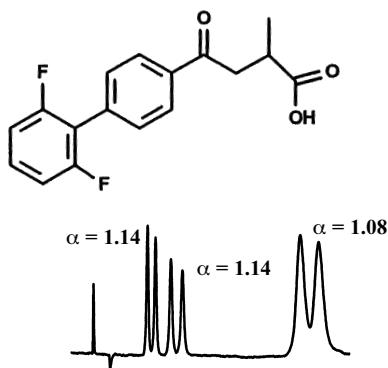
Chlorthalidone

Column = (S,S)-DACH-DNB
25 cm x 4.6 mm
Mobile Phase = (99/1)
CH₂Cl₂/CH₃OH +
0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 20.0 min
 $k'_1 = 9.38$
 $\alpha = 1.18$
reference 46



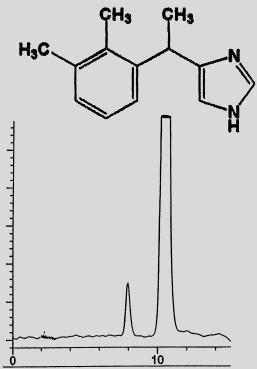
Flobufen Metabolites

Flobufen Metabolites
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (97/3)
Heptane/Glyme +
0.1% TFA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 21.0 min
reference 47



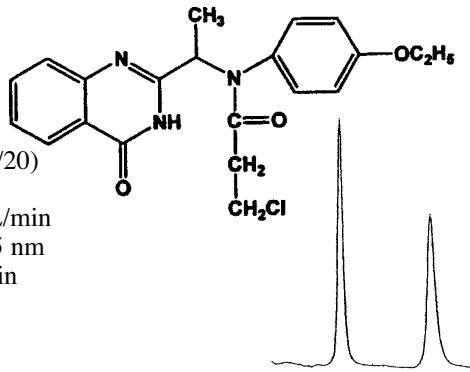
Dexmedetomidine (Enriched)

Dexmedetomidine (Enriched)
Column = (S,S)-Whelk-O 2
25 cm x 4.6 mm
Mobile Phase = (90/10)
Hexane/Ethanol
+ 10 mM Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 220 nm
 $k'_1 = 3.41$
 $\alpha = 1.39$
reference 46



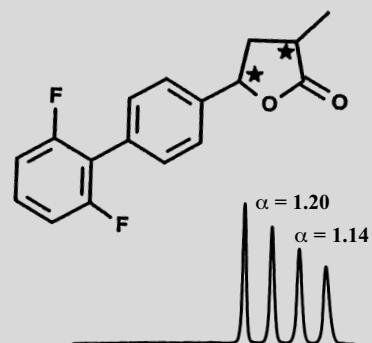
4(3H)-Quinazolone Derivatives

4(3H)-quinazolone derivatives
Column = (S,S)-
Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (80/20)
Hexane/Ethanol
Flow Rate = 1.0 mL/min
Detection = UV 225 nm
Run Time = 17.0 min
 $k'_1 = 2.95$
 $\alpha = 1.62$
reference 58



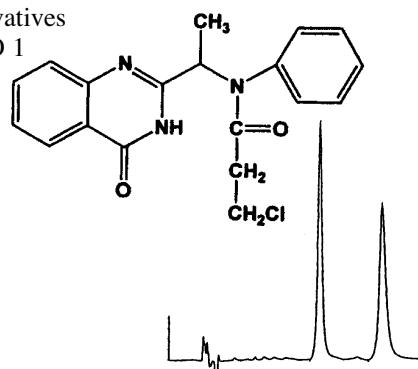
Flobufen and Flobufen Metabolites

Flobufen and Flobufen
Metabolites
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (90/10)
Heptane/IPA + 0.1% TFA
Flow Rate = 2.0 mL/min
Detection = UV 230 nm
Run Time = 24.0 min
reference 47



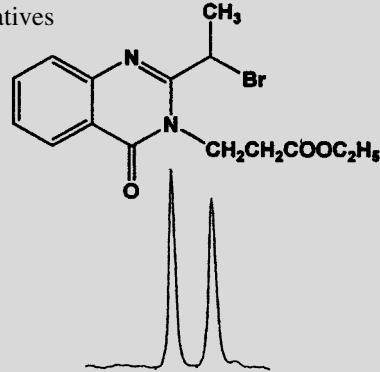
4(3H)-Quinazolone Derivatives

4(3H)-Quinazolone Derivatives
Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (80/20)
Hexane/Ethanol
Flow Rate = 1.0 mL/min
Detection = UV 225 nm
Run Time = 16.0 min
 $k'_1 = 2.88$
 $\alpha = 1.56$
reference 58



4(3H)-Quinazolone Derivatives

4(3H)-quinazolone derivatives
Column = (S,S)-
Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (90/10)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 225 nm
Run Time = 15.0 min
 $k'_1 = 3.54$
 $\alpha = 1.19$
reference 58



4(3H)-Quinazolone Derivatives

4(3H)-Quinazolone Derivatives

Column = (S,S)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (80/20)

Hexane/Ethanol

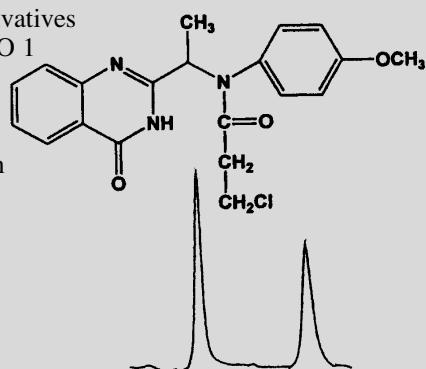
Flow Rate = 1.0 mL/min

Detection = UV 225 nm

Run Time = 21.0 min

 $k'_1 = 3.75$ $\alpha = 1.57$

reference 58

**Ifenprodil**

Ifenprodil

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (85/15)

Hexane/IPA +

0.01 M Ammonium Acetate

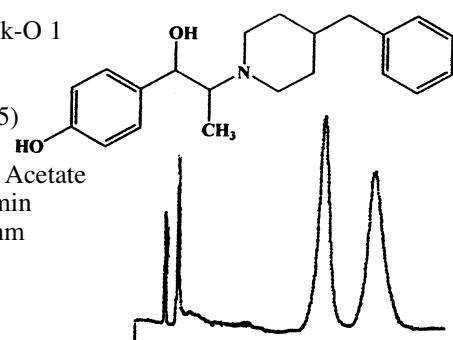
Flow Rate = 1.5 mL/min

Detection = UV 220 nm

Run Time = 16.5 min

 $k'_1 = 6.16$ $\alpha = 1.32$

reference 46

**Tofisopam and it's Conformers**

Tofisopam and it's Conformers

Column = (R,R)- β -Gem 1

25 cm x 4.6 mm

Mobile Phase = (70/30)

Hexane/Ethanol + 0.1% TEA

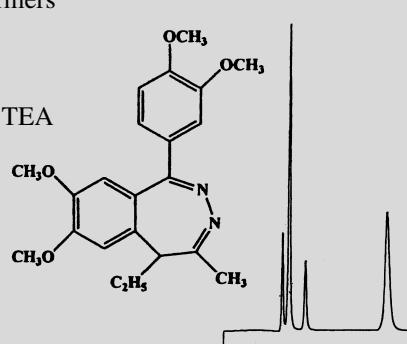
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 25.0 min

 $k'_1 = 2.66$ $\alpha = 3.13$

reference 46

**Coumachlor**

Coumachlor

Column = (R,R)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (65/35)

Hexane/Ethanol + 0.1% Acetic Acid

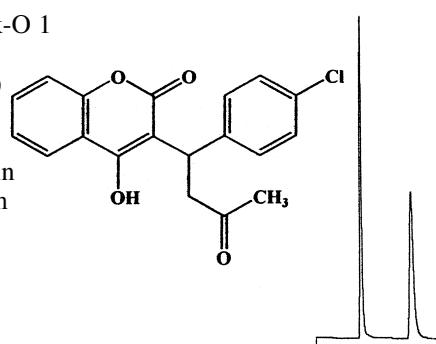
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

Run Time = 10.0 min

 $k'_1 = 1.48$ $\alpha = 2.90$

reference 46

**4(3H)-Quinazolone Derivatives**

4(3H)-Quinazolone Derivatives

Column = (S,S)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (80/20)

Hexane/Ethanol

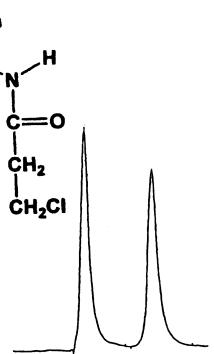
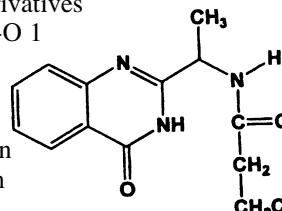
Flow Rate = 1.0 mL/min

Detection = UV 225 nm

Run Time = 15.0 min

 $k'_1 = 3.19$ $\alpha = 1.37$

reference 58

**Ketamine**

Ketamine

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (99/1)

Hexane/IPA + 0.1% TEA

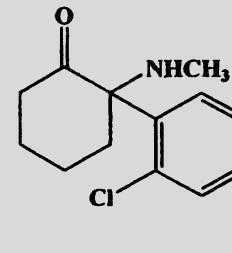
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 22.0 min

 $k'_1 = 6.37$ $\alpha = 1.14$

reference 46

**Ketoconazole**

Ketoconazole

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (46/46/8)

CH₂Cl₂/Hexane/IPA +

0.01 M Ammonium Acetate

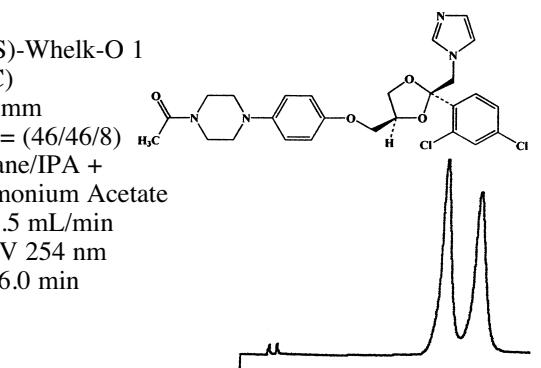
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

Run Time = 16.0 min

 $k'_1 = 6.60$ $\alpha = 1.19$

reference 46

**Sulpiride**

Sulpiride

Column = (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase = (95/5)

CH₂Cl₂/Ethanol +

0.01 M Ammonium Acetate

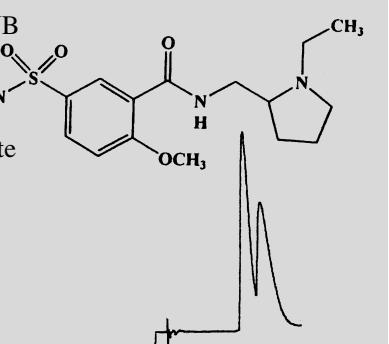
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

Run Time = 14.0 min

 $k'_1 = 5.92$ $\alpha = 1.24$

reference 46



REGIS Miscellaneous Pharmaceuticals

Ofloxacin

Ofloxacin

Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm

Mobile Phase = (43/43/14)

$\text{CH}_2\text{Cl}_2/\text{Hexane/Ethanol} + 0.01 \text{ M Ammonium Acetate}$

Flow Rate = 1.5 mL/min

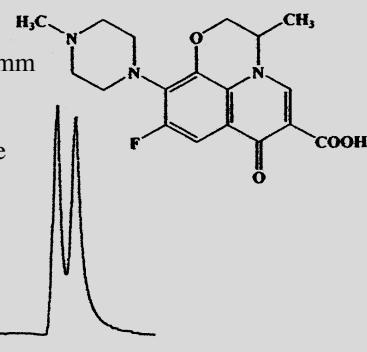
Detection = UV 254 nm

Run Time = 10.0 min

$k'_1 = 2.96$

$\alpha = 1.24$

reference 46



Isoxsuprime

Isoxsuprime

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase = (95/5)

$\text{Hexane/Ethanol} + 0.01 \text{ M Ammonium Acetate}$

Flow Rate = 2.0 mL/min

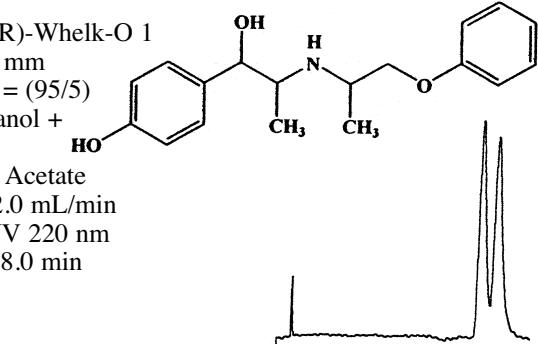
Detection = UV 220 nm

Run Time = 28.0 min

$k'_1 = 17.91$

$\alpha = 1.08$

reference 46



Warfarin (Normal Phase)

Warfarin (normal phase)

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase =

(65/35) Hexane/IPA + 0.1% Acetic Acid

Flow Rate = 1.0 mL/min

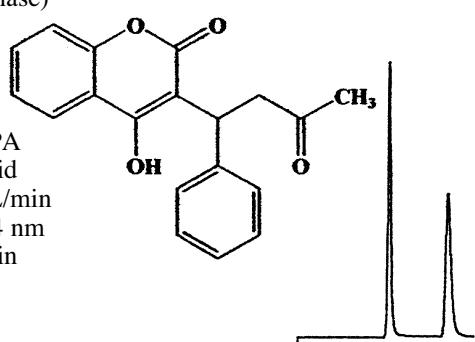
Detection = UV 254 nm

Run Time = 11.5 min

$k'_1 = 1.54$

$\alpha = 2.07$

reference 46



Cromakalim

Cromakalim

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase = (92/8)
Hexane/Ethanol

Flow Rate = 1.5 mL/min

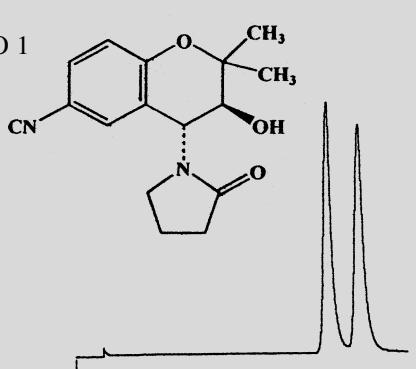
Detection = UV 254 nm

Run Time = 21.0 min

$k'_1 = 9.18$

$\alpha = 1.14$

reference 46



Temazepam

Temazepam

Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm

Mobile Phase =

(80/20) Hexane/IPA + 0.1% Acetic Acid

Flow Rate = 2.0 mL/min

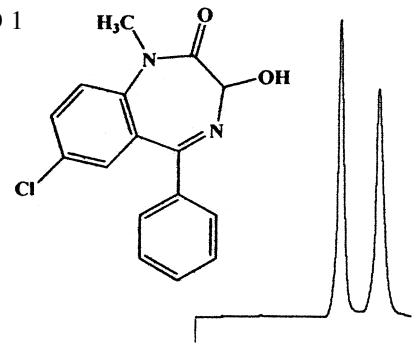
Detection = UV 254 nm

Run Time = 13.0 min

$k'_1 = 6.86$

$\alpha = 1.34$

reference 46



Warfarin (Reversed Phase)

Warfarin (reversed phase)

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase =

(70/30) $\text{CH}_3\text{OH}/\text{H}_2\text{O} + 0.1\%$ Acetic Acid

Flow Rate = 1.0 mL/min

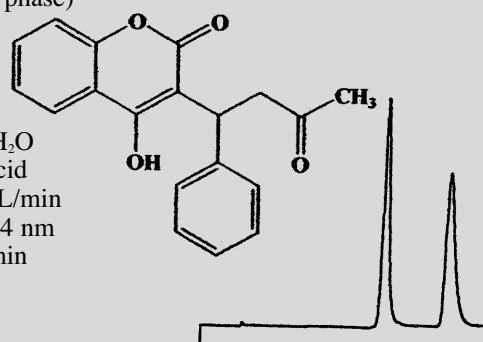
Detection = UV 254 nm

Run Time = 15.0 min

$k'_1 = 3.54$

$\alpha = 1.55$

reference 46



Trichlormethiazide

Trichlormethiazide

Column = (R,R)-ULMO
25 cm x 4.6 mm

Mobile Phase =

(75/25)
Hexane/IPA + 0.1% Acetic Acid

Flow Rate = 1.5 mL/min

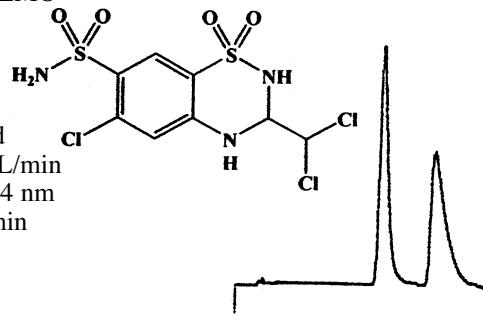
Detection = UV 254 nm

Run Time = 15.0 min

$k'_1 = 5.16$

$\alpha = 1.43$

reference 46



Prilocaine

Prilocaine

Column = (S,S)-ULMO
25 cm x 4.6 mm

Mobile Phase =

(99/1) Hexane/
Ethanol + 0.01 M Ammonium Acetate

Flow Rate = 1.5 mL/min

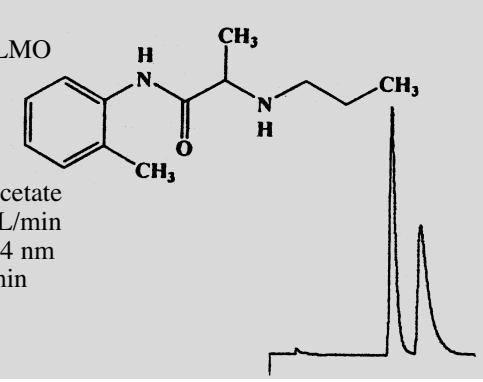
Detection = UV 254 nm

Run Time = 15.0 min

$k'_1 = 5.70$

$\alpha = 1.28$

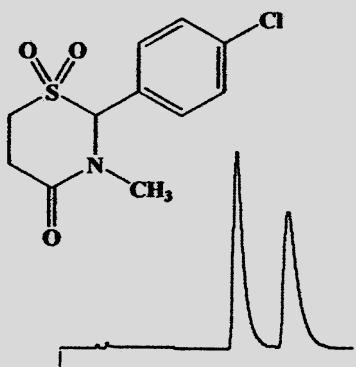
reference 46



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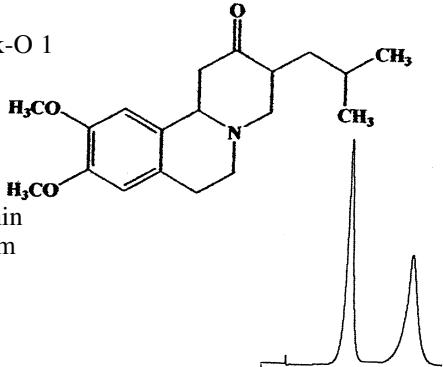
Chlormezanone

Chlormezanone
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (60/40)
 Hexane/IPA
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 13.0 min
 $k'_1 = 4.48$
 $\alpha = 1.36$
 reference 46



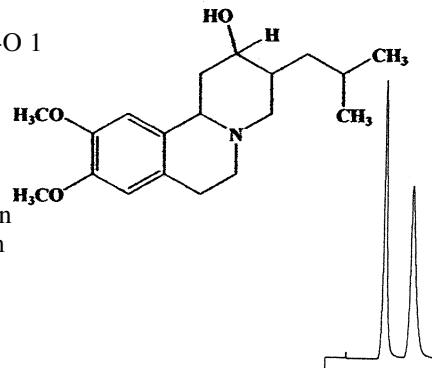
Tetrabenazine

Tetrabenazine
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase =
 (55/45) Hexane/IPA
 + 0.1% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 280 nm
 Run Time = 13.4 min
 $k'_1 = 3.35$
 $\alpha = 1.93$
 reference 46



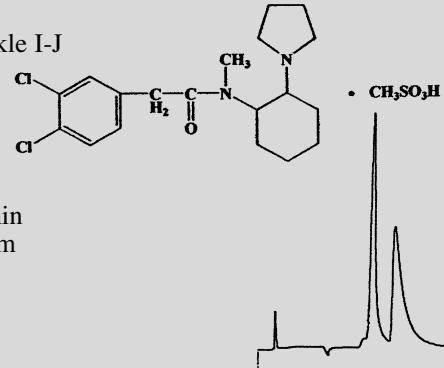
Dihydrotetrabenazine

Dihydrotetrabenazine
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase =
 (60/40) Hexane/IPA
 + 0.1% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 280 nm
 Run Time = 9.3 min
 $k'_1 = 2.50$
 $\alpha = 1.65$
 reference 46



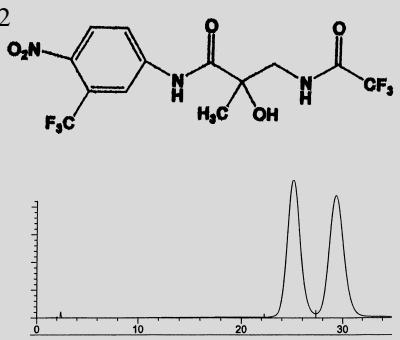
trans-U-50488H

trans-U-50488H
 Column = (3R,4S)-Pirkle I-J
 25 cm x 4.6 mm
 Mobile Phase = (92/8)
 Hexane/Ethanol +
 0.01 M Ammonium
 Acetate
 Flow Rate = 2.0 mL/min
 Detection = UV 220 nm
 Run Time = 12.0 min
 $k'_1 = 6.71$
 $\alpha = 1.27$
 reference 46



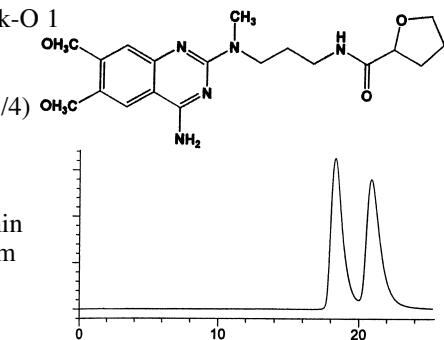
Fluridil

Fluridil
 Column = (S,S)-Whelk-O 2
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase =
 (57/43)
 H₂O/CH₃OH
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 12.9$
 $\alpha = 1.18$
 reference 46



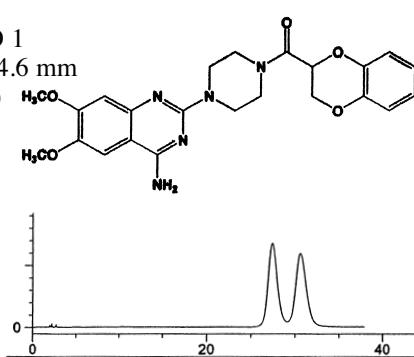
Alfuzosin

Alfuzosin
 Column = (R,R)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase = (68/28/4)
 Hexane/CH₂Cl₂/
 Ethanol + 4 mM
 Ammonium Acetate
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 $k'_1 = 7.37$
 $\alpha = 1.15$
 reference 46



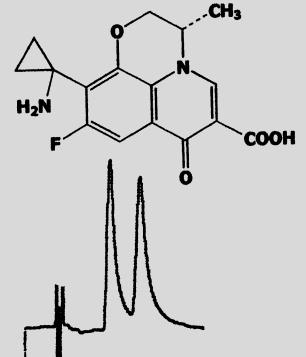
Doxazosin

Doxazosin
 Column = (S,S)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (66/29/5)
 Hexane/CH₂Cl₂/
 Ethanol + 5 mM
 Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 14.2$
 $\alpha = 1.13$
 reference 46



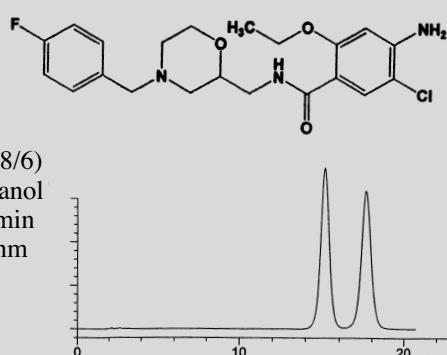
Pazufloxacin

Pazufloxacin
 Column = (S,S)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (40/40/20)
 CH₂Cl₂/Hexane/IPA
 + 0.15% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 6.7 min
 $k'_1 = 1.71$
 $\alpha = 1.58$
 reference 46

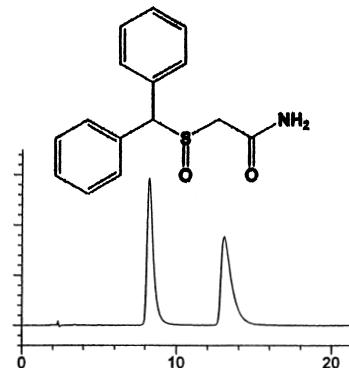


Mosapride

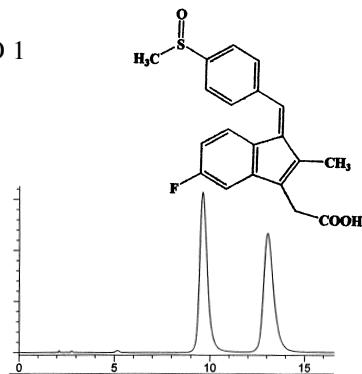
Mosapride
 Column = (R,R)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase = (66/28/6)
 Hexane/CH₂Cl₂/Ethanol
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 7.37$
 $\alpha = 1.19$
 reference 46

**Modafinil**

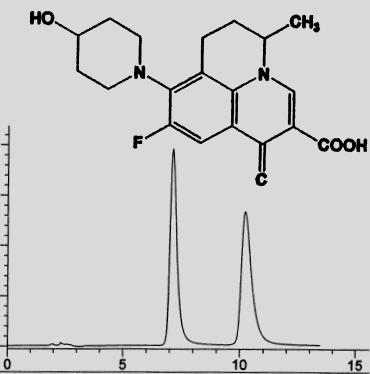
Modafinil
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase = (65/35)
 Hexane/IPA
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 3.57$
 $\alpha = 1.75$
 reference 46

**Sulindac**

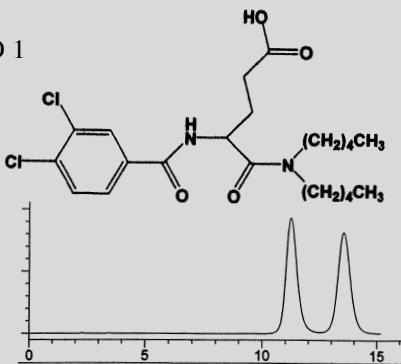
Sulindac
 Column = (R,R)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase = (48/48/4)
 Hexane/CH₂Cl₂/IPA
 + 0.1% Acetic acid
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 4.32$
 $\alpha = 1.45$
 reference 46

**Nadifloxacin**

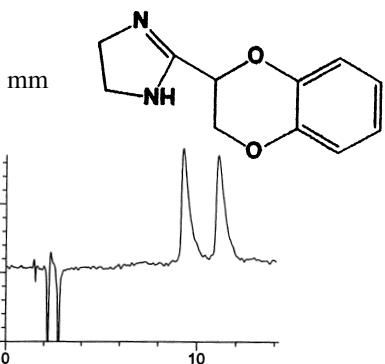
Nadifloxacin
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase = (45/45/10)
 CH₂Cl₂/Hexane/IPA
 + 10 mM Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 $k'_1 = 2.95$
 $\alpha = 1.58$
 reference 46

**Lorglumide**

Lorglumide
 Column = (R,R)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA
 + 0.1% Acetic Acid
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 $k'_1 = 5.22$
 $\alpha = 1.25$
 reference 46

**Idazoxan**

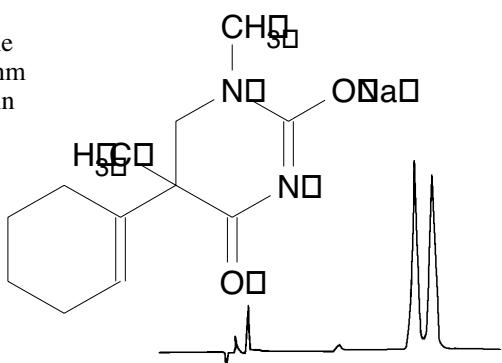
Idazoxan
 Column = (S,S)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (70/29/1)
 Hexane/Methylene Chloride/IPA
 + 0.1% TEA
 Flow Rate = 2.0 mL/min
 Detection - UV 254 nm
 $k'_1 = 5.86$
 $\alpha = 1.23$
 reference 46



REGIS Barbiturates

Hexobarbital

Hexobarbital
5% EtOH/Hexane
0.7 ml/min; 254nm
run time = 16 min
4.6 mm x 25 cm
L-Leucine
 $k'_1 = 2.89$
 $\alpha = 1.10$

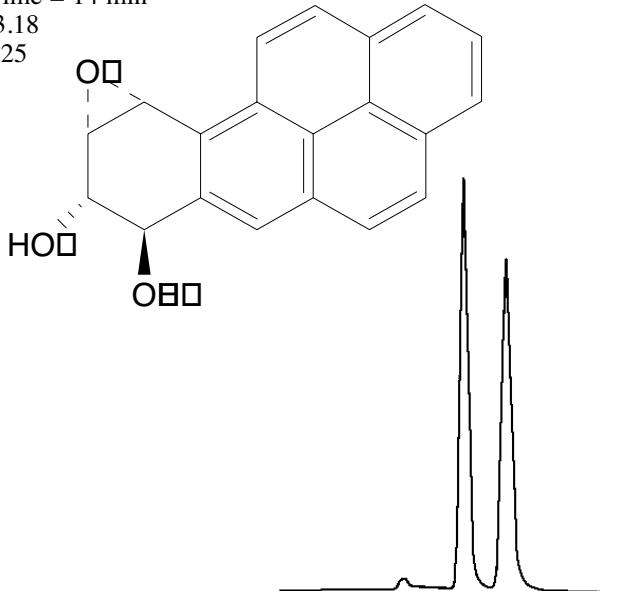


Diol Epoxides

REGIS

r-7,t-8-Dihydroxy-t-9, 10-epoxy-7,8,9, 10-tetrahydrobenzo[a]pyrene

r-7,t-8-Dihydroxy-t-9, 10-epoxy-7,8,9,
10-tetrahydrobenzo[a]pyrene
40% EtOH/Hexane
4.6 mm x 25 cm (*R,R*) β-Gem 1
1 ml/min; 254 nm
Run Time = 14 min
 $k'_1 = 3.18$
 $\alpha = 1.25$

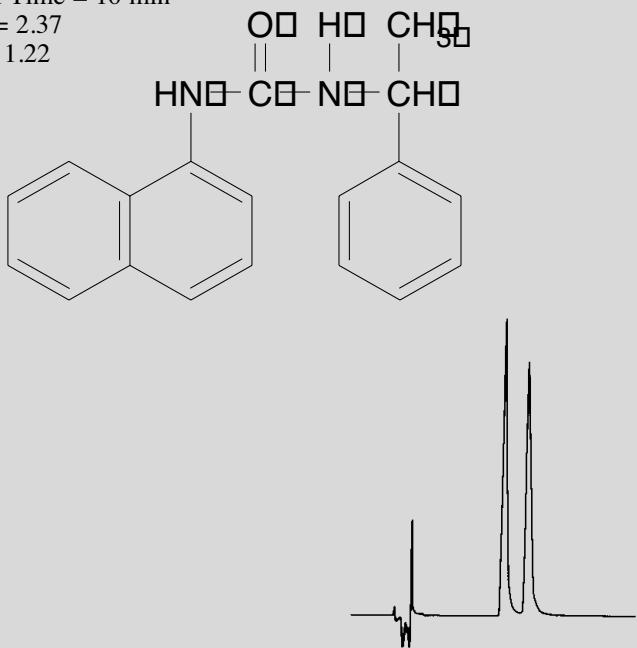


Ureas

REGIS

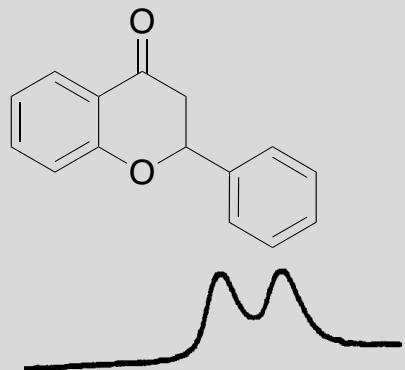
1-Naphthylureaphenethylamine

1-Naphthylureaphenethylamine
30% EtOH/Hexane
4.6 mm x 25 cm
D-Phenylglycine
1 ml/min; 254 nm
Run Time = 10 min
 $k'_1 = 2.37$
 $\alpha = 1.22$

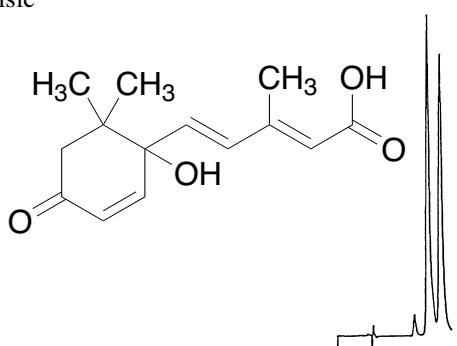


Flavanone

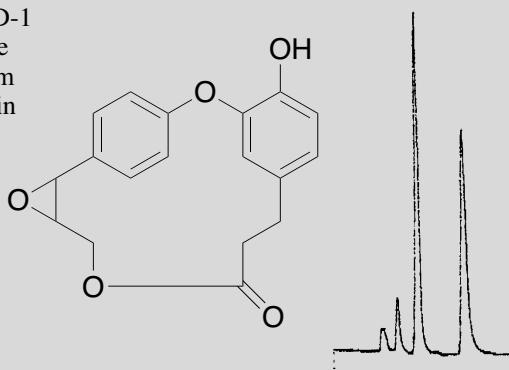
Flavanone
1% IPA/hexane
1 ml/min; 254 nm
Run Time = 25 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 7.08$
 $\alpha = 1.04$
reference 26

**2-trans-4-trans-Abscisic Acid**

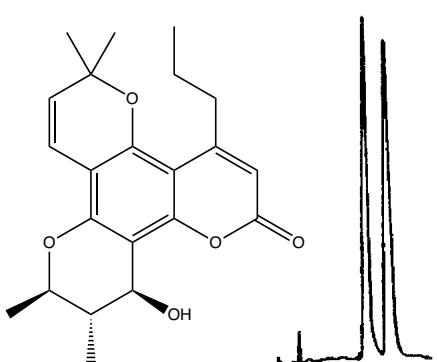
2-trans-4-trans-Abscisic Acid (ABA)
80:20:0.5
Hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 5 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.08$
 $\alpha = 1.21$
reference 9

**Combretastatin D-1**

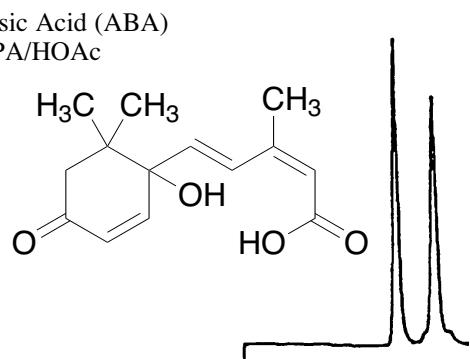
Combretastatin D-1
20% IPA/Hexane
2 ml/min; 254 nm
run time = 13 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 4.54$
 $\alpha = 1.45$
reference 17

**Calanolide A**

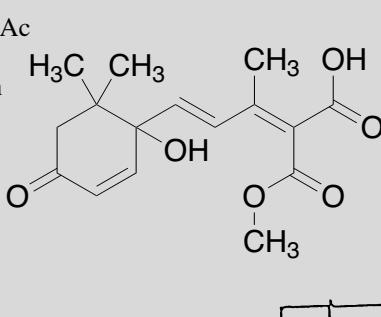
Calanolide A
10% IPA/hexane
1.25 ml/min; 270 nm
run time = 18 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 3.2$
 $\alpha = 1.4$
reference 16

**2-cis-4-trans-Abscisic Acid (ABA)**

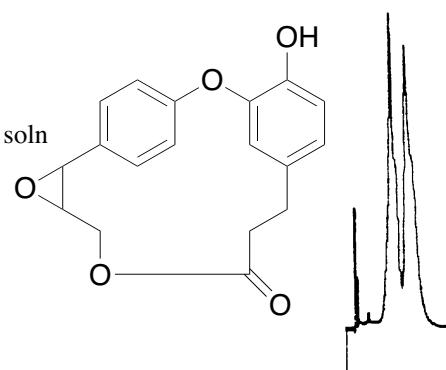
2-cis-4-trans-Abscisic Acid (ABA)
80:20:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 5 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.58$
 $\alpha = 1.39$
reference 9

**ABA Methyl Ester**

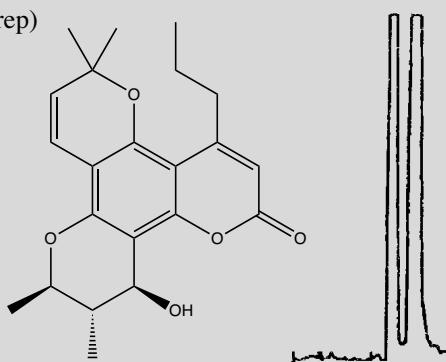
ABA Methyl Ester
80:20:0.5
Hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 5 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 1.31$
reference 9

**Combretastatin D-1**

Combretastatin D-1
semi-prep separation
20% IPA/hexane
2 ml/min; 300 nm
200 μ l of 12.7 mg/ml soln
load = 2.5 mg
run time = 10 min
4.6 mm x 25 cm
Whelk-O 1
reference 17

**Calanolide A (semi prep)**

Calanolide A (semi prep)
10% IPA/hexane
1.25 ml/min; 270 nm
run time = 18 min
4.6 mm x 25 cm
Whelk-O 1
5 mg sample
 $k'_1 = 3.2$
 $\alpha = 1.4$
reference 16



Abscisic Acid

Abscisic Acid

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase = (85/15)
Hexane/IPA +
0.1% Acetic Acid

Flow Rate = 1.5 mL/min

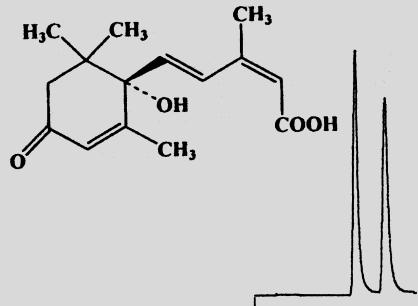
Detection = UV 254 nm

Run Time = 11.0 min

$k'_1 = 3.52$

$\alpha = 1.38$

reference 46



Tetrahydropalmatine

Tetrahydropalmatine

Column = (S,S)-Whelk-O 1
10/100 (FEC)
25 cm x 4.6 mm

Mobile Phase = (50/50)
Hexane/IPA

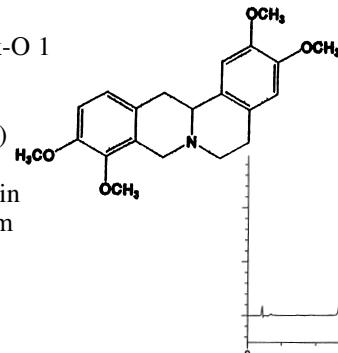
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

$k'_1 = 6.66$

$\alpha = 1.46$

reference 46



Taxifolin

Taxifolin

Column = (S,S)-Whelk-O 2
10/100 (FEC)
25 cm x 4.6 mm

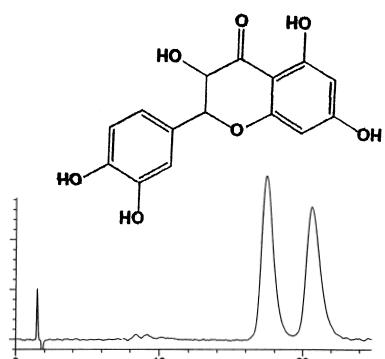
Mobile Phase = (85/15)
Hexane/Ethanol + 0.1% TFA

Flow Rate = 2.0 mL/min
Detection = UV 220 nm

$k'_1 = 11.87$

$\alpha = 1.20$

reference 46



Luciferin

Luciferin

Column = L-Leucine
25 cm x 4.6 mm

Mobile Phase = (60/40)
Hexane/Ethanol + 0.04 mM Ammonium Acetate

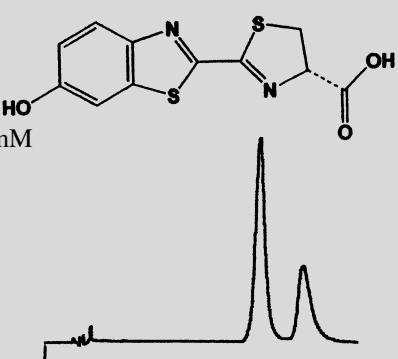
Flow Rate = 1.5 mL/min
Detection = UV 254 nm

Run Time = 15.5 min

$k'_1 = 6.09$

$\alpha = 1.25$

reference 46



REGIS β -Blockers

β -Blocker

β -blocker

Column: (S,S)-DACH-DNB
25 cm x 4.6 mm

Mobile Phase: (92/8)
CH₂Cl₂/IPA

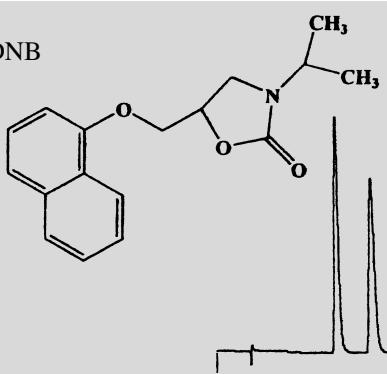
Flow Rate: 1.0 mL/min
Detection: UV 254 nm

Run Time: 11.0 min

$k'_1: 2.27$

$\alpha: 1.42$

reference 59



Bambuterol

Bambuterol

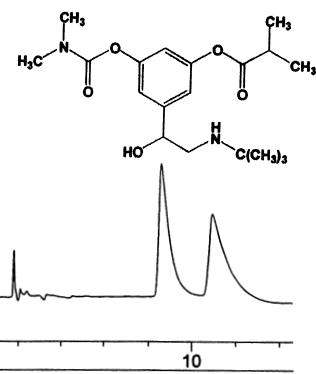
Column = (R,R)-alpha-Burke 2
25 cm x 4.6 mm

Mobile Phase = (40/40/20)
Hexane/Methylene Chloride/Ethanol + 20 mM Ammonium Acetate

Flow Rate = 1.5 mL/min
Detection = UV 254 nm

$k'_1 = 3.74$

$\alpha = 1.35$
reference 46



Alprenolol

Alprenolol

90:5:5 CH₂Cl₂/EtOH/MeOH

10 mM NH₄OAc

1 ml/min; 254 nm

run time = 10 min

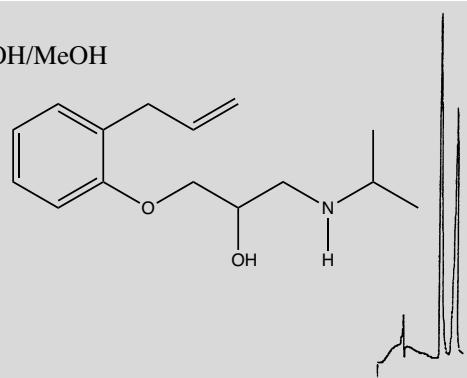
4.6 mm x 25 cm

α -Burke 2

k' ₁ = 1.44

α = 1.44

reference 33



Atenolol

Atenolol

85:10:5

CH₂Cl₂/EtOH/MeOH

15 mM NH₄OAc

1 ml/min; 254 nm

run time = 16 min

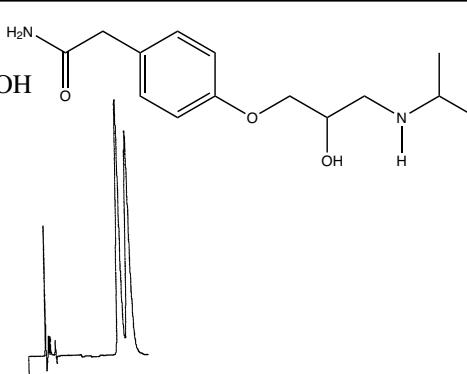
4.6 mm x 25 cm

α -Burke 2

k' ₁ = 4.41

α = 1.13

reference 33



Bufuralol

Bufuralol

Column: (3R,4S)-Pirkle 1-J

25 cm x 4.6 mm

Mobile Phase: (90/10)

CH₂Cl₂/Ethanol

+ 0.02 M Ammonium Acetate

Flow Rate: 1.0 mL/min

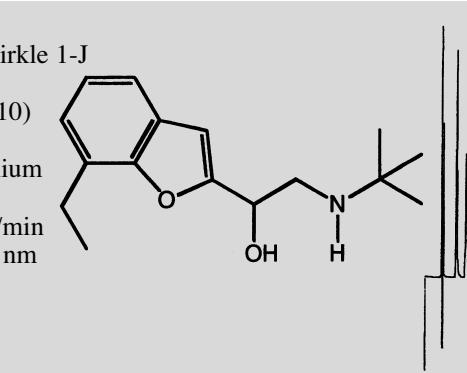
Detection: UV 254 nm

Run Time: 7.0 min

k' ₁: 0.91

α : 2.01

reference 46



Tulobuterol HCl

Tulobuterol HCl

Column: (S)- α -Burke 2

25 cm x 4.6 mm

Mobile Phase: (91/9)

CH₂Cl₂/Ethanol

+ 0.01 M

Ammonium Acetate

Flow Rate: 1.5 mL/min

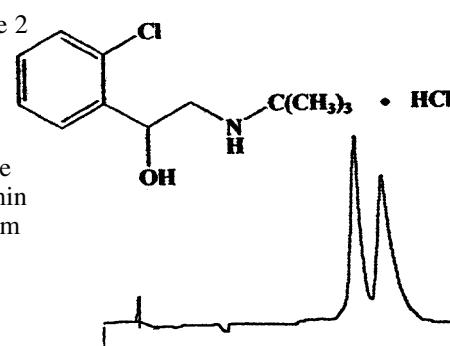
Detection: UV 254 nm

Run Time: 15.0 min

k' ₁: 6.38

α : 1.13

reference 46



Betaxolol

Betaxolol

85:10:5 CH₂Cl₂/EtOH/MeOH

10 mM NH₄OAc

1 ml/min; 254 nm

run time = 11 min

4.6 mm x 25 cm

α -Burke 2

k' ₁ = 2.36

α = 1.25

reference 33



Bupranolol

Bupranolol

Column: (3R,4S)-Pirkle 1-J

25 cm x 4.6 mm

Mobile Phase: (85/15)

CH₂Cl₂/Ethanol
+ 0.015M Ammonium Acetate

Flow Rate: 1.0 mL/min

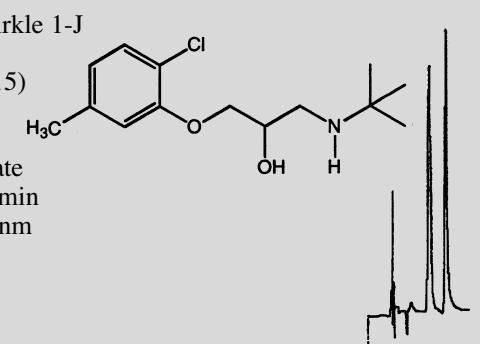
Detection: UV 254 nm

Run Time: 8.5 min

k' ₁: 1.44

α : 1.47

reference 46



Metoprolol

metoprolol

85:10:5 CH₂Cl₂/EtOH/MeOH

10 mM NH₄OAc

1 ml/min; 254 nm

run time = 13 min

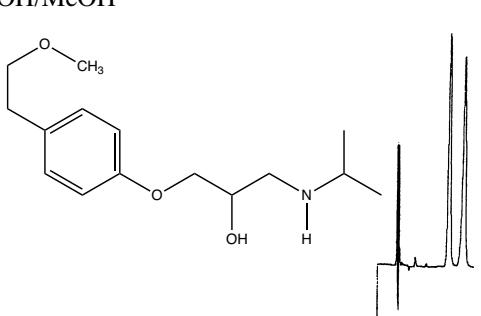
4.6 mm x 25 cm

α -Burke 2

k' ₁ = 2.66

α = 1.28

reference 33



β -Blocker

β -Blocker

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:

(90/10)
CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

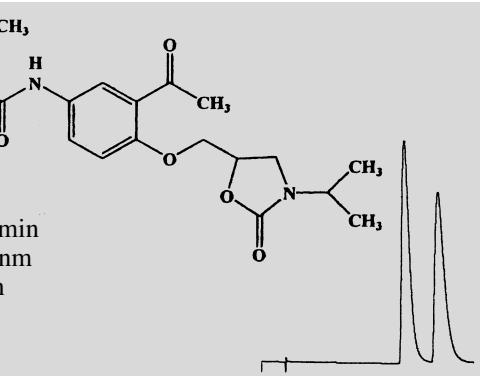
Detection: UV 254 nm

Run Time: 18.0 min

k' ₁: 4.52

α : 1.29

reference 59



REGIS β -Blockers

Nadolol

Nadolol

Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm

Mobile Phase = (78/22)

Hexane/Ethanol +
0.01 M Ammonium
Acetate

Flow Rate = 1.5 mL/min

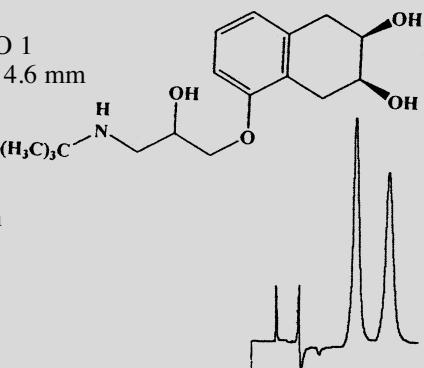
Detection = UV 270 nm

Run Time = 9.5 min

k'_1 = 3.05

α = 1.43

reference 46



Timolol Maleate

Timolol Maleate

Column = (3R,4S)-Pirkle 1-J
25 cm x 4.6 mm

Mobile Phase =
(94/3/3) CH₂Cl₂/
Ethanol/IPA
+ 0.01M Ammonium Acetate

Flow Rate = 1.0 mL/min

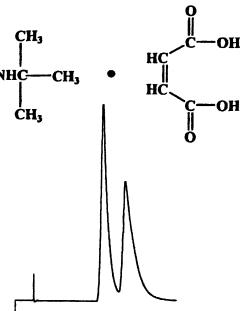
Detection = UV 294 nm

Run Time = 16.0 min

k'_1 = 3.72

α = 1.33

reference 46



Carazolol

Carazolol

Column = (R)- α -Burke 2
25 cm x 4.6 mm

Mobile Phase =

(46/46/8) CH₂Cl₂/
Methanol/Ethanol + 0.01 M
Ammonium Acetate

Flow Rate = 1.5 mL/min

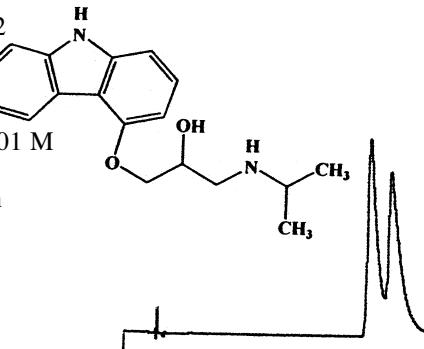
Detection = UV 254 nm

Run Time = 15.0 min

k'_1 = 6.73

α = 1.10

reference 46



Pindolol

Pindolol

Column: (3R,4S)-Pirkle 1-J
25 cm x 4.6 mm

Mobile Phase: (80/20)
CH₂Cl₂/Ethanol + 0.04M
Ammonium Acetate

Flow Rate: 1.0 mL/min

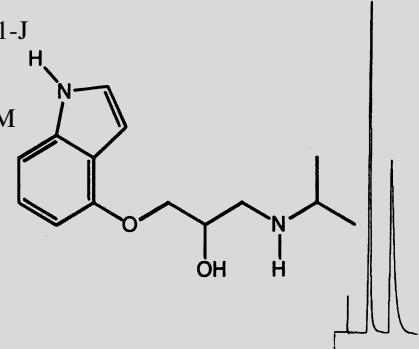
Detection: UV 254 nm

Run Time: 11.0 min

k'_1 : 1.56

α : 2.06

reference 46



Practolol

Practolol

85:10:5 CH₂Cl₂/
EtOH/MeOH

15 mM NH₄OAc

1 ml/min; 254 nm

run time = 19 min

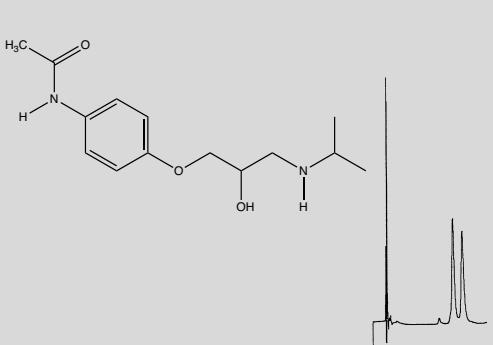
4.6 mm x 25 cm

α -Burke 2

k'_1 = 4.78

α = 1.14

reference 33



Oxprenolol

Oxprenolol

Column: (3R,4S)-Pirkle 1-J
25 cm x 4.6 mm

Mobile Phase: (90/10)

CH₂Cl₂/Ethanol + 0.015M
Ammonium Acetate

Flow Rate: 1.0 mL/min

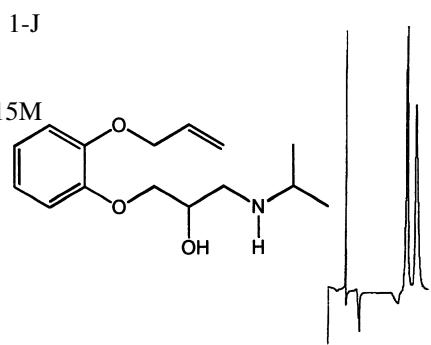
Detection: UV 254 nm

Run Time: 13.5 min

k'_1 : 3.55

α : 1.15

reference 46



Pronethalol

Pronethalol

90:10 CH₂Cl₂/EtOH

15 mM NH₄OAc

1 ml/min; 254 nm

run time = 15 min

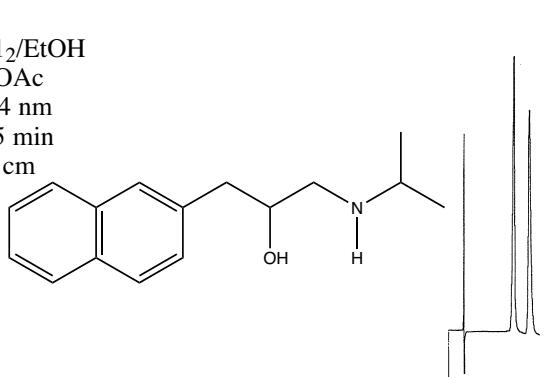
4.6 mm x 25 cm

α -Burke 2

k'_1 = 3.26

α = 1.31

reference 33



Propranolol

Propranolol

Column: (3R,4S)-Pirkle 1-J
25 cm x 4.6 mm

Mobile Phase: (80/20)

CH₂Cl₂/Ethanol + 0.04M
Ammonium Acetate

Flow Rate: 1.0 mL/min

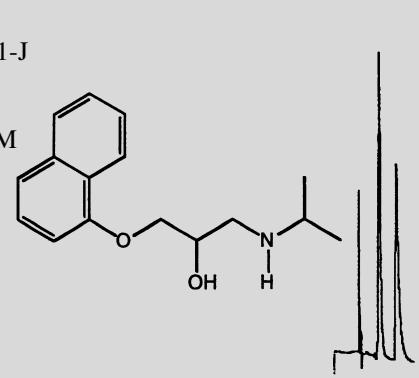
Detection: UV 254 nm

Run Time: 6.5 min

k'_1 : 0.80

α : 1.80

reference 46



Norleucine

Norleucine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O +10 mM Acetic acid

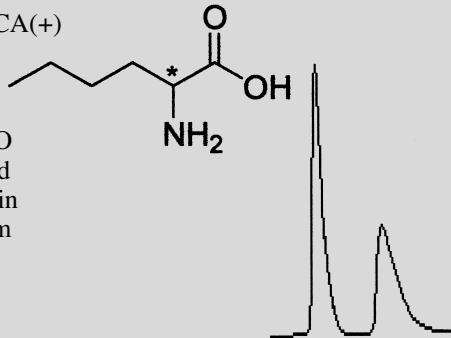
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 5.6 min

k'₁: 1.28

α: 1.75

**1,2,3,4-Tetrahydro-1-naphthylamine**

1,2,3,4-Tetrahydro-1-naphthylamine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (84/16)

CH₃OH/H₂O +10 mM H₂SO₄ and 0.1% TEA

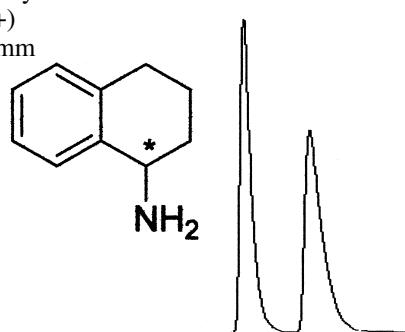
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 3.5 min

k'₁: 0.82

α: 1.76

**1-Aminoindan**

1-Aminoindan

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (84/16)

CH₃OH/H₂O +5 mM HClO₄

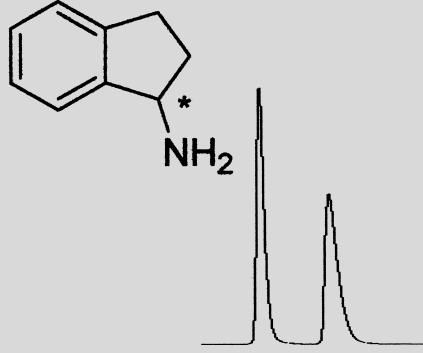
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 4.8 min

k'₁: 1.44

α: 1.91

**Methionine**

Methionine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O +10 mM Acetic acid

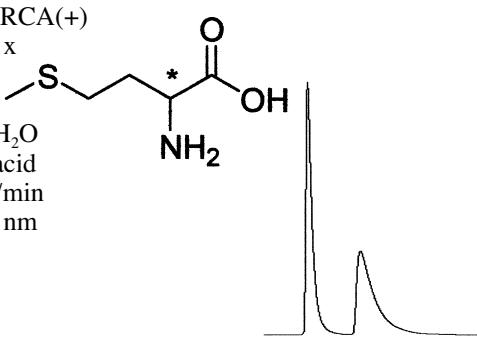
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 7.5 min

k'₁: 1.64

α: 2.04

**4-Fluorophenylalanine**

4-Fluorophenylalanine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (70/30)

CH₃OH/H₂O+10

mM Acetic acid

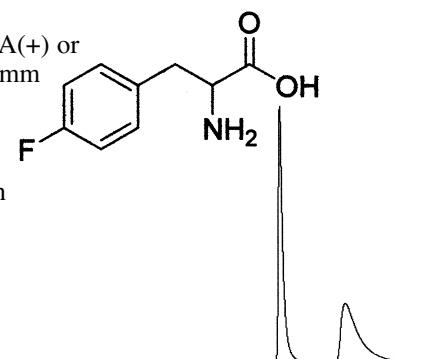
Flow Rate: 1.5 mL/min

Detection: UV 210 nm

Run Time: 9.6 min

k'₁: 2.92

α: 2.56

**Arginine**

Arginine

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(84/16) CH₃OH/H₂O+10 mM H₂SO₄

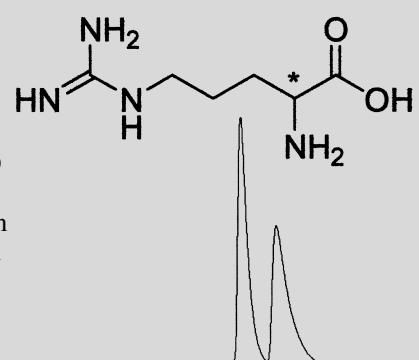
Flow Rate: 0.8 mL/min

Detection: UV 210 nm

Run Time: 4.9 min

k'₁: 1.21

α: 1.64

**Tyrosine**

Tyrosine

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(70/30) CH₃OH/H₂O

+10 mM Acetic acid

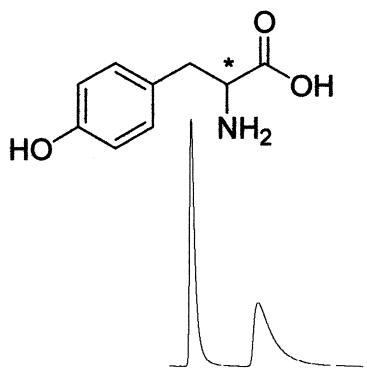
Flow Rate: 1.5 mL/min

Detection: UV 210 nm

Run Time: 9.1 min

k'₁: 2.95

α: 2.38

**Leucine**

Leucine

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O

+10 mM Acetic acid

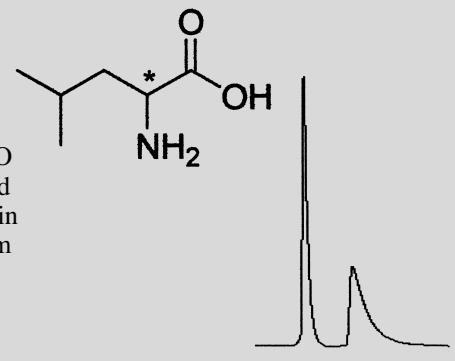
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 5.5 min

k'₁: 1.03

α: 2.14



REGIS Amino Acids

Glutamine

Glutamine

Column: ChiroSil® SCA(-)
25 cm x 4.6 mm

Mobile Phase:
(65/35) CH₃CN/H₂O
+0.01% Acetic acid

Flow Rate: 1.5 mL/min

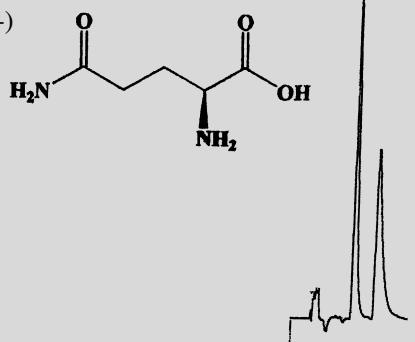
Detection: UV 210 nm

Run Time: 6.5 min

k'_1 : 1.51

α : 1.61

reference 46



Kynurenone

Kynurenone

Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm

Mobile Phase =
(65/35)

H₂O/CH₃OH +

0.1% Acetic Acid

Flow Rate = 1.0 mL/min

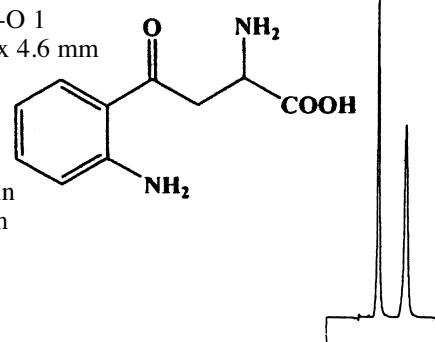
Detection = UV 254 nm

Run Time = 9.0 min

k'_1 = 1.17

α = 1.99

reference 46



D,L-p-Hydroxy-Phenylglycine

D,L-p-Hydroxy-Phenylglycine

Column: ChiroSil®-SCA(+)

15 cm x 4.6 mm

Mobile Phase:

(50/50) CH₃ON/H₂O
+0.02% Acetic acid

Flow Rate: 1.0 mL/min

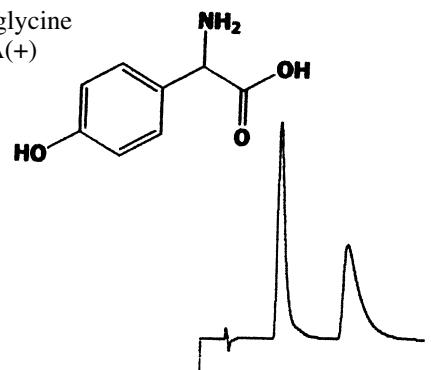
Detection: UV 210 nm

Run Time: 11.0 min

k'_1 : 2.11

α : 2.29

reference 46



Tryptophan

Tryptophan

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (70/30)

CH₃OH/H₂O

+10 mM Acetic acid

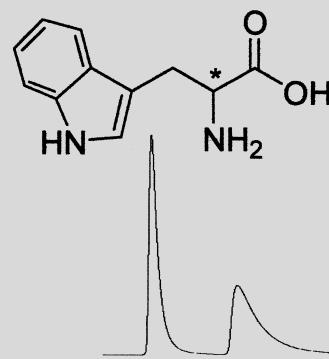
Flow Rate: 1.5 mL/min

Detection: UV 210 nm

Run Time: 11.0 min

k'_1 : 4.06

α : 2.15



Serine

Serine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(84/16) CH₃OH/H₂O
+5 mM HClO₄

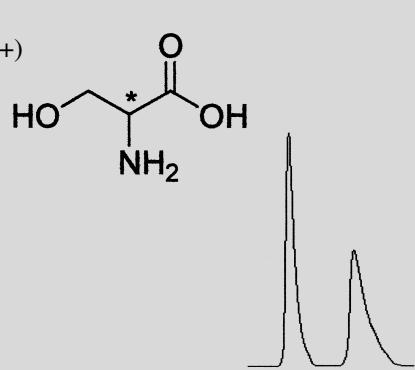
Flow Rate: 0.8 mL/min

Detection: UV 210 nm

Run Time: 6.0 min

k'_1 : 1.37

α : 1.99



Phenylalanine

Phenylalanine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (70/30)

CH₃OH/H₂O

+10 mM Acetic acid

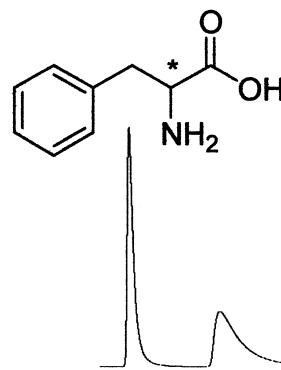
Flow Rate: 1.5 mL/min

Detection: UV 210 nm

Run Time: 8.9 min

k'_1 : 2.66

α : 2.57



Phenylglycine

Phenylglycine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (70/30)

CH₃OH/H₂O
+10 mM H₂SO₄ and 0.1% TEA

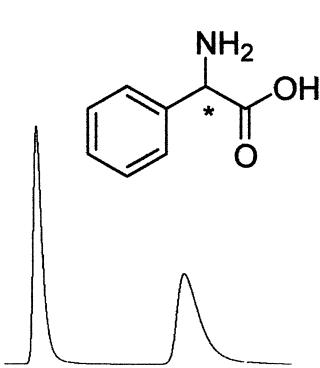
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 13.1 min

k'_1 : 3.14

α : 2.60



Norvaline

Norvaline

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O
+10 mM Acetic acid

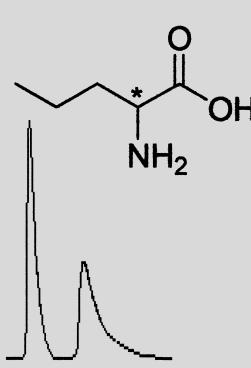
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 5.3 min

k'_1 : 1.15

α : 1.79



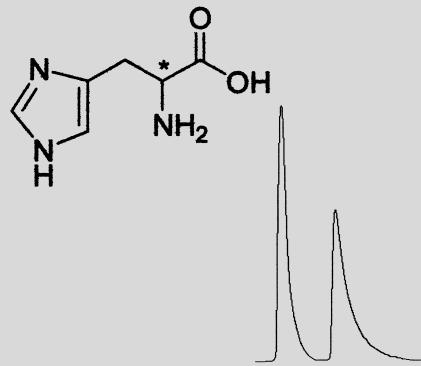
Histidine

Histidine

Column: ChiroSil®

RCA(+) or SCA(-)
15 cm x 4.6 mmMobile Phase:
(45/55) CH₃OH/H₂O
+10 mM Acetic acid
Flow Rate: 1.0 mL/min
Detection: UV 210 nm
Run Time: 26.0 mink'₁: 10.96

α: 1.27

**Glutamic Acid**

Glutamic Acid

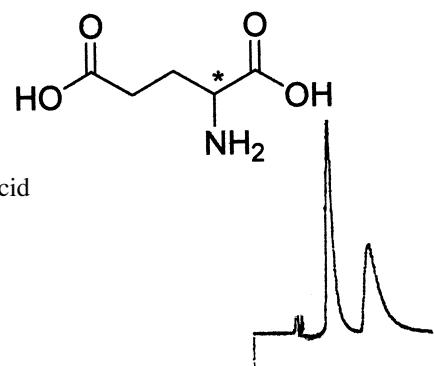
Column: ChiroSil®

RCA(+) or SCA(-)
15 cm x 4.6 mmMobile Phase:
(65/35) CH₃OH/H₂O
+0.05% Phosphoric acid
Flow Rate: 1.0 mL/min
Detection: UV 210 nm

Run Time: 4.5 min

k'₁: 0.71

α: 2.27

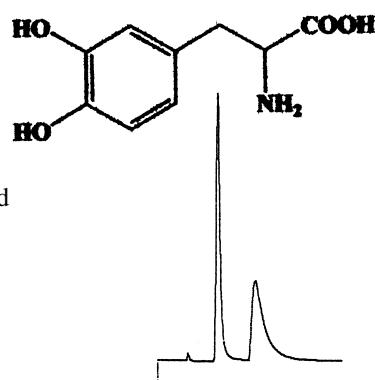
**DOPA**

DOPA

Column: ChiroSil®

RCA(+) or SCA(-)
15 cm x 4.6 mmMobile Phase:
(70/30) CH₃OH/H₂O
+0.01% Phosphoric acid
Flow Rate: 1.0 mL/min
Detection: UV 210 nm
Run Time: 5.5 mink'₁: 0.97

α: 2.30

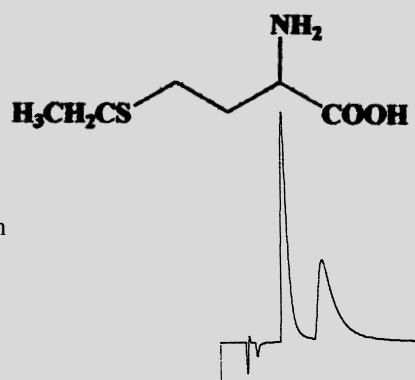
**Ethionine**

Ethionine

Column: ChiroSil®

RCA(+) or SCA(-)
15 cm x 4.6 mmMobile Phase:
(75/25) CH₃OH/H₂O
+0.02% Acetic acid
Flow Rate: 1.0 mL/min
Detection: UV 210 nm
Run Time: 6.2 mink'₁: 1.29

α: 2.07

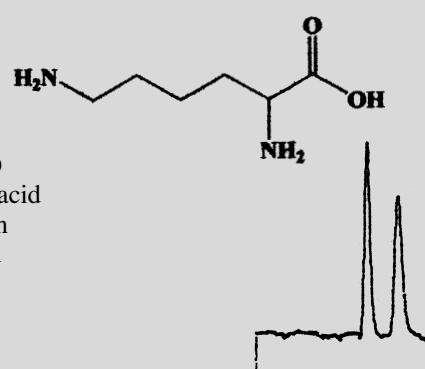
**Lysine**

Lysine

Column: ChiroSil®

RCA(+) or SCA(-)
15 cm x 4.6 mmMobile Phase:
(70/30) CH₃OH/H₂O
+0.01% Phosphoric acid
Flow Rate: 1.0 mL/min
Detection: UV 210 nm
Run Time: 5.3 mink'₁: 1.44

α: 1.48



Frequently Asked Questions

ABOUT PIRKLE-TYPE CHIRAL STATIONARY PHASES AND COLUMNS FROM REGIS

Throughout the past 20 years, the Sales and Technical persons at Regis have fielded hundreds of different questions related to our Chiral Stationary Phases (CSP's). Listed here you will find some of the frequently asked questions. By no means is this a complete list, if you have questions regarding chiral chromatography, please feel free to contact Regis directly or pass your question on through one of our distributors.

What is the pressure rating of your columns?

All analytical (25cm x 4.6 mm i.d.) and semi-preparative (25cm x 10.0 mm i.d.) columns manufactured by Regis can tolerate pressures up to 6000 psi. Larger columns will tolerate 3000 psi. It is very important not to exceed the maximum pressure rating for any HPLC column as you may disrupt the integrity of the silica bed and destroy the column.

Can Regis columns be reversed?

Yes, all columns packed by Regis are fully reversible. In fact, Regis was the first column manufacturer to sell a fully reversible HPLC column. It is recommended to reverse your column frequently. This helps keep the frit surface from becoming clogged with undissolved sample or particulates in the mobile phase, thus extending the column life.

What is the pH range of your columns?

All of Regis' Chiral phases are bonded on silica. The recommended pH range is 2.5 to 7.5. Limited usage outside of this pH range can be tolerated, but it has been proven that extended usage outside of the range will decrease column life.

Can your columns be used in normal and reversed-phase solvents?

Yes, all Pirkle-Type Chiral HPLC columns can be used in *BOTH* normal and reversed-phase solvents. Generally, the Pirkle-Type CSP's will give better separations by using them with normal-phase solvents. There are numerous examples, however; where separations with reversed-phase solvents will outperform those with normal-phase solvents.

Can I use the same column for reversed-phase and normal-phase solvent systems while doing method development?

Yes, just make sure you completely flush out the column with a miscible solvent such as IPA or ethanol. We recommend at least 20 column volumes.

How long does it take your columns to equilibrate?

The column should equilibrate after about 20 column volumes. When you are switching from normal to reversed-phase solvent systems and vice-versa, flush the column with a miscible solvent for 20 column volumes. It should take another 20 column volumes to equilibrate. The equilibration volumes may vary depending on the composition of the mobile phase.

What type of silica do you use?

Regis exclusively uses Exsil® for our 5-micron material and Kromasil, for 10 and 16-micron material. Both brands of silica are 100 angstrom in pore size.

Do you always need a modifier in the mobile phase?

No modifiers are usually needed for initial method development. Modifiers can be used to improve peak shape and resolution when the samples are extremely basic or acidic in nature. Acetic acid or ammonium acetate are recommended for acidic compounds, and triethylamine, diethylamine or ammonium acetate are recommended for basic compounds. Usually 0.1% of modifier is all that is required. Note: Although TFA may be used as a modifier, its use should be limited. Acetic acid usually works as well as TFA.

Can I use your columns for SMB chromatography and SFC?

Yes, many analytical and preparative chromatographers use Pirkle-Type Chiral columns in both SFC and SMB. Special hardware is necessary for certain column dimensions.

What is the difference between Whelk-O 1 and Whelk-O 2?

Although the Whelk-O 1 and Whelk-O 2 both share the same Chiral selector, they have distinct differences. The Whelk-O 1 is monofunctionally bonded to silica and the Whelk-O 2 is trifunctionally bonded. The Whelk-O 2 was designed to tolerate strong acidic modifiers such as TFA. The Whelk-O 2 was designed for preparative use and is not available on 5-micron silica. Due to the fact the Whelk-O 2 is a trifunctional bond, coverage on the silica will be less than with Whelk-O 1. This decrease in the actual number of bonded sites will decrease selectivity and not allow for exact reproducibility of a method developed on a Whelk-O 1 column.

Does my compound need an aromatic ring to achieve separation on a Pirkle-Type Chiral column?

In most cases (not all), yes. Chiral recognition occurs at binding sites. The potential $\pi-\pi$ interaction that can occur between the aromatic rings on the Chiral selector and the aromatic ring on the sample is a major factor in achieving selectivity. Binding does occur at other sites such as acidic sites, basic sites and steric interaction sites—this is why you do not always need a ring—but by far, the $\pi-\pi$ interaction is the major binding site.

Can I use the Pirkle-Type Chiral columns in polar organic mode?

Yes, even though the success rate is very poor, you can use the columns in polar organic mode. We do not recommend dedicating a slot in your method development station for a Pirkle-Type Chiral column if you are exclusively running in polar organic mode. Add another Pirkle-Type column to your normal-phase system to achieve a higher success rate.

What sampling loading can I expect from Pirkle-Type Chiral HPLC columns?

The typical loading range – with relative retention's (α) greater than 1.3—is ~ 4-16 mg of sample per gram of packing. Below are typical loadings for some of the different column sizes: Note: Factors, such as solubility, will greatly affect loading capacity.

Analytical column, 25cm x 4.6mm, ~ 3.5 grams of packing, loading is 14-56 mg/ injection.

Semi-prep column, 25cm x 10.0mm, ~ 16 grams of packing, loading is 64-256 mg/ injection.

Prep column, 25cm x 21.1mm, ~ 72.5 grams of packing, loading is 288-1,152 mg/ injection.

Quick Scheme Method Development

FOR THE REGIS PIRKLE-TYPE CHIRAL STATIONARY PHASES

STEP 1:

Choosing the Appropriate Column:

We recommend using the following sequence of columns to start your method development. When doing method development at Regis, the Whelk-O is our first choice as it exhibits the broadest degree of generality.

Order of Preference:

- Whelk-O
- ULMO
- DACH-DNB
- α -Burke
- β -Gem
- Pirkle 1-J
- Leucine
- Phenylglycine

STEP 2:

Choosing the Mobile Phase:

Certain factors such as solubility and future considerations for preparative work usually help to determine whether to perform your method development with reversed-phase or normal phase solvents. Pirkle-Type phases can be used in both modes, but typically performs the best with normal phase solvents. Since the majority of analytical Chiral methods move on to preparative separations, we recommend using normal phase solvents.

Typical Mobile Phase Combinations:

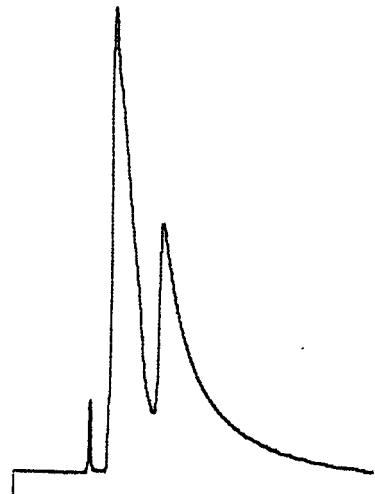
- Hexane/IPA
- Hexane/Ethanol
- Hexane/ CH_2Cl_2 /Ethanol
- Hexane/Ethyl Acetate
- Heptane/Ethanol
- Heptane/ CH_2Cl_2
- Hexane/ CH_2Cl_2
- Heptane/IPA
- Methanol/ H_2O
- Ethanol/ H_2O
- Acetonitrile/ H_2O
- Methanol/ CH_2Cl_2
- THF/ H_2O
- Ethanol/ CH_2Cl_2

STEP 3:

Start with a high percentage (~50%) of strong solvent (ethanol, IPA, ect.):

Starting with a strong solvent system ensures that all peaks will elute off the column quickly.

Sample: Naproxen
Column: (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (50/50)
Hexane/Ethanol
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 6.5 min



k' : 1.37
 α : 1.87
 R_s : 1.59

- If you achieve any resolution, (such as the above sample) move on to STEP 4.
- If your sample comes off in the void, decrease the strong solvent by half.
- If your sample is now out of the void and you have resolution, move on to STEP 4.
- If your sample is out of the void, and there is no resolution, change the column.

Steps 4 and 5 continued on pages 84 & 85

Quick Scheme Method Development

FOR THE REGIS PIRKLE-TYPE CHIRAL STATIONARY PHASES

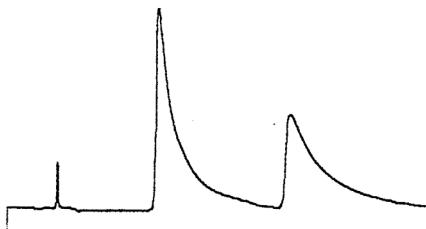
STEP 4:

Add a Mobile Phase Modifier (usually ~ 0.1%)

As you can see, the peak shape of the initial separation is very poor. To rectify this problem, a modifier is usually added. If you are satisfied with the peak shape—you do not need to add a modifier—move on to STEP 5 and optimize your separation.

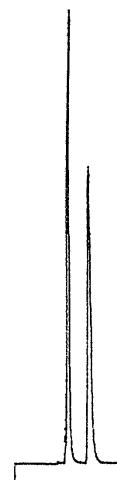
- For basic or amine groups—add triethylamine, diethylamine or ammonium acetate
- For acidic groups—add acetic acid, trifluoroacetic acid or ammonium acetate

Sample: Naproxen
Column: (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (50/50)
Hexane/Ethanol
+ 0.1% TEA
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 19.0 min



k'_1 : 4.63
 α : 2.07
 R_s : 4.14

Sample: Naproxen
Column: (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (50/50)
Hexane/Ethanol
+ 0.1% Acetic Acid
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 4.7 min



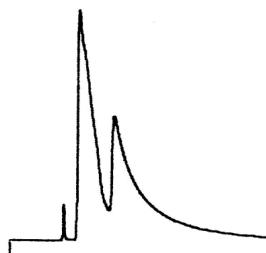
k'_1 : 0.87
 α : 1.85
 R_s : 7.24

- Although resolution increased with the addition of 0.1% of triethylamine to the mobile phase, the peak shape is still very poor.
- Try adding a different modifier

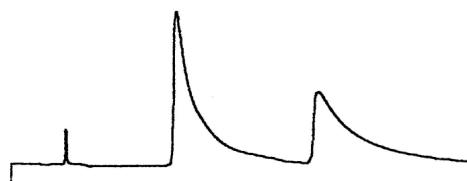
- Resolution and peak shape are excellent with the addition of 0.1% of acetic acid

Recapping The First Four Steps:

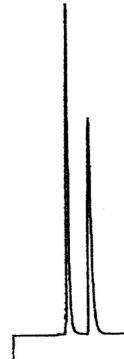
- For this sample, you can stop at 50/50 Hexane/Ethanol + 0.1% acetic acid if you are only looking for a basic method or you can carry it forward to STEP 5 and optimize.



50/50
Hexane/Ethanol
 k'_1 : 1.37
 α : 1.87
 R_s : 1.59



50/50
Hexane/Ethanol
+ 0.1% TEA
 k'_1 : 4.63
 α : 2.07
 R_s : 4.14



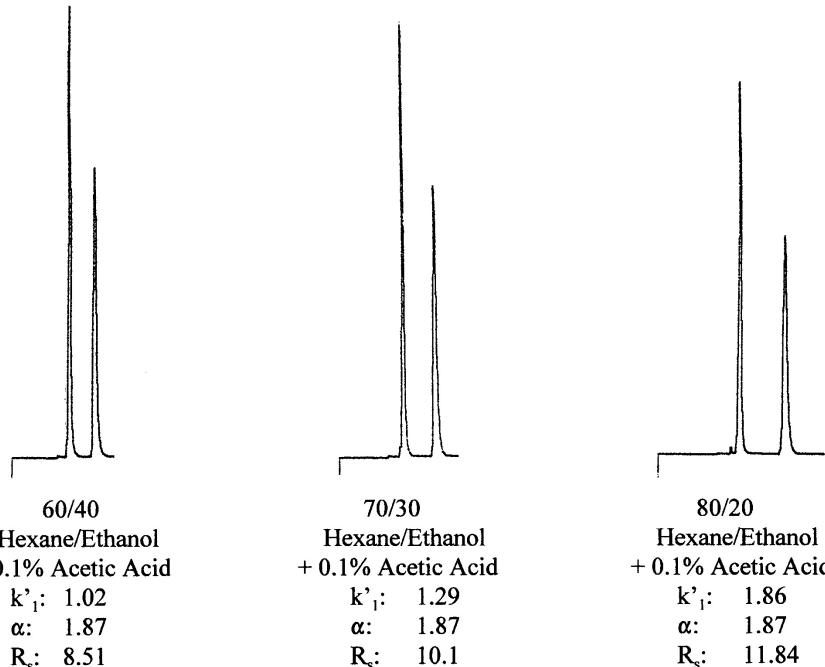
50/50
Hexane/Ethanol
+ 0.1% Acetic Acid
 k'_1 : 0.87
 α : 1.85
 R_s : 7.24

STEP 5:**Optimizing your method:**

Optimizing a Chiral method is very similar to optimizing an achiral method. Changing mobile phase component concentrations and even the components themselves can dramatically change resolution.

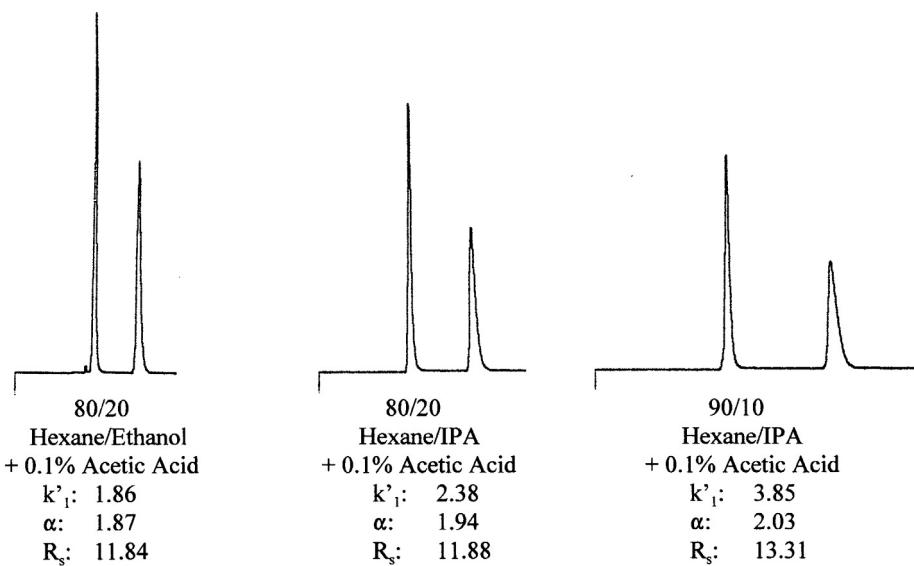
Increase the concentration of the weaker solvent:

- Increasing the hexane content increased the resolution in this example. Again, you can stop here and accept this as optimized or continue on.

**Change the strong solvent:**

- By substituting IPA for ethanol, an increase in both resolution and alpha were achieved.

Optimization of a Chiral method can be as simple or as complicated as you want it to be. Different mobile phase components can be used; modifiers can be changed or eliminated; you can switch to reversed-phase solvents; you can change columns. The possibilities are endless. We suggest you keep it as simple as possible. Once you have achieved an acceptable separation, move on to the next project. Small increases in resolution and alpha are usually not worth the time spent in method development to achieve those increases.



Not sure which chiral column to use for your separation?



sales@registech.com

Let the professional staff at Regis assist you with its free chiral screening service.

Simply fill out the chiral screening data sheet (see page 91 of this Guide) and pre-fax it to Regis or send it along with your sample of interest.

STRICTLY CONFIDENTIAL • FAST TURN AROUND TIME • NO OBLIGATION TO THE CUSTOMER

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Chiral Screening Data Sheet

Contact Information:

Primary Contact _____

Other Contact _____

Company Address _____

City _____

State _____ Zip Code _____

Country _____

Phone _____

Fax _____

e-mail _____

Compound Structure/Name

This compound may or may not
be used in an application booklet.

Separation Requirements:

Analytical

Preparative

Quantity _____ mg gr kg

Do you want your sample returned?

Yes No

Please Note: All samples are destroyed after the screening process is complete.

Physical, Chemical, and Chromatographic Data:

MSDS Available Yes No If yes, include a copy with your sample.

Hazardous Material Yes No Unknown

Special Handling Requirements Yes No Unknown

Appearance:

Powder Crystal Oil Other

Color _____ pKa _____ UV (max) _____ UV (min) _____

Chemical Purity _____

Chiral Screening Data Sheet

Stability/Exposure:

Light	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Moisture	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Temp<40°C	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown



ACIDS:

Acetic Acid (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Trifluoroacetic Acid (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

BASES:

Triethylamine (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Diethylamine (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

Storage Conditions: _____

Solubility

Water	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Methanol	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Ethanol	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
2-Propanol	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Hexane	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Ethyl Acetate	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
CH ₂ Cl ₂	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Acetonitrile	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

Other _____

Chromatographic Analysis:

Column _____

Column Manufacturer _____

Mobile Phase _____

Flow Rate _____ Wavelength _____

Please include a copy of the chromatogram.

Instructions:

Please send the completed form along with your sample. We would like to have at least 25 mg of sample. If you are unable to send us 25 mg, you must include sufficient solubility information. If the compound you are sending is not commercially available, please inquire if you need a confidentiality agreement signed before you send us your sample.



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